VEGF-C and COX-2 expression in papillary thyroid cancer

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Abstract

In papillary thyroid cancer (PTC), age appears to be the most important single prognostic factor. Another characteristic feature is the lack of association between survival and lymph node metastases. Earlier, we found that expression of cyclooxygenase-2 (COX-2) is higher in older PTC patients, in agreement with the finding that older patients have a worse prognosis. Recent findings suggest that COX-2 can up-regulate vascular endothelial growth factor-C (VEGF-C) expression. Here, we investigated whether expression of VEGF-C differs between young and older PTC patients and whether expression of VEGF-C and COX-2 are correlated. Our retrospective study comprised 106 PTC patients selected by age: those under 35 or over 55 at diagnosis. Paraffin-embedded tissue samples were analysed by immunohistochemistry for VEGF-C protein expression. Furthermore, we investigated by quantitative RT-PCR and enzyme immunoassay the relationship between VEGF-C and COX-2 expression in papillary thyroid cancer cells (NPA cells). VEGF-C expression was significantly increased with age. In the tumours from older lymph node-positive (N1) patients, VEGF-C expression was significantly higher than in the tumours from young N1 patients. Moreover, all patients who died of cancer or who developed distant metastases were old, and most tumours from these patients (4 of 5) expressed VEGF-C and had had nodal metastases at the time of primary operation. Immunohistochemically, expression of COX-2 and VEGF-C correlated strongly. In cell culture, this correlation was not so clear, because the COX-2 selective inhibitor, NS-398, did not reduce VEGF-C expression. However, as both COX-2 and VEGF-C were induced by the tumour promoter phorbol 12-myristate 13-acetate (PMA), the same factors may control them both.

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Introduction

Papillary thyroid cancer (PTC) is a unique cancer for two reasons; first, its prognosis depends largely on age, and secondly, lymph node metastases (LNM) do not necessarily indicate poor survival. Many authors have reported that lymph node metastases are associated with an increased rate of locoregional recurrence of disease but not with survival (Simpson et al. 1987, DeGroot et al. 1990, McHenry et al. 1991, Lin et al. 1999, Beasley et al. 2002). Some authors have, however, shown that in older patients, LNM have an adverse effect on survival (Harwood et al. 1978, Mazzaferri & Jhiang 1994, Passler et al. 2004). LNM, common in PTC, are more frequent among young patients whose prognosis is excellent. Such good prognosis in young patients could be explained by the hypothesis that these patients have a unique genetic pattern permitting adherence and retention of cells in regional lymph nodes that is unrelated to their capacity to grow in other organs (Cady 1998). In agreement with this hypothesis, lymph node metastases have been associated with poor prognosis only in older PTC patients (Harwood et al. 1978, Tubiana et al. 1985, Mazzaferri & Jhiang 1994).
We previously reported that cyclooxygenase-2 (COX-2) expression is higher in the tumours from older PTC patients and that this could explain the more aggressive behaviour of PTC in the older age group (Siironen et al. 2004). COX-2 is involved in the formation of prostanoids from arachidonic acid and, in normal tissues, it is induced in response to a wide range of cellular signals. Higher COX-2 expression correlates with a poor clinical outcome in various cancers (Sheehan et al. 1999, Salmennoki et al. 2001, Buskens et al. 2002, Denkert et al. 2002, Ristimäki et al. 2002), and selective inhibition of COX-2 suppresses tumour growth (Molina et al. 1999, Grossman et al. 2000, Higashi et al. 2000, Souza et al. 2000). Moreover, COX-2 expression is associated with lymph node metastasis (Ryu et al. 2000, Costa et al. 2002).

Recently, a novel correlation of COX-2 with vascular endothelial growth factor-C (VEGF-C) was reported in human lung adenocarcinoma, and COX-2 was found to up-regulate VEGF-C expression (Su et al. 2004). Later, correlation of VEGF-C with COX-2 was also demonstrated in head and neck squamous cell carcinoma and in oesophageal squamous cell carcinoma (Byeon et al. 2004, Kyzas et al. 2004). VEGF-C, originally found in the human prostatic adenocarcinoma cell line (Joukov et al. 1996), stimulates lymphatic proliferation and is expressed in cancers metastasizing to lymph nodes (Jeltsch et al. 1997, Mandriota et al. 2001). Papillary thyroid cancer also expresses VEGF-C (Bunone et al. 1999, Fellmer et al. 1999, Shushanov et al. 2000, Tanaka et al. 2002a,b, Hung et al. 2003). Some of these studies have also demonstrated correlations between lymph node metastasis and high VEGF-C levels (Bunone et al. 1999, Tanaka et al. 2002a,b), but no correlations with other clinical parameters. To the best of our knowledge, the relationship between age at diagnosis and VEGF-C expression in PTC has not been reported previously.

To address the issue of age and LNM in PTC, we compared the expression of VEGF-C in young and older PTC patients. Due to the hypothesis of differently behaving lymph node metastases in young and older PTC patients (Cady 1998), we hypothesized that the lymphangiogenic factor VEGF-C is expressed differently in these two age groups. Supporting this idea is our earlier finding of higher COX-2 expression in tumours from older patients (Siironen et al. 2005). As others have demonstrated COX-2-mediated VEGF-C up-regulation (Byeon et al. 2004, Kyzas et al. 2004, Su et al. 2004), we immunohistochemically examined VEGF-C expression in PTC, comparing it with the expression of COX-2, and studied the relationship of VEGF-C and COX-2 expression in a papillary thyroid cancer cell line (NPA). Phorbol 12-myristate 13-acetate (PMA) is a well-known inducer of COX-2 in several cell types and is also known to induce VEGF-C. Previously, we have also shown that PMA stimulates both COX-2 and VEGF-C expression (Enholm et al. 1997, van Rees et al. 2003). To block PMA-induced prostanoid production, we used the COX-2 selective inhibitor, NS-398, to see if VEGF-C expression is dependent on COX-2.

Materials and methods

Patients

During a 10-year period (1987 through 1996), 259 consecutive patients with papillary thyroid cancer were treated at the Department of Surgery of Helsinki University Central Hospital (HUCH). For comparison of young versus old patients, those younger than 35 years and those older than 55 years at diagnosis (cut-off age of tumour-node-metastasis (TNM) classification ±10 years) (Sobin & Wittekind 1997) were selected (n = 128) and their clinical data were retrospectively reviewed. Appropriate clinical information and follow-up data were available on 108 (84%) of these patients. Two patients were excluded; one had histology of tall cell variant and anaplastic carcinoma, and the other had cystic small papillary carcinoma. Therefore, 106 patients were included in this study. Papillary thyroid carcinomas were staged according to the TNM classification of 1997 (Sobin & Wittekind 1997).

Most of these patients underwent total thyroidectomy (98 of 106), and all patients having a positive postoperative whole body scan received radioactive iodine (93 of 98). The eight patients who underwent lobectomy had microcarcinoma.

Histologically, of the 106 tumours, 103 (97%) were of the usual type, two were the diffuse sclerosing variant and one was a Hürthle cell variant (all three patients were under 35 years old). No follicular variant or tall cell variant was included in our series. The typical papillary carcinoma is referred to as the usual type. The diagnosis of PTC is based on nuclear features, which should be present in a significant proportion of the neoplasm (Vickery et al. 1985). Initial lymph node metastases (N1) were present in 26 patients (25%), most of whom (19 of 26, 73%) were under 35 years of age. At the
time of initial surgery, no patients presented with distant metastases, but during follow-up four older patients (aged 61 to 81 years) developed distant metastases (pulmonary 2, mediastinal 1, pulmonary and skeletal 1). Median follow-up in young patients was 10.1 years (range 5.1–15 years) and in older patients it was 9.2 years (range 5.1–14.3 years).

Tumours recurred in 27 patients (25%), defined as evidence of disease (local or distant) appearing more than 6 months after the primary operation. Death was related to PTC in 3 of the 106 patients (3%), all of whom were old (range 61–81 years) and had primary LNM. The patients' clinical characteristics are shown in Table 1. Clinical staging was based on the TNM classification of the International Union Against Cancer (Sobin & Wittekind 1997).

### Immunohistochemistry

Formalin-fixed, paraffin-embedded archival tissue blocks of the 106 tumour samples were freshly cut into 4-μm sections onto slides and dried to fix for 12 to 24 h at 37°C. Freshly cut sections were deparaffinized in xylene and rehydrated through graded alcohol and deionized water. Then sections were heated in a microwave oven in 0.3% citrate buffer (pH 6.0) for 4–5 min, and washed with phosphate-buffered saline (PBS) for 2–5 min. Endogenous peroxidase activity of the tissues was blocked by 30-min incubation in methanol containing 1.6% hydrogen peroxide followed by a PBS wash for 3–5 min. Afterwards, the sections were treated with normal horse serum for 15 min. A rabbit polyclonal antibody against human VEGF-C (Zymed Laboratories, Inc., South San Francisco, CA, USA) was then applied at a dilution of 1:500 overnight.

After this overnight incubation, the sections were first reacted for 30 min with biotinylated anti-mouse IgG diluted in PBS containing 1% normal horse serum or with anti-rabbit IgG diluted in PBS containing 1% normal goat serum, and then for 30 min in avidin-biotinylated peroxidase complex (Vectastain ABC kits, Vector Laboratories, Inc., Burlingame, CA, USA). Between all steps in the

### Table 1 Clinical characteristics of patients with papillary thyroid cancer aged under 35 and over 55 at diagnosis.

<table>
<thead>
<tr>
<th>Patient characteristics</th>
<th>Under 35</th>
<th>Over 55</th>
<th>All patients</th>
<th>P-value*</th>
</tr>
</thead>
<tbody>
<tr>
<td>No. of patients</td>
<td>57</td>
<td>49</td>
<td>106</td>
<td></td>
</tr>
<tr>
<td>Extent of tumour</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>T1 (≤10 mm)</td>
<td>9</td>
<td>14</td>
<td>23</td>
<td>0.055</td>
</tr>
<tr>
<td>T2 (&gt;10 mm, ≤40 mm)</td>
<td>37</td>
<td>24</td>
<td>61</td>
<td></td>
</tr>
<tr>
<td>T3 (&gt;40 mm)</td>
<td>6</td>
<td>1</td>
<td>7</td>
<td></td>
</tr>
<tr>
<td>T4 (beyond the capsule)</td>
<td>5</td>
<td>10</td>
<td>15</td>
<td></td>
</tr>
<tr>
<td>Lymph node metastases</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Absent</td>
<td>38</td>
<td>42</td>
<td>80</td>
<td>0.019</td>
</tr>
<tr>
<td>Present</td>
<td>19</td>
<td>7</td>
<td>26</td>
<td></td>
</tr>
<tr>
<td>Distant metastases</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Absent</td>
<td>57</td>
<td>49</td>
<td>106</td>
<td>0.001</td>
</tr>
<tr>
<td>Present</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td></td>
</tr>
<tr>
<td>TNM stage†</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>I</td>
<td>57</td>
<td>14</td>
<td>71</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>II</td>
<td>0</td>
<td>22</td>
<td>22</td>
<td></td>
</tr>
<tr>
<td>III</td>
<td>0</td>
<td>13</td>
<td>13</td>
<td></td>
</tr>
<tr>
<td>IV</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td></td>
</tr>
<tr>
<td>Size (cm)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt;1</td>
<td>9</td>
<td>14</td>
<td>23</td>
<td>0.617</td>
</tr>
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<td>1–2</td>
<td>27</td>
<td>18</td>
<td>45</td>
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<td>2–4</td>
<td>15</td>
<td>13</td>
<td>28</td>
<td></td>
</tr>
<tr>
<td>&gt;4</td>
<td>6</td>
<td>4</td>
<td>10</td>
<td></td>
</tr>
<tr>
<td>Recurrent disease</td>
<td>20</td>
<td>7</td>
<td>27</td>
<td>0.012</td>
</tr>
</tbody>
</table>

*Analysis by Chi-square test, or by Fisher’s exact test when applicable. NS, not significant.
†Tumour-node-metastases (TNM) stage according to UICC TNM classification of 1997.
staining procedure, sections were washed three times with PBS for 5 min. Staining was visualized with 3-amino-9-ethyl-carbazole (A5754; Sigma, St Louis, MO, USA), 0.2 mg/ml in 0.5 M acetate buffer containing 0.03% hydrogen peroxide (pH 5.0) for 15 min. Subsequently, the sections were washed thoroughly in tap water. Meyer’s haematoxylin served as a counterstain. Finally, the sections were mounted in an aqueous mounting medium (Aquamount; BDH, Poole, Dorset, UK).

Specimens of breast cancer served as positive controls. As the negative control we used PBS instead of the primary antibody.

Scoring of VEGF-C was performed by an experienced pathologist (SN) and by an investigator (PS) without knowledge of the clinical data. Cytoplasmic immunoreactivity for VEGF-C was evaluated according to the extent of staining, with staining intensities scored as: 0, no cells stained; 1, less than 10% of cells stained; 2, 11% to 50% of cells stained; 3, more than 50% of cells stained. For statistical analysis, we grouped patients into two groups, positive expression being defined by staining of more than 10% of the tumour area (scores 2–3).

Statistical analysis

Correlation of VEGF-C with clinical variables (age, gender, stage, tumour size, and recurrent disease) was assessed with the Chi-square test or with Fisher’s exact test when applicable. Data were evaluated in three categories separately: young, old, and all patients. VEGF-C expression was also compared with that of COX-2 in the same specimens (Siironen et al. 2004). A P-value less than 0.05 was regarded as statistically significant.

Cell culture

Human papillary thyroid carcinoma cells (NPA) were grown in RPMI supplemented with 10% fetal calf serum (Promo Cell, Heidelberg, Germany), 2 mM L-glutamine, and antibiotics (BioWhittager, Verviers, Belgium). Cells were incubated with PMA (10 ng/ml; Sigma), NS-398 (5–10 μM; Cayman Chemical Co, Ann Arbor, MI, USA), or arachidonic acid (10 μM; Sigma) for the time periods indicated.

Prostaglandin E2 (PGE2) and VEGF-C enzyme immunoassay (EIA)

PGE2 levels were measured in cell culture medium by an EIA from Cayman Chemical Co., and VEGF-C levels were measured by an EIA from Zymed Laboratories according to the manufacturers’ instructions.

Real-time quantitative RT-PCR

Total RNA from NPA cells was isolated using Trizol Reagents (Invitrogen, Carlsbad, CA, USA), and RNA (1 μg) was converted to cDNA with Moloney murine leukaemia virus reverse transcriptase (Promega, Madison, WI, USA). RNasin (Promega), 2’-deoxyxynucleoside 5’-triphosphates (Amersham Pharmacia Biotech Inc, Bucks, UK), and random primers (Invitrogen). COX-2 and VEGF-C mRNA expression was quantified by TaqMan real-time RT-PCR and the Gene Amp 5700 Sequence Detection System (Applied Biosystems, Foster City, CA, USA). VEGF-C primers and TaqMan probe sequences were those used by Hung et al. (2003) and were purchased from Sigma. Human TATA-binding protein (TBP) served as an endogenous control to normalize expression of VEGF-C and COX-2 mRNA levels. COX-2 and TBP Assays-on-Demand came from Applied Biosystems. Each reaction was run in triplicate, with relative quantitation performed by the comparative Ct method (Applied Biosystems User Bulletin #2).

Results

VEGF-C immunohistochemistry

Tumours from older patients were more often judged to have positive VEGF-C staining (P < 0.001) compared with the younger group. Among all 106 tumours, positive VEGF-C expression was evident in 40 (37.7%): in 12 out of 57 (21.1%) in younger patients and in 28 out of 49 (57.1%) in older patients.

Lymph node metastases were present in 26 patients, and VEGF-C expression was positive in 8 of these tumours. VEGF-C immunoreactivity appeared in 2 of 19 younger and in 6 of 7 older patients, i.e. most of the older node-positive patients were VEGF-C-positive, and conversely, most of the young N1 patients were VEGF-C-negative (P = 0.001) (Table 2).

Of the 106 patients, we classified five patients as being high-risk. These patients either died of cancer or developed distant metastases during follow-up. All five high-risk patients were old. In four patients the tumours expressed VEGF-C (Table 3).
VEGF-C expression correlated strongly with the expression of COX-2 when all patients were analysed together ($P < 0.001$) (Table 4). In the young patient group, this correlation was not significant ($P = 0.120$) but it was significant in the old patient group ($P = 0.006$).

### Cell culture

We showed that NPA cells expressed VEGF-C and COX-2 mRNA and that their expression was induced by PMA (Fig. 1). As a measure of the expression of COX-2 at the protein level, enzyme activity was analysed by measuring PGE$_2$ production after arachidonic acid incubation. PMA induced production of PGE$_2$, and the COX-2 selective inhibitor, NS-398, blocked both basal and PMA-induced PGE$_2$ synthesis (Fig. 2). To study whether COX-2-derived prostanoids are responsible for VEGF-C expression in NPA cells, we treated the cells with PMA and NS-398 and measured VEGF-C protein expression by EIA. PMA caused a 2.7-fold increase in VEGF-C protein production after 24 h incubation. NS-398 failed, however, to reduce the basal or PMA-induced VEGF-C expression (Fig. 3).

### Discussion

In our series of 106 PTC patients, VEGF-C expression was significantly increased with age and, interestingly, most tumours from older N1 patients expressed VEGF-C whereas tumours from younger N1 patients did not ($P = 0.001$). All patients who died of cancer or who developed distant metastases were old, and most tumours from these patients expressed VEGF-C and had had nodal metastases at the time of primary operation.

As age is the most important prognostic factor in PTC, our aim was to compare younger and older PTC patients. In the classification systems for PTC, the cut-off age lies between 40 and 70 years (AGES, 40; AMES, 41 in men, 51 in women; MACIS, 40; SAG, 70; TNM, 45) (Hay et al. 1987, Cady & Rossi 1988, Akslen 1993, Hay et al. 1993, Sobin & Wittekind 1997). Since all these ages are estimates, we assumed that results would be more reliable when comparing patients under 35 with those over 55 years (45 years according to TNM/C6 10 years).

PTC is the only human cancer that shows no clear-cut relationship between lymph node metastases and survival (Bacourt et al. 1986, DeGroot et al. 1990, Samaan et al. 1992). Some studies have, however, shown that LNM also correlate with death in PTC, especially in older patients.

### Table 2

<table>
<thead>
<tr>
<th>Age group</th>
<th>Nodal metastases</th>
<th>VEGF-C-positive (n)</th>
<th>VEGF-C-negative (n)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Under 35 (n = 57)</td>
<td>N0 10</td>
<td>28</td>
<td></td>
</tr>
<tr>
<td></td>
<td>N1 2</td>
<td>17</td>
<td></td>
</tr>
<tr>
<td>Over 55 (n = 49)</td>
<td>N0 22</td>
<td>20</td>
<td></td>
</tr>
<tr>
<td></td>
<td>N1 6</td>
<td>1</td>
<td></td>
</tr>
</tbody>
</table>

* Nodal metastases: N0, absent; N1, present.

Patients with LNM showed significant differences in VEGF-C expression between age groups ($P = 0.001$). Those without LNM also showed significant differences between age groups ($P = 0.017$).

### Table 3

<table>
<thead>
<tr>
<th>Patient number</th>
<th>TNM classification at diagnosis</th>
<th>Age (years)</th>
<th>Sex</th>
<th>Distant metastases during follow-up</th>
<th>Death from PTC</th>
<th>VEGF-C expression</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>T2N1M0</td>
<td>61</td>
<td>Female</td>
<td>+ lung, bone</td>
<td>+</td>
<td>–</td>
</tr>
<tr>
<td>2</td>
<td>T4N1M0</td>
<td>81</td>
<td>Female</td>
<td>+ mediastinum</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>3</td>
<td>T4N1M0</td>
<td>78</td>
<td>Female</td>
<td>– –</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>4</td>
<td>T4N1M0</td>
<td>62</td>
<td>Male</td>
<td>+ lung</td>
<td>–</td>
<td>+</td>
</tr>
<tr>
<td>5</td>
<td>T2N0M0</td>
<td>80</td>
<td>Female</td>
<td>+ lung</td>
<td>–</td>
<td>+</td>
</tr>
</tbody>
</table>

*T = extent of tumour: T1 (≤10 mm), T2 (10 mm, ≤40 mm), T3 (40 mm), T4 (beyond thyroid capsule); N = nodal metastases: N0 absent, N1 present; M = distant metastases: M0 absent, M1 present.

### Table 4

<table>
<thead>
<tr>
<th>COX-2</th>
<th>VEGF-C</th>
<th>No. (%)</th>
<th>Low (%)</th>
<th>High (%)</th>
<th>P-value*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Low</td>
<td>66 (62.3)</td>
<td>52 (76.5)</td>
<td>14 (36.8)</td>
<td>&lt;0.001</td>
<td></td>
</tr>
<tr>
<td>High</td>
<td>40 (37.8)</td>
<td>16 (23.5)</td>
<td>24 (63.2)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*Statistical analysis by Chi-square test, or by Fisher’s exact test when applicable.
Our results agree with these studies. Four older patients developed distant metastases, and three of these also had nodal metastases. As all three patients dying of PTC were old, with LNM, we suggest that LNM in older patients seems to be a sign of aggressive disease as it is in other cancers. Our patients’ LNM were significantly more common among the younger group (33% vs 14%), and local lymph node recurrences were also more often seen in younger patients (35% vs 14%). Even so, none of the younger patients died of cancer or developed distant metastases. Thus, in the young, LNM did not affect survival: the cancer remains in the neck region. We therefore agree with the hypothesis of Cady (1998) that in young patients, metastatic cells in regional lymph nodes lack the capacity to grow in other organs. In our older patients the situation, however, differed, and primary LNM were correlated with poor survival.

It is notable that in our series only 25% of patients had nodal metastases (young patients 33%; older patients 14%). We do not routinely perform lymph node dissection but only remove pathological lymph nodes (so called node picking procedure). Small occult LNMs are apparently treated by routine postoperative radioiodine ablation. It is thus probable that our positive lymph nodes represent only the most aggressive metastases, those that may influence survival in contrast to micro-metastases.

We found higher VEGF-C expression in tumours from older patients, a finding which could explain the more aggressive behaviour of LNMs in those PTC patients. Research has shown that VEGF-C expression in thyroid tumours is higher in patients with nodal involvement, but no correlation has been evident with other clinical parameters (Bunone et al. 1999, Tanaka et al. 2002a). In contrast, other studies demonstrated no correlation between VEGF-C expression and the occurrence of LNMs (Shushanov et al. 2000, Hung et al. 2003). We found no correlation between nodal status and VEGF-C expression when all patients were analysed together. Interestingly, a significant difference was noticeable for young and older patients analysed separately: among lymph node-positive patients, VEGF-C expression was positive in most tumours (6 of 7) from those from older patients, but only 2 of 19 tumours from the young group expressed VEGF-C. Moreover, more than half of the old N1 patients (4 of 7) either died of cancer or developed distant metastases. In older patients,
VEGF-C up-regulation could therefore lead to increased distant dissemination of tumour cells. It could be that in older PTC patients, VEGF-C does not promote formation of lymph node metastases, but rather it induces tumour cell dissemination from lymph nodes to other organs. Supporting this hypothesis, work by Skobe et al. (2001) showed that increased VEGF-C promotes distant metastases. However, in our study, the sample size is not large enough to allow us to make a final statement on how VEGF-C induces tumour cell dissemination. Our results show a trend that VEGF-C could relate to distant dissemination of tumour cells in older patients. We need to study a larger patient series to confirm these observations.

We have reported that COX-2 expression is higher in tumours from older PTC patients (Siironen et al. 2004), and in this same patient series, immunohistochemical analysis of VEGF-C expression showed, consistent with previous results, that VEGF-C and COX-2 expression were correlated (Byeon et al. 2004, Kyzas et al. 2004). To examine any such correlation in vitro, we analysed COX-2 and VEGF-C expression in papillary carcinoma cells known to express COX-2 (Specht et al. 2002), and showed that NPA cells expressed both VEGF-C and COX-2 mRNA and that their expression was induced by PMA. Moreover, PMA induced the production of PGE₂, and the COX-2 selective inhibitor, NS-398, blocked both basal and PMA-induced PGE₂ synthesis, indicating that COX-2 is responsible for PGE₂ synthesis both in untreated NPA cells and under stimulatory conditions. NS-398 did not, however, reduce basal or PMA-induced VEGF-C protein expression. These data suggest that both VEGF-C and COX-2 are induced by PMA in NPA cells, but VEGF-C expression is not dependent on COX-2-derived prostanoids. The differences between our results and those of Su et al. (2004) may relate to the different cell lines studied; they studied human lung adenocarcinoma cell lines, finding that COX-2 enhanced VEGF-C gene expression through the prostanoid receptor (EP₁)-dependent transactivation of the HER-2/Neu receptor. Among the four subtypes of prostanoid receptors (EP₁-₄), EP₃ and EP₂ are expressed in almost all tissues, whereas EP₁ is restricted to organs such as the kidney, lung, and gastrointestinal tract (Watabe et al. 1993, Morimoto et al. 1997); the least abundant receptor is EP₂. Perhaps the difference in tissue distribution of the prostanoid receptors is one reason for these contradictory findings. Thus, because both COX-2 and VEGF-C are induced in NPA cells by PMA, the same factors may control the expression of these two markers. It would have been interesting to study several papillary thyroid cancer cell lines. We could not, however, find another papillary thyroid cancer cell line for our study.

In our study, the combination of old age, primary LNMs, and positive VEGF-C expression seems to indicate worse outcome. In older patients, but not in young ones, LNMs seem to affect survival. Based on our findings, we cannot conclude that VEGF-C is an independent indicator of aggressive disease, but higher VEGF-C in older patients could reflect the fact that in older patients, PTC is a biologically different disease.

Acknowledgements

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