Targeted cytotoxic bombesin analog AN-215 effectively inhibits experimental human breast cancers with a low induction of multi-drug resistance proteins

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Abstract

The cytotoxic analog of bombesin (BN)/gastrin releasing peptide (GRP) AN-215 consisting of 2-pyrrolinodoxorubicin (AN-201), a superactive derivative of doxorubicin linked to a bombesin analog carrier, displays a high affinity to BN/GRP receptors and can be targeted to tumors that express these receptors. We evaluated the antitumor effect and the toxicity of AN-215 in 5 human breast cancer cell lines xenografted into nude mice. In addition, we measured the mRNA expression of multi drug resistance protein 1 (MDR-1), multi drug resistance related protein 1 (MRP-1) and breast cancer resistance protein (BCRP) by real-time PCR analysis after treatment with AN-215. All five cell lines expressed BN/GRP receptors, and AN-215 significantly \( P < 0.05 \) inhibited tumor growth in all models, while its cytotoxic radical AN-201 had no significant effect in four models. In MX-1 tumors, AN-201 had a significantly weaker antitumor effect than AN-215. The effect of AN-215 was nullified by a blockade of BN/GRP receptors with a bombesin antagonist. Low or no induction of MDR-1, MRP-1 and BCRP occurred after treatment with AN-215. In conclusion, targeted chemotherapy with the cytotoxic BN/GRP analog AN-215 strongly inhibits breast cancers that express BN/GRP receptors and might provide a new treatment modality for mammary carcinoma.

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Introduction

Breast cancer is the most common malignancy among women in the United States and Europe and ranks second as a cause of cancer-related deaths (Bray et al. 1995, Hortobagyi 2003, Jemal et al. 2005). Thus, in 2005, an estimated 210,000 new cases of breast cancer with about 40,000 deaths are expected to occur in the USA (Jemal et al. 2005). Most patients succumb to this disease not because of the primary cancer, but because of metastases. In spite of the use of endocrine therapy, systemic chemotherapy and novel approaches such as trastuzumab (Herceptin), an antibody against human epidermal growth factor receptor 2 (HER-2), metastatic disease remains generally incurable with a median survival time of 2 to 3 years (Powles et al. 1980, Todd et al. 1983, Clark et al. 1987, Greenberg et al. 1996, Osborne 1998). Therefore, it is mandatory to develop novel, more effective treatment strategies with low toxicity for the treatment of breast cancer.

The discovery of specific molecular characteristics of malignant cells prompted the development of a new class of drugs known as targeted therapeutics. This new class of antitumor agents includes antibodies against surface structures on tumor cells, such as trastuzumab (Baselga et al. 1996), and hybrid molecules consisting of receptor-specific ligands linked to toxins, radionuclides or chemotherapeutic agents (Magrath 1994, Abou-Jawde et al. 2003, Trail et al. 2003). High-affinity receptors for peptide hormones
expressed on malignant cells can be targeted by cytotoxic hormone analogs consisting of a peptide carrier linked to an antineoplastic agent (Nagy et al. 1996, Schally & Nagy 1999, 2003, 2004). The direct delivery of the cytotoxic radical to the malignant tissue is expected to result in a higher antitumor effect with a reduced systemic toxicity and may overcome chemoresistance of cancer cells. Multidrug resistance (MDR) of cancer cells to a variety of antitumor agents remains a major problem in chemotherapy. The ATP-dependent membrane transporter protein, p-glycoprotein (pGP), the product of the MDR-1 gene, and the recently discovered MDR protein 1 (MRP-1) mediate the increased extracellular efflux of chemotherapeutic agents (Juliano & Ling 1976, Cole et al. 1992). Breast cancer resistance protein (BCRP) is another overlapping but distinct type of MDR, based on drug efflux (Ross et al. 1999). The expression of mRNA for MDR-1 has been detected in about 50% of untreated breast cancer samples (Keith et al. 1990) and induction of the MDR-1 protein pGP was detected in breast cancer specimens after treatment with doxorubicin (Schneider et al. 1989). MRP-1 immunoreactivity was found in 34–80% of primary breast carcinomas (Noote et al. 1997, Filipits et al. 1999). MRP-1 overexpression seems to be associated with a poor prognosis (Noote et al. 1997, Ito et al. 1998, Filipits et al. 1999). Chemotherapeutic drugs targeted directly to tumor cells could overcome MDR based on transmembrane efflux, as the local concentration of the antitumor agent is increased.

Specific receptors for bombesin (BN)-like peptides, such as the gastrin-releasing peptide receptor (GRPR), the neuromedin B receptor (NMBR) and the bombesin receptor subtype 3 (BRS-3), have been found in various human cancer specimens and cancer cell lines (Halmos et al. 1995, Wang et al. 1996, Markwalder & Reubi 1999, Sun et al. 2000, Schally et al. 2001). The presence of binding sites for BN/GRP in human breast cancer specimens was established by our group (Halmos et al. 1995, 1996) and it has recently been reported that the GRPR, which binds bombesin with high affinity, was expressed in 29 of 46 human invasive ductal breast carcinomas (Gugger & Reubi 1999). In addition, the lymph node metastases from GRPR-positive tumors were all found to be GRPR positive, whereas in surrounding lymphoreticular tissue no receptor expression could be found (Gugger & Reubi 1999). In several clinical studies, primary breast cancers and their lymph node metastases were successfully detected using GRPR scintigraphy (Van de Wiele et al. 2000, Scopinaro et al. 2002, Soluri et al. 2003). Therefore, GRPR scintigraphy could be used to select patients who may respond to therapy targeted to BN/GRP receptors. These findings suggest that BN/GRP receptors on breast cancer are a potential target for therapy with cytotoxic bombesin analogs.


This study was designed to evaluate the therapeutic efficacy and the toxicity of the cytotoxic bombesin analog AN-215 and its effect on MDR-1, MRP-1 and BCRP expression in experimental models of human breast cancer in vivo.

Materials and methods

Peptides and cytotoxic radical

The cytotoxic radical 2-pyrrolino-DOX (AN-201), the bombesin analog carrier RC-3094, the cytotoxic bombesin analog AN-215 and the bombesin antagonist RC-3095 were synthesized in our laboratory as described (Nagy et al. 1996, 1997). Before intravenous (i.v.) injection, the compounds were dissolved in 5% (w/v) aqueous D-mannitol solution (Sigma, St Louis, MO, USA).

Cell lines

cell line DU-4475 originated from a cutaneous metastatic nodule of a 70-year-old Caucasian female (Langlois et al. 1979). All cell lines were obtained from the American Type Culture Collection (ATCC, Manassas, VA, USA).

The cells were grown at 37°C in a humidified 95% air:5% carbon dioxide atmosphere, passaged weekly and routinely monitored for mycoplasma contamination using a detection kit (Boehringer Mannheim, Mannheim, Germany). All culture media were purchased from Gibco (Grand Island, NY, USA).

The hormone-independent DOX-resistant human breast cancer xenograft MX-1, originating from a surgical explant, was kindly donated by Dr Richard Camalier (NCI, Frederick Cancer and Development Center, Frederick, MD, USA). Tumors were maintained in donor animals.

Animals

Five- to six-week-old female athymic nude mice (Ncr nu/nu) were obtained from the National Cancer Institute (NCI, Bethesda, MD, USA). The animals were housed in sterile cages under laminar flow hoods in a temperature-controlled room with a 12-h light/12-h darkness schedule. They were fed autoclaved chow and water, which were available ad libitum.

Experimental protocol

Cells of each cell line, except MX-1 and MDA-MB-435 human breast cancers, growing exponentially were implanted into 5 female nude mice by subcutaneous injection of 10^7 cells into both flanks. Tumors resulting after 4 weeks in donor animals were aseptically dissected and mechanically minced. In all experiments except experiment 3, pieces of tumor tissue, about 3mm^3, were transplanted subcutaneously (s.c.) into the experimental animals by a trocar needle. In experiment 3, MDA-MB-435 cells (2×10^6) were directly injected into the mammary fat pad of experimental animals.

Tumor volume (length × width × height × 0.5236) and body weight were measured weekly. The total leukocyte count (WBC) was determined with the Unopette microcollection kit (Becton Dickinson, Franklin Lakes, NJ, USA). The tumor doubling time (TDT) was calculated according to the following formula: (days of treatment × LOG10(2))/(LOG10 (final tumor volume)-LOG10(initial tumor volume)). At the end of each experiment, mice were killed under anesthesia, tumors were excised and weighed, and necropsy was performed. Tumor specimens were snap frozen and stored at −70°C. All experiments were performed in accordance with the institutional ethical guidelines for the care of animals and were essentially in agreement with UKCCCR guidelines (1998) for the welfare of animals in experimental neoplasia. Tumor weights never exceeded 10% of the animals’ body weight. The Institutional Animal Care and Use Committee reviewed the protocol for the animal experiments and gave full approval.

Single dose treatment

Animals bearing MDA-MB-231 (experiment 1), DU-4475 (experiment 2), MDA-MB-435 (experiment 3) or MX-1 (experiment 4) tumors received the following treatment as a single injection into the jugular vein: group 1 (control) was given vehicle solution (5% mannitol), group 2 received the cytotoxic analog AN-215 at 200 nmol/kg, and group 3 was treated with the cytotoxic radical AN-201 at 200 nmol/kg.

In experiment 1, three additional groups were included: group 4 was given an unconjugated mixture of the cytotoxic radical AN-201 and the bombesin analog carrier RC-3094 at 200 nmol/kg, group 5 was injected with RC-3094 alone at 200 nmol/kg, and group 6 received 200 µg of the bombesin antagonist RC-3095 i.v. 15 min prior to the i.v. injection of cytotoxic analog AN-215 at 200 nmol/kg.

Multiple dose treatment

Animals with MDA-MB-231 (experiment 5) tumors were treated twice with i.v. injections (day 1 and day 8). Group 1 received control, vehicle solution, group 2 was given AN-215 at 150 nmol/kg, and group 3 was injected with AN-201 at 150 nmol/kg.

Mice with MDA-MB-468 (experiment 6) tumors were also injected twice i.v. (day 1 and day 29). Group 1 received control, vehicle solution, group 2 was given AN-215 at 200 nmol/kg, and group 3 was injected with AN-201 at 200 nmol/kg.

Receptor binding assay

Binding characteristics of bombesin/GRPR on tumor membrane preparations from control animals and animals after treatment with AN-215 and AN-201 were determined by ligand competition assay using ^125I-labeled [Tyr^4]-bombesin. Preparation of tumor membrane fractions and receptor binding studies of BN/GRP were performed as described previously (Halmos et al. 1994).
RNA isolation and reverse transcriptase–polymerase chain reaction (RT-PCR) analysis

The methods for isolation of total RNA and RT-PCR for analysis of the expression of subtypes of BN/GRP receptors (GRPR, NMBR and BR5-3) have been reported (Kiaris et al. 1999). Ten microliter aliquots of the PCR products were separated on 1.8% agarose gel and visualized by ethidium bromide on an ultraviolet transilluminator. The MDA-MB-468 cell line is known to express all three bombesin receptor subtypes and was therefore taken as a positive control (Kahan et al. 2000).

Real-time PCR for MDR-1, MRP-1 and BCRP mRNA expression

Total RNA was isolated from approximately 100 mg tumor tissue for each sample according to the TRI-Reagent protocol (sigma). One microgram total RNA was subjected to reverse transcription with the Iscript Reagent protocol (sigma). One microgram total RNA was subjected to reverse transcription with the Iscript cDNA synthesis kit (BioRad) following the manufacturer’s protocol. Real-time PCR was employed to measure drug resistance gene expression using the SYBR Green system (BioRad). Primers for MDR-1 were designed to measure drug resistance gene expression using the SYBR Green system (BioRad). Primers for MDR-1 (sense 5′-TCT GGA GGA AGA CAT GAC CAG GTA-3′, antisense 5′-GGC ACC AAA ATG AAA CC T GAA TGT-3′), MRP-1 (sense 5′-AGA GAC AGCT CA GCA GCT CCT-3′, antisense 5′-GCC TTG TCA G CCTCC ATG-3′), BCRP (sense 5′-TAT CAA TG G GAT CAT GAA ACC GTG-3′, antisense 5′-GGC GGG GT CGT CTC ATG GAA C-3′) and β-actin (sense 5′-CTG GAA CGT TGA AGG TGA CA-3′, antisense 5′-AAG GGA CTT CCT GTA ACA ATG CA-3′) were used to measure gene expression. The thermal cycling conditions comprised an initial denaturation step at 95 °C for 3 min, then 40 cycles of two-step PCR including 95 °C for 15 s and 60 °C for 1 min. Data were collected during the 60 °C annealing step and were further analyzed by the iCycler iQ Optical system software (BioRad). Real-time PCR efficiencies (E) for MDR-1 (target gene 1), MRP (target gene 2) and β-actin (reference gene) were calculated from the given slopes in the iCycler software according to the following equation: $E = 10^{(-1/slope)}$ (Rasmussen 2001). Nine tumor samples from each experiment (3 control, 3 AN-215 and 3 AN-201) were analyzed in triplicate. For the mathematical model used in this study it was necessary to determine the crossing points for the transcripts of each sample. Crossing points (CPs) are defined as the number of cycles at which the fluorescence rises appreciably above the background fluorescence. Using CP deviations (ACP) for control and treatment (CPcontrol−CPtreatment) of target and reference gene transcripts, quantification of the target genes in treated groups relative to the controls was performed using a mathematical model by Pfaffl (2001):

$$\text{Ratio} = \frac{(E_{\text{target}})^{\text{ACP target (control-treatment)}}}{(E_{\text{reference}})^{\text{ACP reference (control-treatment)}}}$$

Statistical analysis

Data are expressed as means ± s.e. Differences between mean values were evaluated by two-tailed Student’s t-test. $P < 0.05$ was considered significant.

Results

Anti-tumor effects of the cytotoxic bombesin analog AN-215 in single dose application

In experiments 1, 2 and 3, a single dose of AN-215 at 200 nmol/kg significantly inhibited ($P < 0.05$) the growth of MDA-MB-231, DU-4475 and MDA-MB-435 cancers, as reflected by significant decreases in tumor volume and weight and significantly extended tumor doubling times ($P < 0.05$) (Fig. 1A–C, Table 1). Equimolar doses of AN-201 had no significant effects on the growth characteristics. In experiment 1, a mixture of AN-201 and its bombesin analog carrier, RC-3094, and RC-3094 alone did not suppress tumor growth (Fig. 1A, Table 1). In this experiment, the effect of AN-215 could be nullified by injecting 200 μg of the bombesin antagonist RC-3095 15 min prior to the administration of AN-215. In experiment 4, the cytotoxic analog AN-215 and the cytotoxic radical AN-201 at doses of 200 nmol/kg significantly ($P < 0.01$) inhibited the growth of MX-1 tumors for the first 28 days, while the tumors in the control group grew rapidly (Fig. 1D). Thus, the control animals had to be killed on day 29 because of a high tumor burden ($1695.3 ± 246.1 \text{mm}^3$). After treatment with AN-215, 8 of 10 animals became tumor-free, of which 5 redeveloped palpable tumors but 3 remained tumor-free until the end of the experiment (Fig. 1E). In the group treated with AN-201, the tumors of only 2 mice disappeared, but regrew in one animal before the experiment was terminated (Fig. 1E). At the end of the experiment on day 50, animals injected with AN-215 had a significantly ($P < 0.05$) smaller mean tumor volume ($493.8 ± 188.3 \text{mm}^3$) compared with $1261.7 ± 258.0 \text{mm}^3$ than animals treated with an equimolar dose of AN-201 (Fig. 1D).

Antitumor effects of the cytotoxic bombesin analog AN-215 at multiple doses

In experiment 5, administration of AN-215 at a dose of 150 nmol/kg on day 1 and day 8 significantly ($P < 0.05$)
inhibited the growth of MDA-MB-231 tumors from day 8 until the end of the experiment, when the final tumor volume and weight were 55% smaller than in controls. Tumor doubling time was also significantly ($P < 0.05$) prolonged by AN-215. No significant effect on tumor growth was observed in animals treated with equimolar doses of AN-201 (Fig. 2A, Table 1). In experiment 6, consecutive administration of AN-215 at a dose of 200 nmol/kg on day 1 and day 29 significantly ($P < 0.05$) inhibited the growth of xenografts of the relatively slow-growing MDA-MB-468 cancer cell line. The final tumor volume was 71.5% smaller than in controls, and the tumor doubling time was significantly ($P < 0.05$) prolonged by AN-215. The final tumor
weight was inhibited by 68.4% (P < 0.05). No significant effect on tumor growth was observed in animals treated with equimolar doses of AN-201 (Fig. 2B, Table 1).

Side effects and toxicity
AN-215 did not significantly lower the WBC in any experiment (Table 1). In all experiments, WBC was significantly (P < 0.05) decreased on day 8 by AN-201 at a dose of 200 or 150 nmol/kg (Table 1). In experiment 5, a second dose of AN-201 administered on day 8 significantly (P < 0.01) suppressed the WBC on day 15. In experiment 6, a second dose of AN-201 at 200 nmol/kg administered on day 29 significantly decreased the WBC on day 36. In all experiments, a moderate weight loss occurred 8 days after the injection of AN-215 or AN-201, but weights normalized one week later. In experiments 2 and 3, AN-201 significantly (P < 0.05) lowered the body weight on day 8. AN-215 did not cause any statistically significant loss of body weight in any experiment. The overall mortality was 7% in control animals, 6.8% after treatment with AN-215 and 15.8% in animals treated with AN-201.

In vivo expression and characteristics of BN/GRP receptors
Using gene-specific primers, the expression of mRNA for 3 established subtypes of BN receptors was investigated by RT-PCR in MDA-MB-231, MDA-MB-435, DU-4475 and MX-1 tumors (Fig. 3). mRNAs were detected for all three receptor subtypes in all cell lines investigated. MDA-MB-468 is known to express all 3 receptor subtypes and was therefore used as a positive control (Kahan et al. 2000).

Specific high-affinity binding sites for 125I-labeled [Tyr4]-BN were detected on membrane fractions of MDA-MB-231, MDA-MB-435, DU-4475 and MDA-MB-468 breast cancers (Table 2). AN-215 administered as one dose of 200 nmol/kg or as two consecutive doses of 200 and 150 nmol/kg did not affect the binding characteristics of BN/GRP receptors in DU-4475 and MDA-MB-231 breast cancers respectively (data not shown).

Table 1 Effects of therapy with targeted cytotoxic analog of bombesin AN-215 and its components on the growth of four human breast carcinoma lines xenografted into nude mice

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Tumor weight (mg) (% inhibition)</th>
<th>Tumor doubling time (days)</th>
<th>WBC on day 8 (cells/mm³)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Experiment 1 (MDA-MB-231)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>505.7±125.3</td>
<td>14.6±2.0</td>
<td>10 175±248.0</td>
</tr>
<tr>
<td>AN-215</td>
<td>187.7±42.0 (62.9)*</td>
<td>29.2±7.4*</td>
<td>8085±338.2</td>
</tr>
<tr>
<td>AN-201</td>
<td>479.7±97.1 (5.1)</td>
<td>15.6±1.1</td>
<td>6490±639.8*</td>
</tr>
<tr>
<td>Mixture</td>
<td>469.0±272.9 (7.3)</td>
<td>20.5±3.0</td>
<td>N.D.</td>
</tr>
<tr>
<td>RC-3094</td>
<td>475.2±183.7 (6.0)</td>
<td>17.1±1.7</td>
<td>N.D.</td>
</tr>
<tr>
<td>Blockade</td>
<td>461.8±221.0 (8.7)</td>
<td>18.8±3.1</td>
<td>N.D.</td>
</tr>
<tr>
<td>Experiment 2 (DU-4475)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>2105.4±301.7</td>
<td>4.8±0.22</td>
<td>9925.5±400.1</td>
</tr>
<tr>
<td>AN-215</td>
<td>1245.4±113.8 (40.7)*</td>
<td>7.0±0.9*</td>
<td>7095±1008.7</td>
</tr>
<tr>
<td>AN-201</td>
<td>1860±267.8 (11.6)</td>
<td>5.2±0.6</td>
<td>5390±614.1**</td>
</tr>
<tr>
<td>Experiment 3 (MDA-MB-435)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>494.4±95.4</td>
<td>11.8±0.9</td>
<td>10 037.5±609.8</td>
</tr>
<tr>
<td>AN-215</td>
<td>272.9±31.1 (44.8)*</td>
<td>17.9±2.0*</td>
<td>8772.5±541.5</td>
</tr>
<tr>
<td>AN-201</td>
<td>388.9±64.4 (21.3)</td>
<td>13.8±2.0</td>
<td>6517.5±691.9*</td>
</tr>
<tr>
<td>Experiment 5 (MDA-MB-231)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>589.0±105.6</td>
<td>9.8±1.1</td>
<td>8442.5±939.2</td>
</tr>
<tr>
<td>AN-215</td>
<td>263.3±53.3 (55.3)*</td>
<td>15.0±1.5*</td>
<td>6765±1083.8</td>
</tr>
<tr>
<td>AN-201</td>
<td>498.8±141.2 (15.3)</td>
<td>10.7±3.2</td>
<td>5142.5±853.7*</td>
</tr>
<tr>
<td>Experiment 6 (MDA-MB-468)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>160.1±39.0</td>
<td>34.5±5.6</td>
<td>10 367.5±449.9</td>
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<tr>
<td>AN-215</td>
<td>50.6±5.1 (68.4)*</td>
<td>92.2±18.8*</td>
<td>9377.5±1834.3</td>
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<tr>
<td>AN-201</td>
<td>116.4±38.1 (27.3)</td>
<td>68.4±20.8</td>
<td>6875±1060.3*</td>
</tr>
</tbody>
</table>

*In experiment 5 and 6 the compounds were administered twice. N.D., not determined. **P<0.05, ***P<0.01 (two-sided Student’s t-test).
MDR-1, MRP-1 and BCRP mRNA expression by real-time PCR

mRNA for MDR-1, MRP-1 and BCRP was detected in MDA-MB-231, MDA-MB-435 and DU-4475 breast cancers. On ethidium bromide gels, a single band of the expected size for each gene was seen. The PCR products were of the expected sizes of 95 bp for MDR-1, 127 bp for MRP-1, 140 bp for BCRP and 140 bp for β-actin (Fig. 4). The primers did not amplify any product using genomic DNA or the RT product. The efficiencies (E) were 1.989 for MDR-1, 1.999 for MRP-1, 1.987 for BCRP and 1.997 for β-actin. In MDA-MB-231, MDA-MB-435 and DU-4475 cancers, a single dose of 200 nmol/kg of AN-215 or AN-201 caused a similar, weak induction (1- to 1.8-fold) of MDR-1 gene expression. In MDA-MB-231 tumors, MRP-1 levels were slightly higher after therapy with AN-215 (3.1- vs 2.1-fold), and BCRP expression in this cell line was also higher after administration of AN-201 (1.4- vs 1-fold). In MDA-MB-435 cancers, expression levels of mRNA for MRP-1 and BCRP were slightly higher after treatment with AN-215 (respectively 2.6- vs 1.7-fold and 3.0- vs 0.9-fold). MRP-1 and BCRP mRNA levels of DU-4475 cancers did not increase after treatment with AN-215 and AN-201.

Discussion

Recent findings indicate that a high percentage of human breast carcinomas overexpress specific BN/GRP receptors, which bind bombesin with high affinity (Halmos et al. 1995, Gugger & Reubi 1999).

Figure 3 Expression of mRNA for the GRPR, NMBR and BRS-3 in human breast cancer cell lines as revealed by RT-PCR. PCR products were separated on a 1.8% agarose gel and stained with ethidium bromide. Lane M, 100-base pair DNA ladder; lanes 1–3, MDA-MB-231; lanes 4–6, MDA-MB-435; lanes 7–9, DU-4475; lanes 10–12, MX-1; lane 13, MDA-MB-468 (used as a positive control); lane -, total RNA from MDA-MB-231 tumor samples.

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Scintigraphy with radiolabeled bombesin/GRP analogs targeting BN/GRP receptors was successfully used for the detection of primary breast cancers and their lymph node metastases in several clinical studies (Van de Wiele et al. 2000, Scopinaro et al. 2002, Soluri et al. 2003). For the targeted chemotherapy of BN/GRP receptor-positive cancers, we developed a cytotoxic hybrid of a bombesin analog linked to 2-pyrrolino-DOX. This conjugate, coded AN-215, retains high affinity to BN/GRP receptors and a potent anti-proliferative activity (Nagy et al. 1997).

In the current study, we showed that DU-4475, MDA-MB-231, MDA-MB-435, MDA-MB-468 and MX-1 breast cancer cell lines express specific, high affinity binding sites for BN and mRNA for the GRPR subtype. All the cell lines used in this study are

<table>
<thead>
<tr>
<th>Cell line</th>
<th>Treatment</th>
<th>$K_d$ (nM)</th>
<th>$B_{max}$ (fmol/mg protein)</th>
</tr>
</thead>
<tbody>
<tr>
<td>MDA-MB-231</td>
<td>Control</td>
<td>4.81 ± 0.73</td>
<td>332.8 ± 12.6</td>
</tr>
<tr>
<td>DU-4475</td>
<td>Control</td>
<td>1.41 ± 0.03</td>
<td>534.6 ± 18.1</td>
</tr>
<tr>
<td>MDA-MB-468</td>
<td>Control</td>
<td>1.05 ± 0.06</td>
<td>470.9 ± 60.9</td>
</tr>
<tr>
<td>MDA-MB-435</td>
<td>Control</td>
<td>6.61 ± 0.14</td>
<td>315.8 ± 4.3</td>
</tr>
<tr>
<td>MX-1</td>
<td>Control</td>
<td>0.57 ± 0.02</td>
<td>152.1 ± 33.5</td>
</tr>
</tbody>
</table>

Binding characteristics were obtained from 12-point displacement experiments. $^{125}$I-labeled [Tyr$^4$]-bombesin was used as radioligand.

**Figure 4** Expression of mRNA for MDR-1, MRP-1 BCRP and β-actin in human breast cancers as revealed by RT-PCR. PCR products were separated on a 1.8% agarose gel and stained with ethidium bromide. Specific bands at the expected sizes of 95 bp for MDR-1, 127 bp for MRP-1, 140 bp for BCRP and 140 bp for β-actin were found in all three human breast cancer cell lines. Lanes 1 and 2 are MDA-MB-231 control samples; lanes 3 and 4 are MDA-MB-435 control samples; lanes 5 and 6 are DU-4475 control samples. The negative control is total RNA. The positive controls for MDR-1, MRP-1 and BCRP were the A498 human renal cell cancer cell line, the H4-60 human lung cancer line and the MCF-7 human breast cancer line respectively.
After treatment with AN-215 occurred probably due to the systemic toxicity. The moderate decrease in WBC was not significantly suppressed at any time, while AN-201 significantly lowered WBC 8 days after treatment in all experiments. This indicates that targeted therapy decreases myelotoxicity of AN-215 (due to AN-201) is expected to be further reduced in patients. Nevertheless, as GRPR are known to be expressed in the gastrointestinal tract, some side effects on the alimentary system could be anticipated. To evaluate such potential side effects, in previous studies we determined GRP-stimulated gastrin secretion in animals after treatment with AN-215 (Plonowski et al. 2000, Szereday et al. 2002). It was found that GRP-stimulated gastrin secretion was not significantly affected after treatment with AN-215. In addition, a recent histopathological evaluation of the gastrointestinal tract after treatment with AN-215 or its radical AN-201 at 250 nmol/kg revealed only negligible histological changes. Based on these results, it is reasonable to expect relatively minor side effects to the gastrointestinal system.

The increase in the mRNA levels for the multidrug resistance proteins MDR-1, MRP-1 and BCRP after treatment was small after single doses of AN-215 and AN-201, with no major difference after targeted and nontargeted therapy. Thus, a substantially accelerated development of MDR should not be expected to occur after targeted therapy. However, this topic should be reexamined in further studies to determine the MDR protein levels after multiple doses of AN-215.

In conclusion, our results demonstrate that a panel of five human breast cancer cell lines tested expressed receptors for bombesin/GRP and responded to therapy with the targeted cytotoxic bombesin analog AN-215 with a significant tumor inhibition. Further development of the targeted cytotoxic bombesin/GRP analog AN-215 is in progress. This analog should be available for clinical trials in the near future.

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