Hydrogen peroxide induces DNA single- and double-strand breaks in thyroid cells and is therefore a potential mutagen for this organ

Natacha Driessens¹, Soetkin Versteyhe¹, Chiraz Ghaddhab¹, Agnès Burniat¹, Xavier De Deken¹, Jacqueline Van Sande¹, Jacques-Emile Dumont¹, Françoise Miot¹ and Bernard Corvilain¹,²

¹Institut de Recherche Interdisciplinaire en Biologie Humaine et Moléculaire and ²Department of Endocrinology of Erasme Hospital, Faculty of Medicine, Université Libre de Bruxelles, Bat C Local C4.145, Campus Erasme, 808, Route de Lennik, B-1070 Brussels, Belgium

(Correspondence should be addressed to N Driessens; Email: nadriess@ulb.ac.be)

Abstract

DNA double-strand breaks (DSBs) are considered as one of the primary causes of cancer but their induction by hydrogen peroxide (H₂O₂) is still controversial. In this work, we studied whether the high levels of H₂O₂ produced in the thyroid to oxidize iodide could induce DNA modifications. Scores of DNA damage, in terms of strand breaks, were obtained by comet assay (alkaline condition for single-strand breaks (SSBs) and neutral condition for DSBs). We demonstrated that in a rat thyroid cell line (PCC13), non-lethal concentrations of H₂O₂ (0.1–0.5 mmol/l) as well as irradiation (1–10 Gy) provoked a large number of SSBs (∼2–3 times control DNA damage values) but also high levels of DSBs (1.2–2.3 times control DNA damage values). We confirmed the generation of DSBs in this cell line and also in human thyroid in primary culture and in pig thyroid slices by measuring phosphorylation of histone H2AX. L-Buthionine-sulfoximine, an agent that depletes cells of glutathione, decreased the threshold to observe H₂O₂-induced DNA damage. Moreover, we showed that DNA breaks induced by H₂O₂ were more slowly repaired than those induced by irradiation. In conclusion, H₂O₂ causes SSBs and DSBs in thyroid cells. DSBs are produced in amounts comparable with those observed after irradiation but with a slower repair. These data support the hypothesis that the generation of H₂O₂ in thyroid could also play a role in mutagenesis particularly in the case of antioxidant defense deficiency.

Endocrine-Related Cancer (2009) 16 845–856

Introduction

Thyroid nodules are common and constitute an important clinical problem. They may occur in up to 50% of a population above 60 years old (Ross 2002). Among these nodules, 5% are cancerous, mainly papillary cancers (PTC). While the prevalence of clinically significant PTC remains relatively low, the prevalence of papillary microcarcinoma is clearly higher and reaches 13% in some series of autopsies (Nasir et al. 2000). There is no clear explanation for this high frequency of thyroid tumors. Iodine deficiency increases the prevalence of hot nodules and multinodular goiters but does not modify the global incidence of thyroid cancers even though it raises the relative proportion of follicular carcinoma (Krohn & Paschke 2002). Irradiation is the only environmental risk factor clearly implicated in thyroid cancer pathogenesis (Ron et al. 1995).

The initial event in the majority of PTC consists of an activation of the RAS/RAF/MEK/MAP kinase pathway, either directly by mutation of BRAF and more rarely of RAS, or indirectly by constitutive activation of tyrosine kinase receptors resulting from chromosomal rearrangements (RET/PTC, TRK; Lacroix et al. 2005). After irradiation, carcinogenic processes are mainly attributed to the formation of
DNA double-strand breaks (DSBs; Sarasin et al. 1999). This damage results from a direct effect of irradiation but also from the generation of reactive oxygen species (ROS) formed during water radiolysis (Mikkelsen & Wardman 2003). However, the precise molecular mechanisms of most radiation-induced cancers are largely unknown. Nevertheless, irradiation is certainly not responsible for the majority of thyroid tumors.

Therefore, since 1990, we hypothesized that the elevated frequency of thyroid tumors (benign or malignant) could be partially explained by the prominent mutagenic environment present in the thyroid, resulting from its metabolism producing large amounts of hydrogen peroxide (H$_2$O$_2$; Song et al. 2007). Indeed, in thyroid, H$_2$O$_2$ generated by the DUOX enzymes (Dupuy et al. 1999, De Deken et al. 2000) is required and is the limiting co-factor for iodide oxidation by thyroperoxidase (TPO) and thyroid hormone synthesis (Nunez & Pommier 1982). Quantitatively, a stimulated thyrocyte generates almost as much H$_2$O$_2$ as a stimulated leukocyte (Corvilain et al. 1994). However, while the leukocyte dies soon after activation, the thyrocyte life is much longer (maximum 7 divisions during adulthood; Coclet et al. 1989) allowing mutations to accumulate. H$_2$O$_2$ could facilitate a mutagenic process and lead to tumorigenesis by altering the DNA (oxidation of bases, DNA single-strand breaks (SSBs) and DSBs). H$_2$O$_2$ could also enhance cell proliferation through various mechanisms (Stone & Yang 2006). Arguments to support the involvement of H$_2$O$_2$ in mutagenesis and etiopathogenesis of thyroid nodules were recently reviewed by our group and others (Maier et al. 2006, Song et al. 2007).

The existence of a more mutagenic environment in the thyroid than other organs are suggested: 1) the spontaneous mutation rate is around 10 times higher in the thyroid than in the liver, 2) the comet assay detects increased levels of oxidized pyrimidine and purine in thyroid compared with other organs, and 3) immunohistochemistry methods reveal higher levels of 8-oxoguanine in the thyroid (Maier et al. 2006). A gene expression signature reflecting the differences in cellular response to $\gamma$-radiation and H$_2$O$_2$ could distinguish radiation-induced (from the Chernobyl Tissue Bank) and spontaneous PTCs (from French patients with no history of radiation exposure; Detours et al. 2007). These data reinforce our hypothesis suggesting that in thyroid, H$_2$O$_2$ could provoke DNA damage and mutations. Nevertheless, harmful effects caused by H$_2$O$_2$ are tightly controlled in thyroid, thanks to the restricted apical localization of its production and the presence of various intracellular H$_2$O$_2$ detoxifying enzymes, like seleno-dependent glutathione peroxidase (GPx; Kohrle et al. 2005). In epidemiological studies, it has been shown that the modification of antioxidant capacities in the diet can modulate cancer risk (Bertram et al. 1987, Ip et al. 1994, Vogt et al. 2003). Antioxidant status of cells may also play a role in the mutagenic potency of irradiation and oxidative stress (Kim et al. 2000, Neumann et al. 2003, Wang et al. 2003).

While carcinogenic DNA DSBs are clearly induced by irradiation, their induction by H$_2$O$_2$ is still controversial. Therefore, we analyzed in this work the capacity of H$_2$O$_2$ to generate DNA SSBs and especially DSBs and compared this with the effects observed with a well-known carcinogenic factor (irradiation) to determine if H$_2$O$_2$, like irradiation, could therefore be implicated in the pathogenesis of thyroid tumors.

Materials and methods

Cell lines and culture conditions

PCCI3 cells, a rat thyroid cell line, were cultured as previously described (Rigutto et al. 2007). Non-transformed rat fibroblasts F208 were grown in DMEM with 10% v/v fetal bovine serum, 1% v/v sodium pyruvate, 100 U/ml penicillin, 100 $\mu$g/ml streptomycin, and 2.5 $\mu$g/ml fungizone. Pig thyroids were obtained from freshly killed animals in accordance to ethical guidelines. Thyroids were cut into thin slices (0.3 mm) of $\sim$50 mg wet weight. Slices were preincubated at 37 °C for 1 h in 2 ml Krebs-Ringer–Hepes (KRH) medium and then transferred to fresh KRH before treatment (Corvilain et al. 2000). Primary cultures of human thyroid cells were prepared and incubated as previously described; the experiments were performed, four days after seeding the thyroid follicles, on differentiated thyrocytes (Roger et al. 1988). Human thyroid tissues were obtained from patients undergoing thyroidectomy for multinodular goiter with approval from the institutional ethic committee.

Cell treatment

Irradiation

Cells were exposed to a single dose of $\gamma$-irradiation from a Cesium$^{137}$ source at a dose rate of 2.03 Gy/min at room temperature. To determine the initial DNA damage, cells were placed on ice immediately after treatment. For the kinetic experiments cells were incubated at 37 °C for different periods of time after irradiation.
**Hydrogen peroxide**

H$_2$O$_2$ (Merck) dilutions were prepared in culture medium immediately before use. The H$_2$O$_2$ solutions were diluted in the medium of dishes containing cells in culture or thyroid slices.

**H$_2$O$_2$ generating system**

A relatively stable production of H$_2$O$_2$ was obtained in the cell culture medium using proline (Sigma–Aldrich) in conjunction with 5 mM/l d-amino-oxidase (DAO from Sigma–Aldrich).

**DL-Buthionine-[S,R]-sulfoximine treatment**

Cells were preincubated overnight with 10 mM/l L-buthionine-sulfoximine (BSO; Sigma–Aldrich) before H$_2$O$_2$ or irradiation treatment.

**Cell survival test**

The cytotoxic potential of the different treatments was evaluated by the 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium (MTS) cell proliferation assay. This colorimetric method measures the formation of a soluble formazan product that is directly proportional to the number of living cells in culture. After different intervals of time after treatment, cells were incubated with a MTS/phenazine methosulfate solution for 1–3 h in the dark at 37 °C and in a 5% CO$_2$ atmosphere as described in the Promega (Promega) technical bulletin no. 169. The soluble formazan product has an absorbance maximum at 490–500 nm and was recorded using an ELISA plate reader. Viability was calculated as a percentage of the control. Triton 0.5% v/v was used as a positive control for cell death.

**H$_2$O$_2$ measurement**

H$_2$O$_2$ was measured in the cell culture medium by a sensitive fluorimetric assay based on the H$_2$O$_2$-dependent oxidation of homovanillic acid (3-methoxy-4-hydroxyphenylactic acid, HVA; Sigma–Aldrich) to a highly fluorescent dimer (2,2'-dihydroxy-3,3'-dimethoxydiphenyl-5,5'-diacetic acid) by HRP (Benard & Brault 1971).

**Iodide organification (protein bound iodide)**

Pig thyroid slices were preincubated for 30 min in KRH buffer containing 0.5 g/l BSA supplemented or not with 0.5 mM/l H$_2$O$_2$. The slices were then incubated in fresh medium supplemented with KI (10$^{-5}$ mol/l) and $^{125}$I (1 μCi/ml) for 30 min. Methimazole (1 mM/l), that blocks iodide organification, added during pre-incubation and incubation, was used to estimate the background in the assay. Iodide organification was measured in basal conditions and in slices stimulated by TSH (10 mU/ml) and ionomycin (2 μmol/l). The slices were homogenized in a methimazole solution (1 mM/l). Proteins were precipitated with 10% v/v trichloroacetic acid and counted. Iodide bound to proteins is expressed as picomoles of iodide organified per 100 mg wet weight tissue/30 min.

**Comet assays**

The comet assay was adapted from Singh et al. (1988) and Olive et al. (1990) as previously described (Chico Galdo et al. 2006). This assay is based on the separation from supercoiled DNA of DNA loops containing strand breaks (SSBs and DSBs) that become free to migrate out of the nucleus towards the anode during an alkaline electrophoresis in a solution of 0.3 M NaOH, 1 mM EDTA, pH > 13. To detect DSBs more specifically, the comet assay was adapted to neutral electrophoresis in a buffer of 300 mM sodium acetate, 100 mM Tris–HCl, pH 8.3 as previously described (Wojewodzka et al. 2002).

DNA images were captured after staining with ethidium bromide (20 μg/ml) with a Zeiss Axioplan 2 imaging microscope with a 40× objective lens. For quantification, the comets were classified into different categories. We subdivided cell DNA damage into five stages (1–5) for the alkaline assay (Collins 2004) and four stages (2–5) for the neutral assay according to the length and the intensity of the comet tail as illustrated in Fig. 1A. Stage 1 (no tail) and stage 2 (halo around the nucleus) corresponded to cells without a significant number of DNA strand breaks. Stages 3–5 corresponded to a gradual increase in DNA damage. We measured the comet score in 200 randomly selected cells per slide. Results were, first, expressed as the percentage of each stage of comets per slide. In a second step, a comet score was calculated, following a modification of Collins’ method, as the sum of the percentage of each comet stage n (from 1 to 5) multiplied by n − 1 (Collins 2004). For this calculation, all the negative comets (stages 1 and 2) were considered as stage 2. The scores were expressed in arbitrary units on a scale from 100 (all the comets are in stage 2) to 400 (all the comets are in stage 5). While the scale of the comet assay was the same in alkaline (reflecting SSBs and DSBs) and neutral condition (more specific for DSBs), the observed score remained semi-quantitative and therefore the values could not be compared to extrapolate the number of SSBs.

Comet immunoassays were performed as previously described (Wojewodzka et al. 2002).
Western blotting and immunocytochemistry

Two or four mg of histone extracts obtained after lysis in buffer (150 mmol/l NaCl, 1 mmol/l EDTA, 20 mmol/l Tris–HCl (pH 8), 0.5% v/v NP40) were separated by 15% SDS/PAGE and transferred to nitrocellulose. Immune complexes were detected with HRP-coupled anti-rabbit or anti-mouse IgG antibodies according to the ECL method (NEN Life science product) as previously described (De Deken et al. 2000). Some nitrocellulose membranes were analyzed by the Odyssey infrared imaging system (LI-COR, Biosciences, Erembodegem, Belgium) using fluorogenic secondary antibodies (IRDye 680 Goat Anti-Mouse and IRDye 800 Goat Anti-Rabbit from LI-COR) as previously described (Picariello et al. 2006).

For immunocytochemistry, treated cells were washed with cold Tris 0.05 mol/l, NaCl 0.15 mol/l pH 7.4 (TBS) and fixed in 95% v/v ethanol – 5% v/v acetic acid. Cells were treated with 3% v/v horse serum/TBS before incubation with primary antibodies against phosphorylated Ser139 of histone H2AX (γH2AX) and total histone H2AX (Upstate Cell Signaling Solutions) (Bioconnect, TE Huiissen, The Netherlands) at a 1:500 dilution in blocking buffer for 1 h at room temperature. After washing with TBS, cells were incubated with 1:400 Cy3-conjugated donkey anti-mouse secondary antibody (Jackson Immuno Research, Suffolk, UK) for 1 h at room temperature in the dark. Cells were washed with TBS and counterstained with bis-benzimide. Observations were performed with a Zeiss Axioplan 2 imaging microscope with a 40× objective lens.

Statistical analysis

Non-parametric unpaired tests were performed with the use of GraphPad Prism Software (San Diego, CA, USA).

Results

Assessment of DNA damage after irradiation in a thyroid cell line

We analyzed DNA damage (SSBs and DSBs) by comet assay immediately after γ-irradiation. Irradiation from 1 to 10 Gy of PCCl3 cells induced a dose-dependent increase in the comet score in alkaline condition (SSBs and DSBs) as well as in neutral condition (DSBs). This damage reached a significant score after irradiation of

Table 1 Score of DNA damage measured by comet assay in PCCl3 cells immediately after irradiation (A) and 15 min after addition of various H2O2 concentrations (B). DNA damage was evaluated in alkaline conditions (SSBs + DSBs) and in neutral conditions (DSBs). Data are expressed as mean (arbitrary units on a scale from 100 to 400) ± S.E.M.; (n), number of measurements. Statistical analyses were made by comparison of control values and values obtained after treatments

<table>
<thead>
<tr>
<th></th>
<th>Alkaline</th>
<th>Neutral</th>
</tr>
</thead>
<tbody>
<tr>
<td>(A) Irradiation</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ctl</td>
<td>120.4±2.1 (32)</td>
<td>113.2±1.9 (26)</td>
</tr>
<tr>
<td>1–2 Gy</td>
<td>165.3±15.4 (6)*</td>
<td>201.0±6.1 (8)*</td>
</tr>
<tr>
<td>4–5 Gy</td>
<td>202.1±15.8 (10)*</td>
<td>244.5±30.0 (2)*</td>
</tr>
<tr>
<td>10 Gy</td>
<td>283.0±6.4 (21)</td>
<td>253.5±8.6 (24)*</td>
</tr>
<tr>
<td>(B) H2O2 (mmol/l)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>0</td>
<td>120.1±2.2 (38)</td>
<td>110.5±1.1 (33)</td>
</tr>
<tr>
<td>0.01</td>
<td>125.7±5.7 (9)</td>
<td>105.8±1.9 (6)</td>
</tr>
<tr>
<td>0.05</td>
<td>128.1±7.2 (10)</td>
<td>132.8±11.2 (4)*</td>
</tr>
<tr>
<td>0.1</td>
<td>245.0±11.7 (19)*</td>
<td>215.6±13.4 (14)*</td>
</tr>
<tr>
<td>0.2</td>
<td>321.8±45.2 (6)*</td>
<td>241.9±23.2 (6)*</td>
</tr>
<tr>
<td>0.5</td>
<td>400.0±0.0 (4)*</td>
<td>225.4±42.3 (4)*</td>
</tr>
<tr>
<td>1</td>
<td>391.3±8.1 (15)*</td>
<td>204.5±9.7 (19)*</td>
</tr>
</tbody>
</table>

*P<0.001. SSBs, single-strand breaks; DSBs, double-strand breaks.
1–2 Gy (Table 1). No significant cell death was observed by the MTS assay after an irradiation of 10 Gy (Table 2). We confirmed the presence of DSBs by analysis of phosphorylation of histone variant H2AX on serine 139 by western blotting. Histone H2AX phosphorylation, that reflects the presence of DSBs, increased in a dose-dependent way from 1 to 10 Gy (Fig. 2A). These results were corroborated by an immunocytochemistry method that detects foci of phosphorylated histone H2AX using the same antibody as in western blotting. A similar relationship was observed between the dose of \( g \)-irradiation and the number of foci (pink spots in the nucleus) per cell as shown in Fig. 2B.

### Assessment of DNA damage after exposure to \( \text{H}_2\text{O}_2 \) in thyroid and non-thyroid cell lines

As \( \text{H}_2\text{O}_2 \) is rapidly degraded when added to the cells, we first estimated how long \( \text{H}_2\text{O}_2 \) was present in the incubation medium of the cells in the described experiments. After addition of \( \text{H}_2\text{O}_2 \), the remaining quantities in the medium were measured at different time intervals in comparison with the same concentration of \( \text{H}_2\text{O}_2 \) added to medium without cells. 0.1 mmol/l \( \text{H}_2\text{O}_2 \) added to PCCl3 cells disappeared rapidly; 23 and 1.6% remained after 15 min and 1 h respectively.

DNA damage in PCCl3 cells was quantified by comet assay 15 min after exposure to \( \text{H}_2\text{O}_2 \). In alkaline conditions, no DNA damage was observed up to 0.05 mmol/l \( \text{H}_2\text{O}_2 \) but the scores increased abruptly and significantly above the control values at 0.1 mmol/l \( \text{H}_2\text{O}_2 \) and reached a maximum value (400) between 0.2 and 0.5 mmol/l \( \text{H}_2\text{O}_2 \) (Table 1) corresponding to the upper limit of the assay. The scores obtained in neutral conditions were significantly increased in cells exposed to a concentration of 0.05 mmol/l \( \text{H}_2\text{O}_2 \) compared to non-treated cells (Table 1). These scores reached a plateau at a \( \text{H}_2\text{O}_2 \) concentration of 0.1 mmol/l. We also examined the phosphorylation of histone H2AX after \( \text{H}_2\text{O}_2 \) treatments (Fig. 2C). One hour after the treatment, we observed a concentration-dependent increase of histone H2AX phosphorylation from 0.1 up till 1 mmol/l \( \text{H}_2\text{O}_2 \). Immunocytochemistry confirmed the

---

### Table 2

Cell survival was evaluated by (A) MTS assay in PCCl3 cells and (B) human thyroid in primary culture for 2, 24, and 48 h after addition of various \( \text{H}_2\text{O}_2 \) concentrations and after different doses of irradiation. Triton 0.5% v/v was used as a positive control for cell death. Viability was calculated as a percentage of the control. Data are expressed as mean \( \pm \) S.E.M.; (\( n \)), number of measurements.

<table>
<thead>
<tr>
<th></th>
<th>2 h</th>
<th>24 h</th>
<th>48 h</th>
</tr>
</thead>
<tbody>
<tr>
<td>(A) ( \text{H}_2\text{O}_2 ) (mmol/l)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0</td>
<td>100.0±1.7 (9)</td>
<td>100.0±0.9 (9)</td>
<td>100.0±2.2 (9)</td>
</tr>
<tr>
<td>0.1</td>
<td>101.4±1.5 (9)</td>
<td>101.9±0.6 (9)</td>
<td>99.5±2.2 (9)</td>
</tr>
<tr>
<td>0.2</td>
<td>100.9±1.1 (9)</td>
<td>100.2±0.8 (9)</td>
<td>95.3±1.3 (9)</td>
</tr>
<tr>
<td>0.5</td>
<td>101.1±1.1 (9)</td>
<td>101.0±1.2 (9)</td>
<td>101.9±1.9 (9)</td>
</tr>
<tr>
<td>1</td>
<td>77.4±2.3 (9)†</td>
<td>87.4±2.0 (9)†</td>
<td>94.9±1.5 (9)</td>
</tr>
<tr>
<td>10</td>
<td>14.2±2.2 (9)†</td>
<td>0.9±0.2 (9)†</td>
<td>0.8±0.2 (9)</td>
</tr>
<tr>
<td>Irradiation (Gy)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>116.7±1.8 (9)</td>
<td>115.8±1.9 (9)</td>
<td>108.0±1.6 (9)</td>
</tr>
<tr>
<td>10</td>
<td>125.1±2.1 (9)</td>
<td>112.7±1.7 (9)</td>
<td>104.2±2.4 (9)</td>
</tr>
<tr>
<td>Triton 0.5% v/v</td>
<td>0.8±0.1 (9)†</td>
<td>0.7±0.1 (9)†</td>
<td>0.5±0.2 (9)†</td>
</tr>
<tr>
<td>(B) ( \text{H}_2\text{O}_2 ) (mmol/l)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0</td>
<td>100.0±1.0 (9)</td>
<td>100.0±0.5 (9)</td>
<td>100.0±1.2 (6)</td>
</tr>
<tr>
<td>0.1</td>
<td>102.2±1.7 (9)</td>
<td>102.2±1.4 (9)</td>
<td>99.8±1.3 (6)</td>
</tr>
<tr>
<td>0.25</td>
<td>100.3±0.9 (9)</td>
<td>99.5±1.5 (9)</td>
<td>101.0±2.7 (6)</td>
</tr>
<tr>
<td>0.5</td>
<td>102.9±2.6 (9)</td>
<td>101.6±2.5 (9)</td>
<td>99.2±2.5 (6)</td>
</tr>
<tr>
<td>1</td>
<td>104.2±2.4 (9)</td>
<td>100.2±2.8 (9)</td>
<td>101.5±2.1 (6)</td>
</tr>
<tr>
<td>10</td>
<td>41.8±8.4 (9)†</td>
<td>53.8±5.2 (9)†</td>
<td>59.3±14.3 (6)*</td>
</tr>
<tr>
<td>100</td>
<td>2.0±1.2 (6)†</td>
<td>2.6±1.0 (6)†</td>
<td>1.2±0.6 (6)†</td>
</tr>
<tr>
<td>Irradiation (Gy)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>106.4±1.0 (9)</td>
<td>101.7±1.2 (9)</td>
<td>102.3±1.7 (6)</td>
</tr>
<tr>
<td>5</td>
<td>112.6±3.3 (9)</td>
<td>105.7±3.1 (9)</td>
<td>105.4±2.0 (6)</td>
</tr>
<tr>
<td>10</td>
<td>104.9±2.3 (9)</td>
<td>104.5±2.8 (9)</td>
<td>104.9±1.1 (6)</td>
</tr>
<tr>
<td>50</td>
<td>106.1±4.3 (9)</td>
<td>101.1±2.5 (9)</td>
<td>104.2±1.7 (6)</td>
</tr>
<tr>
<td>Triton 0.5% v/v</td>
<td>1.2±0.3 (9)†</td>
<td>1.2±0.6 (6)†</td>
<td>0.9±0.4 (6)†</td>
</tr>
</tbody>
</table>

*\( P<0.05; \) †\( P<0.001.\)
presence of a rising number of phosphorylated histone H2AX foci, reflecting the presence of DNA DSBs, in cells treated with 0.1 and 1 mmol/l of H2O2 (Fig. 2D).

H2O2 concentrations equal to or below 0.5 mmol/l did not induce significant cell death in contrast to 10 mmol/l H2O2 that provoked a massive cell death (Table 2). A viability of 77.4% in comparison with the control cells was observed after 2 h treatment with 1 mmol/l H2O2 (Table 2).

Western blotting (γ-H2AX) was also used to evaluate DNA damage in a non-thyroid rat cell line, the F208 fibroblast cell line. We observed the presence of DSBs by the detection of significant levels of phosphorylated H2AX with 0.5–1 mmol/l H2O2 (data not shown).

Assessment of DNA damage after exposure to H2O2 in pig thyroid slices and in human thyroid primary culture cells

DSBs were detected 1 h after 1 and 10 Gy irradiation in pig thyroid slices by measuring H2AX phosphorylation levels; 3.9 ± 1.0-fold (mean ± s.e.m.; P = 0.0063) and 32.0 ± 5.0-fold (mean ± s.e.m.; P < 0.0001) in comparison with the control for 1 and 10 Gy irradiation respectively. We also evaluated the induction of DSBs after incubation with different concentrations of H2O2: 0.1, 0.5, 1, and 10 mmol/l. One hour after treatment, a significant induction of phosphorylated H2AX was observed from 0.5 to 10 mmol/l H2O2: 2.3 ± 0.5-fold (mean ± s.e.m.; P = 0.01), 4.9 ± 1.1-fold (mean ± s.e.m.; P = 0.0056) and 33.1 ± 2.9-fold (mean ± s.e.m.; P < 0.0001) in comparison with the control for 0.5, 1 and 10 mmol/l H2O2 respectively.

Cell survival was indirectly evaluated by the capacity of the pig thyroid slices to organify iodide after treatments with H2O2. In the basal condition, 0.5 mmol/l H2O2 did not change the capacity of pig thyroid slices to organify iodide (protein bound iodide (PB125I) measurements); 97.8 ± 11.2% (mean ± s.e.m.) of the control was measured after H2O2 treatment. In slices stimulated with 10 mU/ml TSH or 2 μM ionomycin (that stimulate H2O2 production), H2O2 did not modify extensively the iodide organifying function; 76.5 ± 5.2% of the control and 92.0 ± 11.4% of the control were measured respectively.

In human thyroid in primary culture, the phosphorylation of histone H2AX was increased following a dose-dependent curve between 1 and 10 Gy (data not shown). No significant cell death was observed by the MTS assay 24 and 48 h after an irradiation of 1 to 50 Gy (Table 2). Human thyroid primary culture cells displayed phosphorylation of histone H2AX one hour after 1 Gy irradiation; 3.9-fold (mean ± s.e.m.; P = 0.01) and 32.0 ± 5.0-fold (mean ± s.e.m.; P < 0.0001) in comparison with the control for 1 and 10 Gy irradiation respectively.

Figure 2 DNA DSBs evaluation using phosphorylation of histone H2AX in PCC13 cells an hour after irradiation (A and B) or addition of H2O2 in the medium (C and D). Phosphorylated (γH2AX) and total histones H2AX were detected with specific antibodies. (A) Western blot showing a dose-dependent effect of irradiation on H2AX phosphorylation. (B) Immunocytochemistry detecting phosphorylated histone H2AX foci (pink spots). (C) Western blot showing dose-dependent effect of H2O2 on H2AX phosphorylation. (D) Immunocytochemistry detecting phosphorylated histone H2AX foci (pink spots). DSBs, double-strand breaks.
after treatment with H₂O₂ from 0.2 to 0.5 mmol/l in a dose-dependent way; 1.9 ± 0.3 fold (mean ± s.e.m.; \( P = 0.0199 \)) and 8.2 ± 3.5 fold (mean ± s.e.m.; \( P = 0.0161 \)) in comparison to the control for 0.2 and 0.5 mmol/l H₂O₂ respectively. One representative experiment is shown in Fig. 3. By MTS assay, we measured that H₂O₂ concentrations equal to or below 1 mmol/l did not induce significant cell death 24 and 48 h after treatment in contrast to 100 mmol/l H₂O₂ that provoked a massive cell death already 2 h after treatment (Table 2).

**Effect of depletion of glutathione on DNA damage induced by H₂O₂ and irradiation in PCCl₃ cells**

BSO irreversibly inhibits γ-glutamylcysteine synthetase (γ-GCS) leading to a decrease of glutathione (GSH) concentrations in the cells. GSH is a cofactor for selenium (Se) dependent GPx involved in the detoxification of most cellular H₂O₂. Comet assays in alkaline conditions were performed on PCCl₃ cells preincubated overnight or not with 10 mmol/l BSO before H₂O₂ or irradiation treatment (Fig. 4A). BSO by itself did not have any effect on DNA integrity. Preincubation with 10 mmol/l BSO rendered cells more sensitive to H₂O₂ in terms of DNA breaks: 0.05 mmol/l H₂O₂ provoked more DNA damage in BSO preincubated cells than in corresponding control cells (score of 188 ± 6 in BSO treated cells compared with 126 ± 6 in cells not preincubated with BSO; \( P < 0.0001 \)). BSO also increased the DNA damage observed at 0.1 mmol/l H₂O₂ (\( P < 0.0001 \)). Significant DNA damage was induced after 2 and 4 Gy irradiation but contrary to what we observed after H₂O₂ treatments, BSO did not increase the DNA damage of irradiated cells (Fig. 4A).

H₂O₂ concentrations measured in the incubation medium increased linearly with the dose of irradiation (2–20 Gy) from 1.0 to 10 μmol/l (data not shown). These concentrations of H₂O₂ are largely below those needed to obtain significant DNA damage.
DNA damage in PCCl3 cells after a combined treatment with H2O2 and irradiation

To evaluate DNA damage induced by a combined H2O2 and irradiation treatment, we measured radiation-induced DNA damage on cells preincubated with a H2O2-generating system constituted by proline and DAO. This H2O2-generating system produced in 1 h from 0.06 to 0.1 mmol/l H2O2 with 10 to 100 mmol/l proline (data not shown).

H2O2 generated by using 50 and 100 mmol/l proline during 1 h induced DNA damage with a score evaluated in alkaline conditions at 282±18 and 356±7 respectively. When cells preincubated with DAO/proline at 50 mmol/l during 1 h were irradiated with 4 Gy, the damage score recorded immediately after irradiation nearly reached the sum of the score observed after DAO/proline (50 mmol/l) treatment alone and after irradiation alone (Fig. 4B). This additive effect was not observed with 100 mmol/l proline due to the saturation of the assay (Fig. 4B).

Repair of DNA damage in PCCl3 cells

Kinetics of DNA repair after irradiation

DNA damage repair at different time intervals after irradiation was analyzed by the comet assay (Fig. 5A and B). In alkaline conditions (Fig. 5A), cells submitted to 10 Gy showed a high score of DNA damage (268±18) immediately after the treatment, rapidly decreased with time to reach a score of 135±17 after 1 h, close to the basal score (116±4). In the neutral comet assay (Fig. 5B), repair of DNA DSBs was also completed 1 h after a 10 Gy treatment (score of 254±25 immediately after treatment compared with 117±7 and 147±12 for respectively control and 1 h repair).

Kinetics of DNA repair after H2O2 treatment

The kinetics of DNA break repair after exposure to H2O2 was also studied by the comet assay performed at different time intervals (Fig. 5C and D). Scores of DNA damage induced by a combined treatment with H2O2 and irradiation were evaluated by the comet assay immediately after irradiation and then after various time intervals. Scores of DNA damage induced by 0.2 mM H2O2 were evaluated 15 min after the addition of H2O2 and then after various time intervals. (A and C) SSBs + DSBs measured in alkaline condition. (B and D) DSBs measured in neutral condition. Data are expressed as mean±S.E.M.; (n), number of measurements for each time interval; *P<0.05; **P<0.01; ***P<0.001; SSBs, single-strand breaks; and DSBs, double-strand breaks.

Figure 5 Kinetics of DNA repair after irradiation (A and B) and H2O2 treatment (C and D) in PCCl3 cells. Scores of DNA damage induced by 10 Gy irradiation were evaluated by the comet assay immediately after irradiation and then after various time intervals. Scores of DNA damage induced by 0.2 mM H2O2 were evaluated 15 min after the addition of H2O2 and then after various time intervals. (A and C) SSBs + DSBs measured in alkaline condition. (B and D) DSBs measured in neutral condition. Data are expressed as mean±S.E.M.; (n), number of measurements for each time interval; *P<0.05; **P<0.01; ***P<0.001; SSBs, single-strand breaks; and DSBs, double-strand breaks.
damage were maximal 15 min after exposure to 0.2 mmol/l \( \text{H}_2\text{O}_2 \). In alkaline conditions, the assay was almost saturated (score around 400). Comet scores decreased by \( \sim 50\% \) after 4 h and reached nearly basal values 6 h after treatment (score 128 \( \pm \) 4; Fig. 5C). The repair of DSBs (comet in neutral conditions) was slower than in irradiated cells; comet scores above control values were still observed 24 h after \( \text{H}_2\text{O}_2 \) treatment (104 \( \pm \) 2 for the control and 124 \( \pm \) 1 after 24 h repair; Fig. 5D).

**Discussion**

The thyroid produces large amounts of \( \text{H}_2\text{O}_2 \) that constitute a potentially mutagenic environment (Bjorkman & Ekholm 1988). The aim of this study was to compare, in a rat thyroid cell line (PCCl3) and in more physiological models (human thyroid primary culture cells and pig thyroid slices), DNA damage induced by a well-known carcinogenic factor (irradiation) with that obtained by a putative carcinogenic agent (\( \text{H}_2\text{O}_2 \)).

The physiological levels of \( \text{H}_2\text{O}_2 \) in cells vary from 0.001 \( \mu \text{mol/l} \) to a maximum of 0.7 \( \mu \text{mol/l} \) (Song et al. 2007) but no data are available for the thyrocyte. As the apparent Km of TPO for \( \text{H}_2\text{O}_2 \) is \( \sim 300 \mu \text{mol/l} \), it has been hypothesized that \( \text{H}_2\text{O}_2 \) reaches especially high concentrations but in a restricted place at the periphery of the thyrocyte, in the postulated thyroxosome (Song et al. 2007). In thyroid, \( \text{H}_2\text{O}_2 \) is produced outside the thyrocyte at its apical pole by DUOX and is used by TPO located in the vicinity of DUOX to oxidize iodide. Therefore, our experimental model in which \( \text{H}_2\text{O}_2 \) is added in the incubation medium mimics the in vivo extracellular production. The range of \( \text{H}_2\text{O}_2 \) concentrations used in this study is comparable to those probably needed to oxidize iodide. It is difficult to estimate \( \text{H}_2\text{O}_2 \) concentrations achieved in the limited space of the follicular lumen in vivo but with the generation of 10 \( \mu \text{mol} \)/100 mg per h they could easily reach the micromolar level (Corvilain et al. 2000).

In our experiments, extracellular \( \text{H}_2\text{O}_2 \) is rapidly reduced and has nearly disappeared after 15 min. At the concentrations used in this study, \( \text{H}_2\text{O}_2 \) was not lethal as confirmed by viability tests. Apoptosis has not been specifically evaluated in this work but previous studies performed on thyroid cells failed to detect apoptosis for similar doses of irradiation and showed apoptosis only in a very small number of cells when treated with similar concentrations of \( \text{H}_2\text{O}_2 \) (Yang et al. 1997, Riou et al. 1999). These previous observations, along with the absence of mortality in \( \text{H}_2\text{O}_2 \) or irradiation treated cells exclude that apoptosis may significantly contribute to the measured comet scores.

Comet assays were used to evaluate DNA strand breaks in individual cells. We studied the effects of \( \text{H}_2\text{O}_2 \) and irradiation on generation of SSBs and DSBs. Interestingly, thyroid cancers occur at doses as low as 0.1 Gy with a linear dose–response curve and the related risk increases by 10 times at 1 Gy (Ron et al. 1995). In our different thyroid models, we observed a significant number of SSBs and DSBs after an irradiation of 1 Gy or more. No differences in sensitivity to irradiation were observed between PCCl3 cells, pig thyroid slices and human thyroid primary culture cells. High levels of DSBs formation were confirmed by estimation of phosphorylation of histone H2AX by western blotting and by immunocytochemistry. Experiments on non-transformed rat fibroblasts (F208) showed a significant production of DSBs with 1 Gy. These results are in keeping with previous data showing that the formation of \( \sim 35 \) DSBs per gray, per cell, and per cell cycle is a constant (Rogakou et al. 1998, Wojewodzka et al. 2002, Takahashi et al. 2005).

DSBs provoked by \( \text{H}_2\text{O}_2 \) are considered to be rare events: 1 DSBs for \( \sim 2000 \) SSBs (Bradley et al. 1979, Takahashi et al. 2005). However, Bradley & Kohn (1979) showed that in mouse leukemia L1210 cells, \( \text{H}_2\text{O}_2 \) induced DSBs with a ratio of DSBs to SSBs comparable with that caused by X-rays. Takahashi & Ohnishi (2005) reviewed one study demonstrating formation of DSBs and histone H2AX phosphorylation by immunocytochemistry in normal human fibroblasts exposed to 0.1 mmol/l \( \text{H}_2\text{O}_2 \) for 2 h (Takahashi et al. 2005).

DSBs are considered to be more carcinogenic than SSBs. We demonstrated in PCCl3 cells that high but non-lethal concentrations of \( \text{H}_2\text{O}_2 \) provoke a large number of SSBs but also as many DSBs as irradiation. In the presence of large amounts of \( \text{H}_2\text{O}_2 \)-induced SSBs, some apparent DSBs could be due to closely spaced SSBs (Bradley et al. 1979). Therefore, we confirmed the presence of real DSBs detected by neutral comet assays by highlighting phosphorylated histone H2AX. In this test, PCCl3 cells demonstrated the same apparent damage caused by \( \text{H}_2\text{O}_2 \) (0.05–0.1 mmol/l) as by irradiation. Data obtained on human thyroid primary culture cells and on pig thyroid slices showed a threshold of respectively 0.2 and 0.5 mmol/l \( \text{H}_2\text{O}_2 \) for the appearance of DSBs. We also observed a significant number of DSBs after a 0.5 mmol/l \( \text{H}_2\text{O}_2 \) treatment of a non-thyroid cell line (F208, a non-transformed rat fibroblast cell line) meaning that many
if not all mammalian cells are sensitive to such concentrations of H$_2$O$_2$. Thus, our work is clearly demonstrated by two different methods an induction of DSBs by H$_2$O$_2$ in a thyroid cell line and, more akin to the in vivo situation, in human thyroid primary cultures and pig thyroid slices. The demonstration of H$_2$O$_2$-induced DNA damage does not necessarily imply a mutagenic role but it can be extrapolated. A difficulty for this extrapolation is that levels of DNA damage acutely achieved in vitro must be compared with lower levels accumulated over years. However, several arguments support such an extrapolation: 1) H$_2$O$_2$, as the well accepted mutagen X-ray, induces DNA DSBs, 2) thyroid in which oxidative DNA damage has been demonstrated in vivo displays a higher level of mutations than liver (Maier et al. 2006), and 3) low levels of Se in serum (i.e. presumably lower activity of Se dependent GPx) constitute a risk factor promoting thyroid cancer development (Duntas 2006).

Comparison of the apparent sensitivity of our different models to extra-cellular H$_2$O$_2$ is probably worthless as it may reflect differences in cell membrane permeability and in antioxidant capacities of the cells. When H$_2$O$_2$ is applied to the exterior of cultured cells, the intracellular concentrations are estimated to be $\sim$10-fold lower than the extra-cellular concentrations (Song et al. 2007). In thyroid, under physiological conditions, a part of the H$_2$O$_2$ not used for thyroglobulin iodination may diffuse into the cells where it is degraded by very efficient antioxidant enzymes like Se dependent GPx. BSO decreases intracellular GSH and therefore the activity of Se dependent GPx. In PCC13 cells exposed to H$_2$O$_2$, the presence of BSO decreased the concentration of H$_2$O$_2$ needed to observe DNA strand breaks to 0.05 mmol/l without affecting the damage induced by irradiation. BSO alone in absence of externally added H$_2$O$_2$ did not increase the level of DNA strand breaks probably because basal H$_2$O$_2$ production in PCC13 cells is not sufficient to induce DNA damage (De Deken et al. 2002). Thyroid destruction in myxoedematous endemic cretinism has been related to impaired H$_2$O$_2$ degradation in stimulated but Se deficient thyroids (Contempre et al. 2004). Therefore, we may extrapolate that in vivo the potential DNA damaging effect of H$_2$O$_2$ will increase in case of deficient antioxidant defense.

Because radiation increases 8-oxoguanine modifications, it was suggested that both radiation and endogenous oxidative stress could synergistically lead to the initiation of thyroid cancer (Riou et al. 1998). We looked therefore for a possible synergic effect of a combined treatment of irradiation and H$_2$O$_2$ on DNA damage. The observed effects of irradiation on cells preincubated with a H$_2$O$_2$-generating system were additive with no synergy whatever the conditions used. The very low H$_2$O$_2$ concentrations measured after irradiation of the culture medium as well as the absence of a potentiating effect of BSO suggest that in PCC13 cells, H$_2$O$_2$ produced through the radiolysis of water is not the main mechanism involved in DNA damage following irradiation.

As it is well known that DNA repair deficiencies are strongly associated with high cancer risk in humans, we compared the kinetics of repair of DNA breaks induced by irradiation and H$_2$O$_2$. DNA damage induced by a 10 Gy irradiation and measured by the comet assay in PCC13 cells was completely repaired after 1 h. The kinetics of repair was clearly made slow for a similar amount of DSBs induced by H$_2$O$_2$. This observation is probably related to different parameters: 1) the very high quantity of SSBs produced by H$_2$O$_2$ could saturate the repair systems. 2) H$_2$O$_2$ may induce DNA damage, but also have direct inhibitory effects on DNA repair. H$_2$O$_2$ at 0.1 mmol/l can inactivate the human DNA mismatch repair system (Chang et al. 2002) and inhibit the repair of certain types of DNA lesions through redox control of ADP ribosylation and unscheduled DNA synthesis (Pero et al. 1990). 3) In any case, the delay in H$_2$O$_2$-induced damage repair is not due to the persistence of H$_2$O$_2$ in the medium as we demonstrated that in our experimental conditions, H$_2$O$_2$ rapidly (15 min) disappeared from the medium.

In conclusion, H$_2$O$_2$ produces DNA damage in the thyroid. Concentrations of H$_2$O$_2$ that cause significant DNA damage are not lethal for the cells and do not modify cell functioning. These observations reinforce the hypothesis that H$_2$O$_2$ is a potential carcinogenic agent in the thyroid. H$_2$O$_2$ induces SSBs but also more mutagenic DSBs in amounts comparable with what is obtained with irradiation. The low repair efficiency of DNA DSBs induced by H$_2$O$_2$ strengthens the possible role of H$_2$O$_2$, generated in the thyroid to oxidize iodide, in thyroid tumorigenesis. Therefore, chronic endogenous exposure of thyroid cells to H$_2$O$_2$ could be a key to explain the high frequency of thyroid tumors and thyroid microcarcinoma, particularly in case of antioxidant defense deficiency as demonstrated by the increase of damage observed in the presence of BSO and suggested in epidemiological studies in case of Se deficiency (Duntas 2006, Kaprara & Krassas 2006).

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.
Funding

This work was supported by Fonds Erasme pour la Recherche Médicale, Télévie, Fonds de la Recherche Scientifique Médicale (FRSM), Actions de Recherches Concertées de la Communauté Française de Belgique (ARC), European Union Contract FP6-36495 (GENRISK-T) and the ICT Impulse program 2006, Brussels Capital Region, Belgium (In Silico Project). XDD: postdoctoral researcher at the Fonds National de la Recherche Scientifique (FRS-FNRS).

Acknowledgements

We thank Claude Massart, Bernadette Bournonville, and Chantal Degraef for their excellent technical assistance.

References

Benard B & Brault J 1971 Production of peroxide in the thyroid. L’Unión Médicale du Canada 100 701–705.
 Duntas LH 2006 The role of selenium in thyroid autoimmunity and cancer. Thyroid 16 455–460.


