Novel gene rearrangements in transformed breast cells identified by high-resolution breakpoint analysis of chromosomal aberrations

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Abstract

Chromosomal copy number alterations and chromosomal rearrangements are frequent mutations in human cancer. Unlike copy number alterations, little is known about the role and occurrence of chromosomal rearrangements in breast cancer. This may be due to the fact that chromosome-based breakpoint analysis is widely restricted to cultured cells. In order to identify gene rearrangements in breast cancer, we studied the chromosomal breakpoints in radiation-transformed epithelial breast cell lines using a high-resolution array-based approach using 1 Mb bacterial artificial chromosome (BAC) arrays. The breakpoints were further narrowed down by fluorescence in situ hybridisation (FISH) with clones from the 32 k BAC library. The analysis of the cell lines B42-11 and B42-16 revealed rearrangements of chromosomes 7, 8, 10 and 12. We identified the genes Has2, Grid1, Ret, Cpm, Tbx3, Tbx5, Tuba1a, Wnt1 and Arf3 within the breakpoint regions. Quantitative RT-PCR showed a deregulated expression of all of these candidate genes except for Tbx5 and Tbx3. This is the first study demonstrating gene rearrangements and their deregulated mRNA expression in radiation-transformed breast cells. Since the gene rearrangements occurred in the transformed and tumourigenic cell lines only, it is likely that these were generated in conjunction with malignant transformation of the epithelial breast cells and therefore might reflect early molecular events in breast carcinogenesis. Initial studies indicate that these gene alterations are also found in sporadic breast cancers.

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Introduction

Chromosome rearrangement is a key mechanism for gene alteration and subsequent deregulation of cellular processes such as proliferation, apoptosis, genome stability, angiogenesis and motility, leading to malignant tumour development (Hanahan & Weinberg 2000, Albertson et al. 2003). Whereas in breast cancer, numerous studies on copy number alterations exist...
(Climent et al. 2007), there is only little knowledge on the role of chromosomal rearrangements in this type of cancer. This might be due to the fact that high-resolution chromosome analysis requires metaphase chromosomes from vital cells, which are not available from archived clinical samples. However, cell culture models are highly informative tools for the investigation of molecular aberrations (Riches et al. 1997, Zitzelsberger et al. 2001, Climent et al. 2007), which moreover can be used for the functional assessment of candidate genes (Neve et al. 2006). This study supports the hypothesis that gene rearrangements observed in radiation-transformed breast cell lines partly mirror those occurring in primary breast cancer. In order to identify such gene rearrangements, we made use of an experimental approach, which combines the detection of chromosomal breakpoints by cytogenetic techniques, high-throughput isolation of aberrant chromosomes using fluorescence-activated chromosome sorting (FACS) and high-resolution mapping of chromosomal breakpoints using 1 Mb bacterial artificial chromosome (BAC) arrays. This approach has already been applied in both cancer and standard clinical cytogenetics (Darai-Ramqvist et al. 2006, Neve et al. 2006, Backx et al. 2007).

In this study, we present a comprehensive analysis of chromosome rearrangements in two radiation-transformed human epithelial breast cell lines showing a tumourigenic phenotype. The identification of rearranged candidate genes by chromosome breakpoint analysis in transformed breast cells and their altered gene expression point to an involvement of these gene rearrangements in the development of breast cancer.

Material and methods

Cell lines

Primary cultures of human mammary epithelial cells were established from surgically removed tissue samples from regions remote from the tumour in a patient with breast cancer presenting for mastectomy (Romanov et al. 2001). The cells were grown in mammary epithelial cell growth medium (MEGM) with additives (Lonza, Basel, Switzerland) and immortalised by transduction with the human telomerase catalytic subunit (hTERT). The hTERT gene was cloned into the retroviral vector (pBABE-puro), and supernatants from the packaging cell line jCRIP were used to transduce the mammary epithelial cells (Wyllie et al. 2000). A cloned cell line B42-htLG was established following selection with puromycin and subsequently exposed to 20 fractions of 2 Gy gamma irradiation and screened for anchorage-independent growth in soft agar (0.3%). In contrast to the irradiated cell line, the parent line B42 does not produce significant numbers of anchorage-independent colonies. Cloned cell lines were established from individual anchorage-independent colonies derived from the irradiated parent cells, from which two cell lines (B42-11 and B42-16) were used for the current experiments. The cell lines were tested for tumourigenicity by s.c. injection into athymic nude mice, which were subsequently monitored for the induction of tumour growth.

Spectral karyotyping

Metaphase preparations were generated from the radiation-transformed cell lines and from the non-tumourigenic parent cell line B42-htLG according to standard procedures (Zitzelsberger et al. 2001). Spectral karyotyping (SKY) analysis was performed as described previously (Zitzelsberger et al. 1999).

Chromosome sorting

Metaphase chromosomes from cell lines B42-11 and B42-16 were prepared according to standard flow sorting protocols (Ferguson-Smith 1997) with optimised buffers (Ng & Carter 2006).

Chromosome sorting was performed on a FACS Vantage (Becton Dickinson, Franklin Lakes, NJ, USA) equipped with two water-cooled lasers (Innova 300 and Innova 70, Coherent, Santa Clara, CA, USA) tuned at multi-line u.v. (330–360 nm) to excite Hoechst 33258 and chromomycin A3 (457 nm). Sorting speed was ~10 chromosomes/s using a 70-μm nozzle tip. For flow karyogram analysis and gating the programme, CellQuest (Becton Dickinson) was used. For each rearrangement, about 500 chromosomes were sorted into PCR tubes.

Amplification of genomic chromosomal DNA

DNA from chromosomes was amplified using the single cell WGA kit (Sigma) according to the manufacturer’s protocol. DNA purity and quantity were determined using a spectrophotometer (NanoDrop Technologies, Wilmington, DE, USA).

Hybridisation of amplified DNA to metaphase spreads

One microgram of WGA amplification product was labelled with digoxigenin by nick translation, hybridised to normal human metaphase spreads and detected using Cy3-conjugated anti-digoxigenin. FISH signals
were analysed using an epifluorescence microscope (AxioVision, Zeiss) and captured using the FISHView software (Applied Spectral Imaging, Neckarhausen, Germany).

**Array painting**

DNA from aberrant chromosomes (M11-1, M16-1 to -5; Table 1) was hybridised onto 1 Mb BAC arrays according to Fiegler et al. (2003) in an automated hybridisation and washing station (HS400, Tecan, Männedorf, Switzerland). Slides were scanned using a GenePix Personal 4100A array scanner (Molecular Devices, Sunnyvale, CA, USA), and feature intensities were extracted using the GenePixPro 6.0 software (Molecular Devices). The datasets were normalised using the MANOR package (Neuvial et al. 2006) and segmented using the DNAcopy package (Venkatraman & Olshen 2007).

**Confirmatory fluorescence in situ hybridisation (FISH) analysis of array-painting results**

DNA from BAC clones spanning or flanking the chromosomal breakpoints was labelled either with biotin or with digoxigenin by nick translation according to standard protocols. Hybridisation to metaphase spreads of the tumorigenic cell lines was performed as previously described (Zitzelsberger et al. 2001).

**Array CGH**

DNA was extracted from fresh cultured cells (B42-11, B42-16 and B42-htLG) using the DNA mini kit (Qiagen). Totally, 450 ng male genomic reference DNA (Promega) and 450 ng cell line DNA were labelled and co-precipitated with 135 µg Cot-1 DNA (Roche) prior to hybridisation. Hybridisation and washing were performed as described above.

**Table 1** Summary of marker chromosomes in cell lines B42-11 and B42-16 detected by spectral karyotyping (SKY) and array painting

<table>
<thead>
<tr>
<th>Cell line</th>
<th>Marker</th>
<th>Cytogenetic description (SKY)</th>
<th>BAC-based description (array painting with 1 Mb BAC clone set)</th>
</tr>
</thead>
<tbody>
<tr>
<td>B42-11</td>
<td>M11-1</td>
<td>der(7)(7pter→q11.1::10q11.2→qter)</td>
<td>der(7)(7pter→RP5-905H7::RP11-38B21→qter)</td>
</tr>
<tr>
<td></td>
<td>M16-1</td>
<td>der(4)(4pter→4q31.3::12q24.2→qter)</td>
<td>der(4)(4pter→RP11-28E24::RP11-8A1→qter)</td>
</tr>
<tr>
<td>B42-16</td>
<td>M16-2</td>
<td>der(8)(8pter→q24.1::10q23.2→qter)</td>
<td>der(8)(8pter→RP11-115M9::RP11-396M20→qter)</td>
</tr>
</tbody>
</table>

Data were normalised and segmented using the R-package MANOR (Neuvial et al. 2006) and DNAcopy (Venkatraman & Olshen 2007).

**Cloning of fusion products using rapid amplification of cDNA ends**

In order to analyse the rearranged sequences of *Has2* and *Grid1*, rapid amplification of cDNA ends (RACE)-PCR (Frohman et al. 1988) was performed. The reactions were carried out according to the manufacturer’s protocols using the 3' and 5' RACE systems for RACEs (Invitrogen). Gene-specific ‘nested’ primers binding within exons 1 and 2 of the gene *Has2* and exon 2 of gene *Grid1* were used for 3' RACE analysis, and primers binding within exons 3 and 4 of the gene *Grid1* and exon 3 of the gene *Has2* were used for 5' RACE analysis.

The amplification products were cloned into a TOPO cloning vector (Invitrogen) according to the manufacturer’s protocol. Transfected clones were checked for inserted DNA using PCR with M13 forward and reverse primers. Clones showing PCR amplicons of the expected size were grown for 14 h at 37 °C in LB medium (ampicillin) followed by a DNA extraction using the Nucleo Plasmid kit (Macherey-Nagel, Düren, Germany). The plasmid DNA was amplified using the BigDye Terminator v3.1 Cycle Sequencing kit (Applied Biosystems, Fosters City, CA, USA) and analysed using an ABI3730 sequencer (Applied Biosystems). DNA sequences were aligned to genomic and cDNA sequences gathered from the Ensembl database using the NTI Vector software (Invitrogen).

**RNA isolation and reverse transcription**

RNA was isolated from ~5×10^4 cells by phase separation using Trizol (Invitrogen) and residual DNA was digested using the Turbo DNA-free kit (Ambion, Austin, TX, USA). Five micrograms of total RNA was
reverse transcribed using the Superscript II kit (Invitrogen), and the resulting cDNA was directly used for RT-PCR.

**Real-time RT-PCR**

We used commercial exon-spanning Taqman probe sets (Applied Biosystems) for the genes *Has2*, *Grid1*, *Arf3*, *Cpm*, *Tbx3*, *Tbx5* and β-actin (reference gene). Reactions (25 μl) were carried out in triplicates and under standard cycling conditions (40 cycles) in an ABI7300 real-time PCR thermocycler (Applied Biosystems) in combination with the analysis software SDS (Applied Biosystems). Probe sets for exons 10–11 and 12–13 of the *Ret* gene were used as previously described (Rhoden et al. 2004).

Relative quantitation was performed using the ΔΔCt method (Livak & Schmittgen 2001). ΔCt values were calculated relative to the Ct of β-actin and Ct values in the non-transformed B42-htLG (calibrator cell line). If genes were not expressed in the calibrator cell line, expressions are given qualitatively (Table 2).

**Validation of gene alterations of Has2, Ret and Grid 1 by FISH analysis of a human breast cancer tissue array**

A human breast cancer tissue microarray (TMA, BRC1501, Pantomics Inc., San Francisco, CA, USA) was hybridised with the following YAC and BAC FISH probes: RP11-115N9/RP11-711N15 (*Has2* gene), RP11-369M20/RP11-711N15 (*Grid1* gene) and CEPH-YAC clones 313F4/214H10/344H4 (*Ret* gene). The TMA comprised 5-μm FFPE sections (in duplicate) of 70 mostly high-grade breast cancers. Hybridisation and analysis of the TMA were performed as described earlier (Unger et al. 2004). For evaluation, fused and separated FISH signals were scored in each cell to detect gene rearrangements (split signals), the wild-type gene (fused signals) or altered copy number (increased or decreased number of FISH signals). FISH signals were counted as split when the distance between the red and the green signal was >10 μm.

**In silico validation of candidate genes in a public available gene expression set**

In order to validate gene expressions of the found candidate genes, a publicly available gene expression set GSE3494 (Miller et al. 2005) was downloaded from the Gene Omnibus website (http://www.ncbi.nlm.nih.gov/geo/). The cel files that resulted from Affymetrix U133A and U133B Genechip hybridisations were normalised separately using the
R-package gcrma (http://www.bioconductor.org/packages/2.0/bioc/html/gcrma.html), combined and clinical data attached. After subsetting data according to clinically meaningful groups (grade, nodal status and estrogen receptor (ER) status) expression values (of candidate genes Has2, Grid1, Ret) were tested group by group for significantly different levels of expression either using Mann–Whitney test in case of testing two groups or using Kruskal–Wallis test in case of testing three groups. The test result was significant when the *P* value was within the 95% confidence interval. Plots of expressions along with test results are given in the Supplementary data file, see section on supplementary data.

**Results**

We have identified chromosomal rearrangements and copy number alterations in the radiation-transformed breast cell lines B42-11 and B42-16 producing small tumours in nude mice (tumour induction in each of the three mice tested per cell line). No tumours developed from cells of the parent cell line B42-htLG in any of the eight nude mice tested indicating a tumourigenic phenotype for the radiation-transformed cell lines. Copy number changes were detected using 1 Mb BAC arrays, whilst the chromosomal breakpoints were fine-mapped and confirmed by FISH and mRNA expression levels of the resulting candidate genes were measured by quantitative RT-PCR. Further, we cloned the rearrangement products of the genes Has2 and Grid1 using RACE.

**Isolation of marker chromosomes**

All markers of the cell lines B42-11 and B42-16 identified by SKY (Fig. 1, cytogenetic description of cell lines is given in Supplementary data, see section on supplementary data, section 2) were successfully isolated by FACS. A cytogenetic description and a schematic illustration of the marker chromosomes are

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**Figure 1** Structural aberrations in the radiation-transformed cell lines B42-11 and B42-16. (A) Marker chromosomes (indicated by red arrows) identified by spectral karyotyping (SKY) in the cell line B42-11 (upper panel) and B42-16 (lower panel). B42-11 shows one marker chromosome M11-1 (mar(7)) and B42-16 marker chromosomes M16-1 (mar(4)), M16-2 (i(8)(q)), M16-3 (mar(10)) and M16-4 (mar(12)). Schematic illustration of chromosomal rearrangements in B42-11 demonstrating a translocation (B) and in B42-16 exhibiting complex exchange aberrations (C). Thin black lines with cytogenetic band designation indicate positions of chromosomal breakpoints. Chromosomal fragments indicated by a red cross are deleted in the cell lines. Blue lines indicate exchange processes between chromosomes at the marked breakpoints. Grey small boxes indicate candidate genes, which may be affected by the chromosomal rearrangement, whilst red arrows indicate elevated mRNA expression. (D) Marker chromosomes of cell lines B42-11 and B42-16 drawn in 500 band resolution according to their ISCN cytogenetic description using the CyDAS online analysis site (http://www.cydas.org/OnlineAnalysis, Hiller B, Bradtke J, Balz H & Rieder H 2005).
given in Table 1. The DNA of sorted chromosomes was amplified and subsequently FISH hybridised to normal human metaphase spreads in order to verify the origin of the sorted chromosomes. Regions of the metaphase chromosomes showing a hybridisation signal represent the chromosomal composition of the isolated marker chromosomes (data not shown).

Array painting and identification of candidate genes

In total, six marker chromosomes were cytogenetically characterised by flow-sorting and hybridised to 1 Mb BAC arrays (array painting). An exemplary illustration of such an analysis is given in Fig. 2, while Table 1 summarises the results of the array-painting analyses. The region in between breakpoint-flanking BAC clones (given in Table 1) identified the candidate genes Has2 (chromosome 8), Grid1, Ret (both chromosome 10), Cpm, Tbx3/5, Wnt1 and Arf3 (chromosome 12). These were further investigated by quantitative RT-PCR in order to detect aberrant mRNA expression levels. All chromosome rearrangements were fine-mapped and verified by FISH with BAC clones (Supplementary data, see section on supplementary data) on metaphase preparations of the cell lines. This analysis also revealed the deletion of a small region comprising the gene Tbx5, what explains the absent mRNA expression of Tbx5 (Table 2) in the cell line B42-16. A schema showing array-painting results and illustrating exchange processes in both radiation-transformed cell lines is given in Fig. 1.

Array CGH analysis

All cell lines have been investigated for copy number changes by array CGH using 1 Mb BAC arrays (Table 3). In total, gains on chromosomes 7 and 8 and a loss on chromosome 10 were identified in
In B42-16, gains on chromosomes 8, 12 and 13 and losses on chromosomes 4, 10 and X were detected. An isochromosome 8q, which was already seen in SKY, became apparent in the cell lines B42-11, B42-16 and the parent cell line B42-htLG.

**Quantitative RT-PCR**

Transcriptional expression of the candidate genes Has2, Grid1, Wnt1, Arf3, Cpm, Tbx3, Tbx5 and Ret was determined by quantitative real-time PCR (Table 2). Ret, which usually is not expressed in epithelial breast cells (Esseghir et al. 2007), was clearly overexpressed in both cell lines, B42-11 and B42-16. Also Has2 and Grid1 were overexpressed in B42-11 and B42-16 relative to B42-htLG. Expression of the gene Wnt1 was not seen at all in B42-11, whereas in B42-16 it was detected. A slight overexpression of Cpm was found in B42-16, whilst in the same cell line Tbx5 was not expressed at all (Table 2).

**Cloning of aberrant gene products**

In order to identify the fusion partners of the rearranged genes Has2 and Grid1, we used RACE PCR followed by sequencing of the resulting amplification products. With respect to the breakpoints of chromosomes 8 and 10, it turned out that a part (exons 3–16) of the gene Grid1 (10q23.1-23.2) was fused to an intronic sequence of the gene Zhx2 (8q24.13), which is located ~1 Mb distal to Has2 (8q24.13). RACE amplification of Has2 revealed a breakpoint between exons 2 and 3 of the gene, but it was not possible to map the fusion sequence to a particular gene. However, neither 5' RACE using gene-specific primers binding within exons 1 and 2 of Has2 nor 3'-RACE using primers specific for sequences of exons 1 and 2 of Grid1 resulted in successful amplification.

**TMA analysis**

The prevalence of rearrangements of the genes Grid1, Has2 and Ret was determined by FISH using breakpoint-specific probes on a TMA comprising 70 mostly high-grade invasive ductal breast cancer cases. Every section (duplicate cores) was scored for separated (rearranged) FISH signals (Fig. 3). Twenty-eight percentage (19 out of 68 cases) of cases showed alteration of Has2 (rearrangement or copy number amplification), 15% (10 out of 67) rearrangement of Grid1 and 13% (8 out of 62) rearrangement of Ret (Table 4).

**In silico validation of candidate genes in a public available gene expression set**

The dataset was stratified according to grade (Bloom-Richardson), nodal status and ER status. Group-wise gene expressions of the genes Ret, Has2 and Grid1 were statistically tested.

Expression of Has2 was significantly increased in grade 3 cases compared with grade 1 and 2 cases (P value: 0.035). Grid1 showed elevated expression in grade 3 compared with low-grade (grade 1 and grade 2 cases combined, P value: 0.03) and in node-positive

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**Table 3 Summary of array CGH results**

<table>
<thead>
<tr>
<th>Chromosome</th>
<th>Region start (clone name)</th>
<th>Region end (clone name)</th>
<th>Region start (bp)</th>
<th>Region end (bp)</th>
<th>Region start (cytogenetic band)</th>
<th>Region end (cytogenetic band)</th>
<th>Size (Mb)</th>
</tr>
</thead>
<tbody>
<tr>
<td>B42-11</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Gains</td>
<td></td>
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<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>7</td>
<td>CTB-164D18</td>
<td>RP4-725G10</td>
<td>178219</td>
<td>56275548</td>
<td>7p22</td>
<td>7p11.2</td>
<td>56.1</td>
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<tr>
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<td>CTD-2115H11</td>
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<td>RP11-218L14</td>
<td>62268206</td>
<td>154713754</td>
<td>Xcen</td>
<td>Xqter</td>
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</table>
grade 1/2 compared with node-negative grade 1/2 cases ($P$ values: 0.007 and 0.03). Expression levels of the $Ret$ gene were significantly higher ($P$ value: 0.04) in ER-positive cases compared with ER-negative cases. Plots of results are given in Supplementary data, see section on supplementary data (section 1).

**Discussion**

A combined approach of high-resolution breakpoint analysis of marker chromosomes, PCR-based characterisation of chromosomal breakpoints and FISH was used to investigate chromosomal rearrangements and changes in expression of resulting candidate genes in two radiation-transformed cell lines (B42-11 and B42-16). A significant prevalence of the alterations which we identified from an *in vitro* model was demonstrated by FISH on a set of clinical breast cancers. Furthermore, we could show by *in silico* expression analysis that the genes $Has2$, $Grid1$ and $Ret$ are differentially expressed in clinically relevant groups of breast cancers.

The parent cell line B42-htLG did not induce tumours in nude mice and showed a stable karyotype 47,XX,i(8)(q). The isochromosome 8q has already been reported to be associated with an immortal phenotype.

**Figure 3** Fluorescence *in situ* hybridisation on a FFPE breast cancer tissue array. FISH probes (BACs RP11-115M9, red and RP11-497O14, green) flanking the chromosomal breakpoint of $Has2$ in B42-16 were hybridised onto FFPE breast cancer tissues. (A) Two polyploid cells of a breast cancer specimen with multiple red FISH signals representing a high level amplification. (B) Two breast cancer cells from which the upper exhibits a rearrangement of $Has2$ (split red and green signals, indicated by white arrow) and the lower shows a normal hybridisation pattern (fused signals). (C) A rearrangement of $Grid1$ is demonstrated by split FISH signals (indicated by white arrow) in a breast cancer cell. (D) A rearrangement of the $ret$ locus is indicated by split FISH signals (indicated by white arrow) in a breast cancer cell.
Table 4 Summary of interphase FISH results on a breast cancer tissue microarray (TMA)

<table>
<thead>
<tr>
<th>Gene</th>
<th>BAC/YAC clones used as DNA probes</th>
<th>Nr. of tumour tissues analysed</th>
<th>Nr. of cases with gene rearrangement (%)</th>
<th>Cases with gene alteration (position on array)</th>
</tr>
</thead>
</table>

M. metastasising tumour: either N1-3 or M1; AR, androgen receptor-positive case; ER, estrogen receptor-positive case; PR, progesterone receptor-positive case; HER, HER2/neu amplification; ID, invasive ductal carcinoma; IL, invasive lobular carcinoma; CIS, ductal carcinoma in situ.

*Three cases showed an amplified HAS2 signal instead of a translocation.

of epithelial breast cells (Caruso et al. 2001). Consequently, the marker chromosomes in the transformed cell lines (Table 1) must have developed during radiation-induced malignant transformation and can be attributed to the tumourigenic phenotype of the cells. The complex nature of the rearrangements of the marker chromosomes M16-3 and M16-4 in the cell line B42-16 could be classified as a translocation between chromosomes 10 and 12 with an additional terminal deletion of 10p and a deletion of the gene Tbx5 on 12q24.2. The lack of expression of Tbx5 (Table 2) supports the idea that copy number alterations often lead to altered gene expression of the affected gene.

Has2 was translocated one megabase distal to the Grid1-containing region on chromosome 10. The breakpoint of Has2 occurred between exon 2 and 3 affecting the largest (putatively cytosolic) domain of the protein. The DNA sequence fused to Has2 could not be ascribed to a particular gene, and therefore, the mechanism of activation in the B42-16 cell line remains unclear. Has2 encodes for a high molecular weight hyaluronan (Itano et al. 1999), which at increased expression levels reduces contact inhibition of cell growth and promotes cell migration and thus contributing to features of a transformed phenotype such as anchorage-independent and invasive growth (Itano et al. 2002, Zoltan-Jones et al. 2003). Consequently, in breast cancer, an elevated Has2 expression was shown to be associated with increased cellular proliferation and an invasive phenotype (Udabage et al. 2005a,b, Cook et al. 2006). In our studied set of primary breast cancers, Has2 was altered in 28% of (TMA), and expression was elevated in the grade 3 tumours of the analysed in silico validation set (Miller et al. 2005). This is in accordance with another study correlating Has2 overexpression with an unfavourable outcome (Tammi et al. 2008). In this context, gene rearrangement of Has2 represents a novel mechanism of its activation and offers considerable potential as a therapeutic target in breast cancer.

The location of the chromosomal breakpoint within the Grid1 gene (between exon 2 and 3) indicates an alteration of the ligand-binding N-terminal domain of the resulting protein. Grid1 belongs to a group of glutamate receptors that were believed to exclusively...
play a role in the central nervous system (CNS). However, it was recently shown that the glutamatergic system is also involved outside the CNS in malignant processes (Rzeski et al. 2002, Haas et al. 2005, Kalariti et al. 2005).

From a clinical perspective rearrangement of Grid1 in a substantial subset of breast cancers and elevated expression in grade 3 and low-grade lymph-node-positive breast cancers makes the gene, which is reported in this context for the first time attractive as a potential tumour marker.

Another candidate gene from this study is the receptor tyrosine kinase (TK) Ret, which was surprisingly overexpressed in both transformed cell lines, although only B42-11 exhibited a rearrangement of the gene. In addition both cell lines expressed the ligand-binding (EC) and TK domains of Ret at almost similar levels, which is remarkable in case of B42-11, since in papillary thyroid carcinomas with RET/PTC rearrangement usually only the TK domain is expressed (Ciampi & Nikiforov 2007). Thus, it is likely that in breast cells Ret is activated by a different mechanism or also an activation of the Ret-EC domain occurs in case of a rearrangement. In a study on mostly Her2/PR-positive breast cancers, it was recently reported that Ret is co-expressed with its co-receptor Gfra1 and its ligand GDNF (Esseghir et al. 2007), and another study demonstrated overexpression of Ret in breast cancer cell lines showing RET-induced anchorage-independent growth (Boulay et al. 2008). Both studies suggest interaction of RET with the ER pathway. This is supported by the finding that Ret expression is associated with positive ER status in the expression set from Miller et al. (2005). With regard to RET as a therapeutic target for TK inhibitors such as sunitinib, activation by rearrangement might be of particular interest. The above-mentioned studies would not pick up Ret expression after rearrangement due to the methodology they have used. Therefore, Ret expression in breast cancer is likely to be underestimated, and usage as a therapeutic target or diagnostic/prognostic marker should be taken into consideration.

Our study demonstrated that the molecular analysis of chromosomal breakpoints in radiation-transformed epithelial breast cell lines is able to reveal novel breast cancer candidate genes and that rearrangement of these lead to altered gene expression. Prevalence of rearrangement and subsequent upregulation of Grid1, Has2 and Ret could be validated in sporadic breast cancer. This provides evidence that genomic alterations and gene rearrangements in particular can be used for the identification of attractive candidates in diagnosis and treatment of breast cancer.

**Supplementary data**

This is linked to the online version of the paper at http://dx.doi.org/10.1677/ERC-09-0065.

**Declaration of interest**

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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