Neuroendocrine-like prostate cancer cells: neuroendocrine transdifferentiation of prostate adenocarcinoma cells

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Abstract

Neuroendocrine (NE) cells represent a minor cell population in the epithelial compartment of normal prostate glands and may play a role in regulating the growth and differentiation of normal prostate epithelia. In prostate tumor lesions, the population of NE-like cells, i.e., cells exhibiting NE phenotypes and expressing NE markers, is increased that correlates with tumor progression, poor prognosis, and the androgen-independent state. However, the origin of those NE-like cells in prostate cancer (PCa) lesions and the underlying molecular mechanism of enrichment remain an enigma. In this review, we focus on discussing the distinction between NE-like PCa and normal NE cells, the potential origin of NE-like PCa cells, and in vitro and in vivo studies related to the molecular mechanism of NE transdifferentiation of PCa cells. The data together suggest that PCa cells undergo a transdifferentiation process to become NE-like cells, which acquire the NE phenotype and express NE markers. Thus, we propose that those NE-like cells in PCa lesions were originated from cancerous epithelial cells, but not from normal NE cells, and should be defined as ‘NE-like PCa cells’. We further describe the biochemical properties of newly established, stable NE-like lymph node carcinoma of the prostate (LNCaP) cell lines, transdifferentiated from androgen-sensitive LNCaP cells under androgen-deprived conditions. Knowledge of understanding NE-like PCa cells will help us to explore new therapeutic strategies for treating PCa.

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Introduction

Neuroendocrine (NE) cells are the third cell type in the epithelial compartment of normal prostate glands in addition to basal and secretory epithelial cells (Abrahamsson 1999). Though prostatic NE cells were described decades ago, their functional roles in prostate, especially in prostate cancer (PCa) progression, have received attention only recently. In normal prostate, NE cells are apparently involved in regulating epithelial cell growth and differentiation in an androgen-independent (AI) manner. In prostate carcinoma, the NE cell population, as characterized by the expression of NE markers, in tumor foci is increased, which evidently correlates with tumor progression, poor prognosis, and the hormone-refractory stage. The role of NE cells in PCa progression is suggested by the fact that those cells express and secrete a variety of neuropeptides which have mitogenic effects on adjacent cancer cells and thus can contribute to the AI proliferation of PCa cells (di Sant’Agnese 1992, 1998, di Sant’Agnese & Cockett 1996). Further studies also show that NE differentiation is found in the metastatic lesion and significantly associated with the mortality of PCa patients (Cheville et al. 2002, Roudier et al. 2003). Although the origin of NE cells and the molecular mechanism of NE cell enrichment during PCa progression are not fully understood, accumulated evidence including in vitro cell culture, in vivo animal model, and clinical sample analyses together indicate that PCa cells can undergo a transdifferentiation process to become NE-like cells, which
acquire the NE phenotype and express NE markers. Thus, we propose to define those NE-like cells in PCa lesions as ‘NE-like PCa cells’.

In this review, we focus our discussion on the potential origin of NE-like PCa cells and the molecular mechanism of NE transdifferentiation. In the first section, we discuss the differences between NE-like PCa and normal NE cells in their biochemical properties and origins. In the second section, we present data from in vitro and in vivo studies suggesting the multipathway of NE transdifferentiation occurring in PCa cells and discuss their underlying molecular mechanisms. In the final section, we describe the biochemical properties of newly established NE-like LNCaP stable subclone cells which represent a useful model mimicking clinical NE-like cells in prostate tumors. Knowledge of NE-like PCa cells will provide us with new insights for developing novel therapeutic strategies for treating PCa.

NE-like PCa versus normal NE cells
Differences in biochemical characteristics between NE-like PCa and normal NE cells

Several reviews have extensively described the characterizations of prostatic NE cells and their functions in both physiological and pathological states (di Sant’Agnese 1992, 1998, di Sant’Agnese & Cockett 1996, Abrahamsson 1999, Sciarra et al. 2003, Vashchenko & Abrahamsson 2005). However, an important question of whether NE-like PCa cells differ from normal NE cells is less addressed. While several studies in PCa archival specimens have shown that NE-like PCa cells share similar biochemical properties with NE cells in the normal prostate, including the expression of NE markers as well as the lack of AR and PSA, these reports also suggest several differences existing between NE-like PCa and normal NE cells (Table 1; Abrahamsson 1999, Vashchenko & Abrahamsson 2005). First, immunohistochemical studies have shown that NE cells in the normal prostate gland express cytokeratin 5 (K5), a basal cell marker (Schalken & van Leenders 2003, Hudson 2004); while NE-like PCa cells exhibit characteristics of luminal secretory cells by expressing prostatic acid phosphatase (PAcP; Huang et al. 2006) and K18, a luminal cell-specific cytokeratin (van Bokhoven et al. 2003, Vashchenko & Abrahamsson 2005). Huang et al. (2006) further showed that CgA-positive NE-like cells in PCa archival specimens are positively stained with antibodies (Abs) against luminal secretory cell-associated cytokeratins; while they are negative for staining with Abs against basal cell markers, including high molecular weight cytokeratin and p63 (Wojno &

<table>
<thead>
<tr>
<th>Table 1</th>
<th>Comparison of normal neuroendocrine (NE) and NE-like prostate cancer (PCa) cells in prostate</th>
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</table>
| **Similarities** | Lack of AR and PSA expression  
| | Autocrine/paracrine effects  
| | Non/low-proliferative activity  
| | Neuron-like morphology  
| | Expression of NE markers  
| | NSE  
| | CgA  
| | CgB  
| | Serotonin  
| | PTHrP  
| | NT  
| | Bombesin  
| **Dissimilarities (normal NE versus NE-like PCa cells)** | Expression of basal cell markers versus luminal secretory cell markers  
| | Non-aggressive versus highly aggressive  
| | No Bcl-2 expression versus Elevated Bcl-2 expression  
| | No AMACR expression versus AMACR expression  

NSE, neuron-specific enolase; CgA, chromogranin A; CgB, chromogranin B; PTHrP, parathyroid hormone-related protein; NT, neurotensin; Bcl-2, B cell lymphoma protein 2; AMACR, α-methylacyl-CoA racemase.  
* Some reports show no neuron-like phenotype, i.e., without processing, in NE-like PCa cells.

Epstein 1995, Signoretti et al. 2000). Secondly, compared with normal NE cells with non-proliferating and non-tumorigenic activities, NE-like PCa cells, such as small cell carcinoma cells being completely NE differentiated (Yao et al. 2006), are highly aggressive and exhibit tumorigenic activity (Oesterling et al. 1992). Thirdly, while NE cells from normal prostate and benign prostatic hyperplasia do not express anti-apoptotic protein B cell lymphoma protein 2 (Bcl-2) (Xue et al. 1997), studies in archival PCa samples show a correlation relationship between the expression levels of Bcl-2 and neuron-specific enolase (NSE; Segal et al. 1994). Fourthly, α-methylacyl-CoA racemase (AMACR), an enzyme involved in the β-oxidation of fatty acids, is only expressed in NE-like PCa cells and non-NE prostate tumor cells but not in normal prostatic cells including NE cells (Huang et al. 2006), suggesting the close linkage between NE-like PCa and prostate carcinoma cells. Finally, histological studies have clearly shown that normal NE cells exhibit two distinct morphologies which have a complex appearance with irregular dendrite-like processes extending between adjacent epithelial cells (di Sant’Agnese 1992), while some NE-like PCa cells may lack the typically neuron-like processes and are morphologically similar to the surrounding carcinoma cells (Xing et al. 2001). These data together suggest that NE-like PCa cells are clearly distinguished from NE cells in the normal prostate.
Origin of NE-like PCa versus normal NE cells

The distinctions between NE-like PCa and normal NE cells may be attributed to their origins. As shown in Fig. 1, results of studies indicate that normal NE cells, as basal and luminal epithelial cells, are originated from stem cells in the basal layer (Hudson 2004, Long et al. 2005). It is also possible that normal NE cells represent an independent cell lineage from the neurogenic origin, differing from secretory and basal cells (Aumuller et al. 1999). Thus, the origin of NE cells in normal prostate glands requires further investigations.

Similarly, the origin of NE-like PCa cells remains controversial. It is proposed that NE-like PCa cells share the same origin with normal NE cells and are differentiated from the intermediate stem cells (Bonkhoff et al. 1995, Bonkhoff 1998). The differentiation processes of those stem cells may be aberrantly regulated under pathological conditions like androgen ablation, resulting in an abnormally increased NE cell population. Although this stem cell-based hypothesis is attractive, it requires further validation. Alternatively, accumulated evidence suggests that adenocarcinoma cells can undergo a transdifferentiation process to become NE-like cells, which acquire a similar phenotype to normal NE cells and express several NE markers. Importantly, they still retain some epithelial characteristics (Fig. 1; Vashchenko & Abrahamsson 2005). This notion is supported by the observations that NCI-H660 and PSK-1 cells, two human prostatic small cell carcinoma cell lines, express K8/18 of luminal cell markers, but not K5/14 of basal cell markers (van Bokhoven et al. 2003). Furthermore, Bcl-2 expression is found in all specimens examined for prostatic small cell carcinoma (18/18; 100%), whereas its expression is observed in two out of ten cases (20%) of prostate adenocarcinoma (Yao et al. 2006) and no expression in normal NE cells (Xue et al. 1997), which suggests the correlative relationship of NE-like PCa cells and adenocarcinoma cells. In addition, NE differentiation can be seen in the bone metastatic lesion but not within the primary tumor foci, suggesting the occurrence of transdifferentiation process in metastatic PCa cells (Cheville et al. 2002, Roudier et al. 2003). Importantly, results of genetic analyses on clinical archival specimens further reveal that NE-like PCa cells share essentially identical allelic profiles with non-NE PCa cells, but not that of normal prostatic epithelium or normal NE cells (Sauer et al. 2006). These results collectively provide the evidence that NE-like PCa cells are originated from cancerous luminal epithelial cells, which undergo a transdifferentiation process (Fig. 1), and thus these cells should be defined as ‘NE-like PCa cells’, but not ‘NE cells’.

Multipathways involved in NE transdifferentiation of PCa cells

Androgen depletion-induced NE differentiation

Androgen ablation therapy is the main option for treating metastatic PCa; however, androgen withdrawal also contributes to increased NE
differentiation in PCa (Ismail et al. 2002). It has been shown that patients who had received a longer course of androgen ablation therapy would have higher levels of NE differentiation. Ito et al. (2001) reported that in archival specimens from 137 PCa patients, 70.5% of 68 patients who had received hormonal therapy for more than 13 months exhibited NE differentiation, higher than 44% of 25 patients with hormonal therapy less than 12 months and 31.8% in 44 patients without therapy. A similar result was also shown in Hirano et al. (2004) study, which supports the notion that androgen ablation therapy increases NE differentiation of PCa cells.

To support the observations in clinical specimens, androgen withdrawal-induced NE differentiation of PCa cells is also seen in in vitro cell culture (Table 2) and in vivo animal model (Table 3) studies. The initial study revealed that androgen-sensitive (AS) LNCaP cells exhibit the capability of being transdifferentiated into an NE-like phenotype in response to androgen deprivation (Shen et al. 1997). Our data further showed that androgen-reduced media can induce NE transdifferentiation of AS LNCaP cells, and these cells acquire the neuronal phenotype and express multiple NE markers, including NSE, CgA, gastrin, and neurotensin (Zelivianski et al. 2001, Yuan et al. 2006). This NE transdifferentiation induced by steroid-reduced (SR) medium is reversed by the addition of DHT. Conversely, Casodex, an androgen antagonist, can compete out DHT effect and restore NE transdifferentiation. These results suggest that androgen-activated AR signaling suppresses NE differentiation. This notion is supported by the observation that knockdown of AR in AS LNCaP cells induces NE transdifferentiation, indicated by acquiring NE-like morphology and expressing NE markers (Wright et al. 2003).

Studies on in vivo animal models validate in vitro observations that castration of mice bearing AS PC-295 or PC-310 xenograft tumors promotes their NE differentiation, indicated by the elevated expression of NE markers (Jongsma et al. 1999, 2000). Similar results were also observed in castrated animals bearing xenografts of AS LNCaP (Burchardt et al. 1999) and CWR22 cells (Huss et al. 2004) as well as castrated TRAMP mice (Kaplan-Lefko et al. 2003). Consistent to clinical observations, data from in vitro and in vivo studies collectively support the notion that androgen depletion induces transdifferentiation of AS adenocarcinoma cells to become NE-like cells in PCa lesions.

Mechanisms of androgen depletion-induced NE differentiation

Receptor protein tyrosine phosphatase α (RPTPα)-mediated signal pathway

RPTPα is a member of the transmembrane subfamily of PTPs and is widely expressed in mammalian tissues (Sap et al. 1990). In addition to the regulation of integrin signaling and potassium channel activity, RPTPα plays a critical role in neuronal differentiation (Pallen 2003). In AS LNCaP cells, the expression of RPTPα at both mRNA and protein levels is upregulated by androgen depletion, correlating with NE differentiation and NSE expression (Zelivianski et al. 2000, 2001). Furthermore, addition of androgens abolishes steroid depletion-induced RPTPα elevation, while Casodex can compete out androgen effects and restores RPTPα as well as NSE expression (Zhang et al. 2003). Further studies show that even in the regular culture condition containing androgenic activity, increased expression of RPTPα by cDNA transfection in AS LNCaP cells results in elevated levels of multiple NE markers, including NSE, chromogranin A (CgA), CgB, parathyroid hormone-related protein (PTHrP), and NT as well as acquired NE-like phenotype, suggesting a role of RPTPα in NE differentiation of AS PCa cells (Zhang et al. 2003, Yuan et al. 2006).

The molecular mechanism of RPTPα-induced NE differentiation in AS PCa cells is closely associated with the activation of c-Src and ERK/MAPK (Zhang et al. 2003, Yuan et al. 2006). This notion is supported by observations that elevated RPTPα expression in LNCaP cells leads to c-Src activation, while the expression of Y789F-RPTPα, a mutant of RPTPα which is devoid of interaction with c-Src, does not have an effect on c-Src activation and fails to induce NE marker expression as well as NE-like morphology (Yuan et al. 2006). Similarly, increased RPTPα expression in LNCaP cells also leads to ERK/MAPK activation, correlating with elevated expression of multiple NE markers, while this RPTPα-effect is abolished by PD98059, an MEK inhibitor (Zhang et al. 2003, Yuan et al. 2006). Importantly, PP2, an inhibitor of c-Src family kinases, can effectively abolish RPTPα-induced ERK/MAPK activation as well as NE marker expression, and thus blocks SR medium-induced NE-like morphology in LNCaP cells (Yuan et al. 2006). These results collectively suggest that in androgen depletion condition, RPTPα is elevated and constitutively activates the c-Src-MEK-ERK/MAPK signal pathway.
<table>
<thead>
<tr>
<th>Induction</th>
<th>Cell types</th>
<th>NE-like morphology</th>
<th>Growth arrest</th>
<th>Marker expression</th>
<th>PSA/AR expression</th>
<th>Biological phenotypes</th>
<th>Paracrine (CM) effects</th>
<th>R/IR</th>
<th>References</th>
</tr>
</thead>
</table>
| Androgen depletion | LNCaP           | Yes                | Yes           | NSE/S-
| SR medium        |                 |                   |               | 100/Bombesin       | Decreased PSA/AR     | ND                    | ND   | Shen et al. (1997)                  |
| LNCaP (C-33)     |                 | Yes                | ND            | NSE/CgA/GS/NT     | Decreased PSA     | ND                   | ND                     | ND   | Zelivianski et al. (2001)           |
| LNCaP (C-81)     |                 | ND                 | ND            | No effect         | Decreased PSA     | ND                   | ND                     | ND   | Zhang et al. (2003)                 |
| LNCaP (C-33)     |                 | Yes                | Yes           | NSE               | ND                | ND                   | ND                     | ND   | Yang et al. (2005)                  |
| LNCaP            |                 | ND                 | ND            | NSE               | ND                | ND                   | ND                     | ND   | Burchardt et al. (1999)             |
| cAMP-related     | LNCaP (C-33)    | Yes                | ND            | No effect         | Increased PSA     | ND                   | ND                     | ND   | Zelivianski et al. (2001)           |
| db-cAMP          | LNCaP (C-33)    | Yes                | ND            | NSE               | Decreased AR      | ND                   | ND                     | ND   | Bang et al. (1994)                  |
| db-cAMP + IBMX   | LNCaP (C-81)    | ND                 | ND            | NSE               | CgA               | ND                   | ND                     | ND   | Cox et al. (1999)                   |
| PC-3M            |                 | Yes                | Yes           | NSE/CgA/S-100     | ND                | ND                   | ND                     | ND   | Deeble et al. (2001)                |
| Forskolin + IBMX | LNCaP/C4-2      | Yes                | Yes           | NSE/Serotonin     | ND                | ND                   | ND                     | R    | Cox et al. (1999)                   |
| Epinephrine      | LNCaP           | Yes                | Yes           | NSE               | Decreased PSA/AR  | Decreased PSA/AR     | Decreased invasiveness | ND   | Wang et al. (2004b)                 |
| Epinephrine + IL-6 | LNCaP           | Yes                | Yes           | NSE               | Decreased PSA/AR  | Decreased PSA/AR     | Decreased invasiveness | R    | Deeble et al. (2001)                |
| Cytokines        | IL-6            | LNCaP | Yes | ND | NSE | ND | ND | ND | ND | Qin et al. (1998)                  |
|                 | LNCaP           | Yes                | Yes           | NSE               | ND                | ND                   | ND                     | ND   | Mori et al. (1999)                  |
|                 | LNCaP           | Yes                | Yes           | NSE               | ND                | ND                   | ND                     | ND   | Spiotto & Chung (2000a, b)          |
|                 | LNCaP (C-33)    | Yes                | Yes           | NSE               | Decreased PSA     | ND                   | ND                     | ND   | Deeble et al. (2001)                |
|                 | LNCaP (C-81)    | ND                 | ND            | No effect         | Decreased PSA     | ND                   | ND                     | ND   | Zelivianski et al. (2001)           |
|                 | LNCaP           | Yes                | Yes           | NSE/I/III tubulin | Decreased PSA/AR  | Decreased growth     | ND                     | ND   | Wang et al. (2004b)                 |

*Table 2 In vitro studies of neuroendocrine (NE) differentiation of prostate cancer (PCa) cells*


<table>
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<tr>
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<th>Cell types</th>
<th>NE-like morphology</th>
<th>Growth arrest</th>
<th>Marker expression</th>
<th>PSA/AR expression</th>
<th>Biological phenotypes</th>
<th>Paracrine (CM) effects</th>
<th>R/IR</th>
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<tbody>
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<td>IL-1β/IL-6</td>
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<td>ND</td>
<td>CgA</td>
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<td>ND</td>
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<td>ND</td>
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<td>ND</td>
<td>ND</td>
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<td>ND</td>
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<td>ND</td>
<td>NSE/CgA</td>
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<td>ND</td>
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<td>ND</td>
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<td>DU145</td>
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<td>ND</td>
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<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
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<td>Yes</td>
<td>NT</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>Farini et al. (2003)</td>
</tr>
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</table>

CM, conditioned medium; R/IR, reversible/irreversible phenotype; SR, steroid reduced; CA, constitutively active; K, cytokeratin; LNCaP (C-33), androgen-sensitive LNCaP cells; LNCaP (C-81), androgen-independent LNCaP cells; ND, not determined.
resulting in NE transdifferentiation of PCa cells (Fig. 2).

**PTP1B-mediated signal pathway**

The expression of PTP1B, a cytosolic PTP, is also found to have an elevated level in NE-differentiated LNCaP cells induced by androgen depletion (Wu et al. 2006). In the regular culture condition containing androgenic activity, increased expression of PTP1B in LNCaP cells can lead to NE differentiation. In an androgen-deprived condition, increased PTP1B expression or activity in LNCaP cells synergistically enhances NE marker expression. Conversely, the expression of a dominant-negative mutant of PTP1B blocks the upregulation of CgA expression in response to androgen withdrawal (Wu et al. 2006). Thus, PTP1B may play a role in androgen depletion-induced NE differentiation. Consistently, immunostaining of PCa archival specimens revealed the co-expression of PTP1B and CgA in tissues examined (Wu et al. 2006). Although the underlying mechanism of PTP1B-induced NE differentiation of PCa cells requires further investigation, PTP1B may activate the transdifferentiation process through upregulating the Ras-ERK/MAPK signaling (Dube et al. 2004). Alternatively, PTP1B, similar to RPTPα, can directly activate c-Src as observed in breast cancer cells (Bjorge et al. 2000). PTP1B-induced NE differentiation may also be mediated by AKT activation (Wu & Huang 2007).

**Protocadherin-PC (PCDH-PC)-mediated signal pathway**

PCDH-PC, a member of the protocadherin gene family encoded on the human Y chromosome, is selectively expressed in hormone-resistant PCa cells (Chen et al. 2002, Terry et al. 2006). The expression of PCDH-PC is upregulated in androgen-depleted LNCaP cells and in LNCaP xenograft tumors from castrated male mice (Chen et al. 2002). Increased expression of PCDH-PC leads to the activation of Wnt signaling, including the nuclear translocation of β-catenin and increased Tcf transcriptional activity (Yang et al. 2005). Elevated expression of PCDH-PC by cDNA transfection in LNCaP and PC-3 cells upregulates NSE and CgA expression, while its decreased expression by siRNA oligomers in LNCaP cells abolishes androgen depletion or PCDH-PC-induced NSE expression. Furthermore, NE transdifferentiation of LNCaP cells could be induced by β-catenin cDNA transfection or blocked by its siRNA oligomers (Yang et al. 2005).
These data together suggest that increased PCDH-PC expression in response to androgen deprivation via Wnt signaling plays a role in NE differentiation of PCa cells.

cAMP and its agonists-induced NE differentiation

Several lines of evidence show that elevation of intracellular cAMP leads to NE differentiation of AS and AI PCa cells (Table 2). Bang et al. (1994) first showed that increased cAMP, by adding cAMP analogs and phosphodiesterase inhibitors to AS LNCaP or AI PC-3M cell culture medium, induces their NE differentiation by acquiring neuron-like morphology, expressing NE markers, and suppressing cell growth. Further studies showed that elevation of intracellular cAMP by forskolin, an activator of adenylate cyclase, or epinephrine, a β-adrenergic receptor agonist, induces NE transdifferentiation of LNCaP cells (Cox et al. 1999). Additionally, androgen depletion also leads to cAMP elevation in AS LNCaP cells, correlated with NE differentiation (Burchardt et al. 1999). Nevertheless, the NE characteristics induced by cAMP-elevating agents are fully reversible upon the removal of the stimuli (Cox et al. 1999).

Mechanisms of cAMP-induced NE differentiation

Since increased cAMP induces the activation of protein kinase A (PKA) and treatment of epinephrine also leads to PKA activation (Deeble et al. 2001), it is thus proposed that PKA is mediating cAMP-induced transdifferentiation of PCa cells. This notion is supported by the observation that overexpression of PKA catalytic subunit in LNCaP cells leads to NE differentiation, while expression of a dominant-negative mutant of PKA blocks NE differentiation induced by cAMP-elevating agents (Cox et al. 2000). Since ERK/MAPK signal pathway crosstalks with cAMP/PKA-regulated proliferation and differentiation in several cell types (Stork & Schmitt 2002), ERK/MAPK may also play a role in the cAMP-elevating agent-induced NE differentiation of PCa cells. For example, epinephrine and db-cAMP treatments in LNCaP cells lead to ERK/MAPK activation (Chen et al. 1999). However, in other studies, the withdrawal of db-cAMP/IBMX from NE-differentiated LNCaP cells results in ERK/MAPK activation (Cox et al. 1999). Additionally, epinephrine treatment has no significant effect on ERK/MAPK activity in LNCaP cells, although it can lead to an increased cAMP level, the PKA activation, and the induction of NE differentiation (Deeble et al. 2001). The inconsistent observations could be contributed by different experimental conditions. It has been shown that transient cAMP accumulation in LNCaP cells leads to proliferation through PKA-dependent ERK/MAPK activation, while sustained elevation of cAMP level increases

Figure 2 Signal transduction pathways associated with the induction of NE transdifferentiation of PCa cells. The solid line indicates the reported pathways, while the dash line indicates the hypothetical pathways.
PKA activity resulting in NE differentiation by a PKA-dependent but ERK/MAPK-independent mechanism (Farini et al. 2003). The role of ERK/MAPK in cAMP/PKA signaling-mediated NE differentiation in PCa cells requires further investigation.

Cytokines-induced NE differentiation

Cytokines, a diverse group of secreted peptides, function as signaling molecules to mediate immune and inflammatory responses as well as to regulate the proliferation and maturation of immune cells. Importantly, cytokines such as interleukins can also regulate the proliferation and differentiation of non-immune cells, including PCa cells. Results of clinical studies reveal that the serum level of interleukin-6 (IL-6) is elevated in patients with PCa, especially hormone-refractory and metastatic PCa (Adler et al. 1999, Drachenberg et al. 1999, Shariat et al. 2001). Furthermore, the levels of IL-6 and its receptor in PCa tissues are significantly higher than that in normal prostate tissues (Siegsmund et al. 1994, Giri et al. 2001). These results clearly show the correlation of IL-6 expression with PCa progression and thus suggest that IL-6 may serve as a marker for PCa morbidity (Twillie et al. 1995). Since elevated IL-6 level is associated with advanced PCa and correlated with increased NE differentiation, it is proposed that IL-6 is one of the important inducers of NE differentiation of PCa cells. Indeed, IL-6 treatment induces NE transdifferentiation of LNCaP cells (Qiu et al. 1998), indicated by neuron-like morphology, reduced cell growth, and elevated NE marker expression (Table 2). Furthermore, in the absence of ligand, elevated PKA activity resulting in NE differentiation is closely associated with cholesterol-rich membrane lipid rafts. In LNCaP cells, the disruption of lipid rafts inhibits IL-6-mediated effects, probably because IL-6 receptors predominantly localizes to the lipid raft membrane compartment (Kim et al. 2004). Additionally, tyrosine kinase Etk/Bmx has been shown to play an important role in IL-6-induced NE differentiation (Qiu et al. 1998). Etk/Bmx may activate c-Src, Stat3, and/or PI3K for NE differentiation (Spiotto & Chung 2000b). Interestingly, IL-6-mediated Stat3 phosphorylation, nuclear translocation, and NE differentiation are closely associated with cholesterol-rich membrane lipid rafts. In LNCaP cells, the disruption of lipid rafts inhibits IL-6-mediated effects, probably because IL-6 receptors predominantly localizes to the lipid raft membrane compartment (Kim et al. 2004). Additionally, tyrosine kinase Etk/Bmx has been shown to play an important role in IL-6-induced NE differentiation (Qiu et al. 1998). Etk/Bmx may activate c-Src, Stat3, and/or PI3K for NE differentiation (Spiotto & Chung 2000b). Alternatively, c-Src can activate the Etk/Bmx-Stat3 pathway (Tsai et al. 2000). Further investigation is apparently needed.

Clinical relevance of IL-6-induced NE differentiation

IL-6-induced NE differentiation has been clearly demonstrated in LNCaP cells in culture; however, the role of those IL-6-induced NE-like cells in clinical prostate tumor progression is less understood. Studies showed that IL-6 inhibits the growth of LNCaP xenografts, despite the fact that those tumors exhibit NE characteristics including elevated levels of NSE and β-III tubulin (Wang et al. 2004a). Furthermore, co-culture of IL-6-induced NE-like cells with LNCaP, PC-3, or DU145 cells leads to cell cycle arrest and DNA synthesis inhibition (Wang et al. 2004b). These results suggest that IL-6-induced NE differentiation leads to the growth inhibition of surrounding PCa cells, probably via the release of inhibitory factors by IL-6-induced NE-like cells (Wang et al. 2004b). While the underlying mechanism still requires further investigations, these results are opposite to the observations in clinical specimens that the NE cell-proximal PCa cells exhibit higher proliferation indices than the distal PCa cells (Bonhoff et al. 1995). The data suggest that clinical NE cells in PCa lesions secrete growth stimulatory factors to enhance proximal PCa cell proliferation. It should also be noted that IL-6 exhibits a growth stimulatory effect on the primary prostate epithelial cells as well as a cell line derived from
precancorous lesion and high-grade prostatic intraepithelial neoplasia (Giri et al. 2001, Liu et al. 2002b). The contradictory effects of IL-6 on both PCa cell proliferation and NE differentiation may result from the bifunctional character of IL-6. It has been proposed that the paracrine effect by IL-6 leads to the growth inhibition and NE differentiation, while the autocrine effect by IL-6 causes the growth stimulation (Lee et al. 2007). Evidently, the role of IL-6-induced NE differentiation of PCa cells in clinical PCa progression under AI condition requires further clarification.

Other factors in NE differentiation

Results from studies clearly suggest multiple pathways for inducing NE transdifferentiation of PCa cells (Table 2 and Fig. 2; Zelivianski et al. 2001). For example, HB-EGF, an EGFR ligand serving as a mitogenic and survival factor for PCa cells, can also promote NE differentiation in vitro and in vivo (Adam et al. 2002, Kim et al. 2002). HB-EGF activates EGFR via ERK/MAPK to induce NE differentiation of LNCaP cells (Kim et al. 2002). Similarly, activation of EGFR by EGF can also induce NE differentiation of DU145 cells although the underlying molecular basis remains unclear (Humez et al. 2006). Contrarily, treatment of genistein, a classical tyrosine kinase inhibitor, has been shown to be associated with NE differentiation of LNCaP cells, which is correlated with the activation of Stat3 and ERK/MAPK (Pinski et al. 2006). Additionally, treatment of genistein abolishes EGF-induced NE differentiation of DU145 cells (Humez et al. 2006), and conditioned media from genistein-induced NE-like LNCaP cells have an inhibitory effect on the growth of PC-3 and DU145 cells (Pinski et al. 2006). Vasoactive intestinal peptide (VIP) is a neuropeptide and can also promote NE differentiation of LNCaP cells (Jaunrannz et al. 2001). Further studies reveal that VIP-induced NE differentiation is mediated by the activation of multiple signaling molecules, including PKA, ERK/MAPK, and PI3K (Gutierrez-Canas et al. 2005). Interestingly, intracellular calcium level is also associated with the development of NE phenotype in PCa cells (Vanoverberghe et al. 2004). Calcium chelator BAPTA/AM inhibits VIP-induced NE phenotype in LNCaP cells (Collado et al. 2005), while increased expression of calcium channel proteins in LNCaP cells promotes NE differentiation (Mariot et al. 2002). The treatments of some anticancer agents, such as jolkinolide B and silibinin, are also reported to be correlated with the induction of NE differentiation in LNCaP cells (Zi & Agarwal 1999, Liu et al. 2002a). The data collectively support the notion that NE differentiation of PCa cells is mediated by multiple signal pathways, in part due to the heterogeneity of PCa cells (Zelivianski et al. 2001), and PCa cells can indeed transdifferentiate into NE-like cells.

Summary

Although several studies have shown interesting results regarding the NE transdifferentiation of PCa cells in cultures as well as in animal models, it remains for further investigations to determine whether those transdifferentiated NE-like cells exhibit the similar biochemical properties as that of clinical NE-like PCa cells. For example, the biochemical information of those transdifferentiated NE-like cells, such as the expression profiles of NE markers, the paracrine effect and the tumorigenic activity, is limited. It is because most of transdifferentiation processes induced by various reagents are transient and cells can fully revert to their original phenotype in the absence of inducers. As such, they are apparently different from those terminally differentiated clinical NE-like PCa cells; nor can they be used for studying the interaction between NE-like PCa and prostatic carcinoma cells. To elucidate the molecular basis of NE differentiation and their roles in PCa progression, stable NE-like PCa cell lines of clinical relevance are imperatively needed.

An NE-like PCa cell model

Establishment and characterization of NE-like LNCaP subclone cells

To investigate the molecular mechanism of NE differentiation and its role in PCa, we established NE-like PCa cell lines by culturing AS LNCaP cells in an androgen-depleted condition, resembling clinical androgen ablation therapy. By prolonged culturing in an androgen-reduced medium, AS LNCaP cells transdifferentiate into NE-like cells and several subclone cells grew to become independent cell lines. These NE-like LNCaP subclone cells exhibit a neuronal morphology, differing from AS LNCaP parental cells (Zhang et al. 2003, Yuan et al. 2006). Importantly, these NE-like LNCaP subclone cells share the same genetic profile with AS LNCaP parental cells (Yuan et al. 2006), corresponding to the observation that NE-like PCa cells in tumor lesions have essentially identical genetic profiles with adjacent exocrine PCa cells (Sauer et al. 2006).

Expression of neuron-specific markers and neuropeptides is one of the major hallmarks of NE-like PCa cells in tumor lesions. Studies in three NE-like LNCaP...
### Table 4 Prostatic neuroendocrine (NE) xenografts and cell models

<table>
<thead>
<tr>
<th>Name</th>
<th>Origin of cells</th>
<th>NE-like morphology</th>
<th>Marker expression</th>
<th>PSA/AR expression</th>
<th>Biological phenotypes</th>
<th>Paracrine (CM) effects</th>
<th>References</th>
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<tr>
<td><strong>Xenograft models</strong></td>
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<tr>
<td>UCRU-Pr-2</td>
<td>Primary NE tumor (human)</td>
<td>ND</td>
<td>NSE/somatostatin β-endorphin/ATCH</td>
<td>No PSA/AR</td>
<td>ND</td>
<td>ND</td>
<td>van Haaften-Day <em>et al.</em> (1987) and Jelbart <em>et al.</em> (1988)</td>
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<tr>
<td>WISH-PC2</td>
<td>Primary tumor (human)</td>
<td>ND</td>
<td>NSE/CgA/synaptophysin/-K8/K18/Bcl-2</td>
<td>No sPSA/PSA/AR</td>
<td>Metastatic activity</td>
<td>High Ki-67 staining</td>
<td>Pinthus <em>et al.</em> (2000)</td>
</tr>
<tr>
<td>LuCaP 49</td>
<td>Metastatic tumor (human)</td>
<td>ND</td>
<td>NSE/synaptophysin/VEGF</td>
<td>No sPSA/AR</td>
<td>75% of cells with Ki-67 staining</td>
<td>ND</td>
<td>True <em>et al.</em> (2002)</td>
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<td><strong>SCC cell lines</strong></td>
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<tr>
<td>SO-MI</td>
<td>Primary NE tumor (human)</td>
<td>Yes</td>
<td>NSE/CgA</td>
<td>No PSA/AR</td>
<td>ND</td>
<td>ND</td>
<td>Okada <em>et al.</em> (2003)</td>
</tr>
<tr>
<td>NE-CS</td>
<td>NE-10 allograft (mouse)</td>
<td>Yes</td>
<td>Secreted serotonin</td>
<td>No AR</td>
<td>Tumor development in nude mice</td>
<td>Up-regulation of cell migration and invasion</td>
<td>Uchida <em>et al.</em> (2005, 2006)</td>
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<tr>
<td>Trans differentiated NE cell line</td>
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</tr>
<tr>
<td>NE-like LNCaP subclone cells</td>
<td>Androgen depleted-LNCaP (C-33) cells</td>
<td>Yes</td>
<td>NSE/CgA/CgB/NT/PTHrP/-K8/K18</td>
<td>No PSA/AR</td>
<td>Tumor development in nude mice</td>
<td>Up-regulation of AI growth, PSA secretion</td>
<td>Yuan <em>et al.</em> (2006)</td>
</tr>
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CM, conditioned medium; SCC, small cell carcinoma; sPSA, secreted PSA; AI, androgen-independent; LNCaP (C-33), androgen-sensitive LNCaP cells; ND, not determined.
subclone cells have revealed that they differentially express elevated levels of multiple NE markers and neuropeptides, including NSE, CgA, CgB, NT, and PTHrP, compared with LNCaP parental cells (Yuan et al. 2006; Table 4). Furthermore, these NE-like LNCaP subclone cells express K8 and K18, two prominent markers for luminal epithelial cells, higher than LNCaP parental cells (Yuan et al. 2006). Similarly, in clinical archival specimens, over 90% of serotonin-positive prostatic NE cells and two cell lines derived from prostatic small cell carcinomas express K18 (Xue et al. 1997, van Bokhoven et al. 2003; Table 4). This observation is in parallel with the expression of PAcP, a prostate secretory epithelium-specific differentiation antigen, in these NE-like LNCaP subclone cells and clinical NE-like PCa cells in prostate carcinomas (Abrahamsson, Yuan et al. 2006). Importantly, these NE-like LNCaP subclone cells do not express AR and PSA (Yuan et al. 2006), similar to those NE-like PCa cells found in the archival specimens (Bonkhoff 2001, Huang et al. 2006). Additionally, these NE-like LNCaP subclone cells express an elevated level of Bcl-2 (Yuan et al. 2006), correlated with the observation in clinical small cell carcinoma cells (Yao et al. 2006). Significantly, even after a 3-month re-culturing in a medium containing androgenic activity, these NE-like LNCaP subclone cells retain the NE phenotype and express NE markers, indicating a terminal transdifferentiation. Collectively, our NE-like LNCaP subclone cells resemble those NE-like PCa cells in clinical samples and are suitable for further biochemical characterizations.

The functional characterizations further reveal the clinical relevance of these NE-like subclone cells. Results of studies in prostatic small cell carcinomas indicate that those clinical NE-like PCa cells are highly aggressive and exhibit a malignant phenotype (Abrahamsson 1999). Corresponding to the clinical observations, our NE-like LNCaP subclone cells exhibit high tumorigenicity. These NE-like subclone cells have a greater tumorigenic activity in xenograft animals and a higher clonogenic activity, but a lower proliferation rate in culture, than AS LNCaP parental cells (Yuan et al. 2006). Importantly, conditioned media (CM) by these NE-like subclone cells, but not by LNCaP parental cells, can stimulate the growth and PSA secretion from LNCaP cells under an androgen-reduced condition (Yuan et al. 2006), corresponding to the observation that clinical NE-like PCa cells have a mitogenic effect on adjacent cancer cells. Thus, the data collectively suggest that these NE-like LNCaP subclone cells, similar to NE-like PCa cells observed in clinical studies, exhibit a malignant phenotype and have a paracrine effect on AI proliferation of PCa cells.

### RPTPα signaling in NE-like LNCaP subclone cells

As described earlier that RPTPα plays a role in androgen depletion-induced NE differentiation, RPTPα is highly expressed in these NE-like LNCaP subclone cells and its expression level is associated with increased ERK/MAPK activity as well as elevated NSE expression, compared with LNCaP parental cells (Fig. 3; Zhang et al. 2003, Yuan et al. 2006). We further compared our NE-like LNCaP subclone cells with NCI-H660 cells, a prostatic small cell carcinoma cell line (Johnson et al. 1989). As shown in Fig. 3, NCI-H660 cells express a high level of RPTPα, even higher than both NE-1.3 and NE-1.8 cells, correlated with activation of ERK/MAPK by phosphorylation. As a control, LNCaP parental cells express a low level of RPTPα and exhibit very low ERK/MAPK activity. Furthermore, NCI-H660 cells, similar to NE-1.3 and NE-1.8 cells, expressed elevated levels of NE markers but no AR and PSA expression (Fig. 3).
These results further support the functional importance of RPTPα in NE differentiation of PCa cells.

Conclusions and prospectives
Accumulated evidence has suggested the roles of NE-like PCa cells in PCa progression and has also revealed that their biological characteristics are different from the normal NE cells. Furthermore, NE-like PCa cells are originated from cancerous luminal epithelial cells, but not from normal NE cells, via a transdifferentiation process. This notion is supported by the fact that those NE-like cells in PCa lesions exhibit the same genetic profile as PCa cells in cancerous lesions but not NE cells in normal tissues both from the same specimen (Sauer et al. 2006). We propose that those NE-like cells in PCa lesions should be defined as ‘NE-like PCa cells’ to accurately reflect their origin. While several factors may induce NE transdifferentiation via multiple signal pathways, the constitutive ERK/MAPK activation apparently plays a critical role in converging multiple signalings for NE differentiation in PCa cells (Fig. 2). Thus, we propose that activation of ERK/MAPK is one of the major mechanisms in the transdifferentiation of PCa cells into NE-like cells. We further propose that sustained, but not transient, hyperactivation of ERK/MAPK is required to support the NE transdifferentiation process of PCa cells, similar to NE differentiation of PC-12 cells (Marshall 1995). This notion is supported by the observations in NCI-H660 and our NE-like stable subclone cells that exhibit a high and sustained level of ERK/MAPK activity (Fig. 3). In parallel, constitutive activation of ERK/MAPK is observed in PCa archival specimens, including advanced hormone-refractory tumors (Gioeli et al. 1999, Price et al. 1999). Therefore, ERK/MAPK activation plays an important role in NE transdifferentiation of PCa cells and may serve as a target for developing new therapeutic approaches for PCa therapy.

It should also be pointed out that NE tumors are rare for PCa, and the post-mitotic NE-like PCa cells in tumor lesions may be one or two steps away from becoming NE tumors. The transition from the post-mitotic NE-like PCa cells to highly aggressive NE tumor cells, such as small cell carcinoma cells, may occur after additional genetic alterations. In parallel, the NE-like LNCaP subclone cells were established after growth silence in SR condition. Interestingly, studies from the TRAMP mouse model as well as the Rb/p53-null mice showed that the simultaneous inactivation or knockout of both p53 and Rb genes promotes NE differentiation of PCa (Perez-Stable et al. 1997, Zhou et al. 2006). While LNCaP cells express the wild-type p53 (Isaacs et al. 1991, Carroll et al. 1993), NCI-H660, and PSK-1, two human prostatic small cell carcinoma cell lines, exhibit truncated and mutated p53 proteins respectively (van Bokhoven et al. 2003). Despite that further investigation on the molecular characterization of those NE-like LNCaP subclone cells is required; it is possible that those NE-like LNCaP subclone cells have acquired additional genetic alterations, such as p53 or Rb mutation/deletion seen in a subpopulation of advanced PCa, which cause the transition from the post-mitotic NE-like PCa cells to highly aggressive NE tumor cells. Thus, our NE-like LNCaP subclone cells provide a useful model to study both NE-like PCa and NE tumor cells.

It is an urgent need to have proper cell lines for studying the biological significance of NE-like PCa cells. Our NE-like LNCaP stable subclone cells acquires many features of clinical NE-like PCa cells, including irreversible NE phenotype, the same genetic profile as the parental PCa cells, expression of multiple NE markers, lack of AR or PSA expression, and high tumorigenicity in xenograft animals. Importantly, the paracrine effect of these NE-like subclone cells on AI proliferation as well as PSA secretion by LNCaP parental cells in SR conditions is in parallel with clinical observations that the proliferation index of cancer cells adjacent to NE-like PCa cells is increased and the circulation level of PSA in a hormone-refractory PCa patient is rebound. In summary, these NE-like LNCaP stable subclone cells can serve as a useful cell model to further explore the molecular mechanism of NE differentiation and its role in PCa for developing targeted therapy.

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References

Abrahamsson PA 1999 Neuroendocrine cells in tumour growth of the prostate. Endocrine-Related Cancer 6 503–519.


Bonkoff H, Stein U & Remberger K 1995 Endocrine-paracrine cell types in the prostate and prostatic adenocarcinoma are postmitotic cells. Human Pathology 26 167–170.


Cox ME, Deeble PD, Bissonette EA & Parsons SJ 2000 Activated 3′,5′-cyclic AMP-dependent protein kinase is sufficient to induce neuroendocrine-like differentiation of the LNCaP prostate tumor cell line. Journal of Biological Chemistry 275 13812–13818.

Deeble PD, Murphy DJ, Parsons SJ & Cox ME 2001 Interleukin-6- and cyclic AMP-mediated signaling potentiates neuroendocrine differentiation of LNCaP prostate tumor cells. Molecular and Cellular Biology 21 8471–8482.


Dube N, Cheng A & Tremblay ML 2004 The role of protein tyrosine phosphatase 1B in Ras signaling. PNAS 101 1834–1839.


Mori S, Murakami-Mori K & Bonavida B 1999 Interleukin-6 induces G1 arrest through induction of p27(Kip1), a cyclin-dependent kinase inhibitor, and neuron-like morphology in LNCaP prostate tumor cells. *Biochemical and Biophysical Research Communications* 257 609–614.


Qiu Y, Robinson D, Pretlow TG & Kung HI 1998 Etk/Bmx, a tyrosine kinase with a pleckstrin-homology domain, is an effector of phosphatidylinositol 3'-kinase and is involved in interleukin 6-induced neuroendocrine differentiation of prostate cancer cells. *PNAS* 95 3644–3649.


Spiotto MT & Chung TD 2000a STAT3 mediates IL-6-induced growth inhibition in the human prostate cancer cell line LNCaP. *Prostate* 42 88–98.


Stork PJ & Schmitt JM 2002 Crosstalk between cAMP and MAP kinase signaling in the regulation of cell proliferation. *Trends in Cell Biology* 12 258–266.

Terry S, Queires L, Gil-Diez-de-Medina S, Chen MW, de la Taille A, Allory Y, Tran PL, Abbou CC, Buttyan R &


