Single photon emission computed tomography imaging for temporal dynamics of thyroidal and salivary radionuclide accumulation in 17-allyamino-17-demothoxygeldanamycin-treated thyroid cancer mouse model

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Abstract

Selective iodide uptake and prolonged iodine retention in the thyroid is the basis for targeted radioiodine therapy for thyroid cancer patients; however, salivary gland dysfunction is the most frequent nonthyroidal complications. In this study, we have used noninvasive single photon emission computed tomography functional imaging to quantify the temporal dynamics of thyroidal and salivary radiiodine accumulation in mice. At 60 min post radionuclide injection, radionuclide accumulation in the salivary gland was generally higher than that in thyroid due to much larger volume of the salivary gland. However, radionuclide accumulation per anatomic unit in the salivary gland was lower than that in thyroid and was comparable among mice of different age and gender. Differently, radionuclide accumulation per anatomic unit in thyroid varied greatly among mice. The extent of thyroidal radioactive accumulation stimulated by a single dose of exogenous bovine TSH (bTSH) in triiodothyronine (T3)-supplemented mice was much less than that in mice received neither bTSH nor T3 (nontreated mice), suggesting that the duration of elevated serum TSH level is important to maximize thyroidal radioactive accumulation. Furthermore, the extent and duration of radioactive accumulation stimulated by bTSH was less in the thyroids of the thyroid-targeted RET/PTC1 (thyroglobulin (Tg)-PTC1) mice bearing thyroid tumors compared with the thyroids in wild-type (WT) mice. Finally, the effect of 17-allyamino-17-demothoxygeldanamycin on increasing thyroidal, but not salivary, radioactive accumulation was validated in both WT mice and Tg-PTC1 preclinical thyroid cancer mouse model.

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Introduction

The primary function of the thyroid gland is to synthesize thyroid hormones, triiodothyronine (T3) and thyroxine (T4), which play important roles in systemic metabolism. Iodine is an essential component of thyroid hormone yet it is frequently not abundant in the diet; therefore, the thyroid gland has evolved an effective way to sequester iodine 20 to 40 times against its concentration gradient from the systemic circulation. Iodide is transported into thyroid follicular cells by the sodium iodide symporter (NIS), an intrinsic membrane glycoprotein expressed on the basolateral...
membrane of the thyroid follicular cells. Once in the thyroid cells, iodine is incorporated into thyroglobulin (Tg) through a process called iodine organification and stored in the follicular lumen of the thyroid. Effective radioiodine accumulation in thyroid tissues, mediated by both active uptake and organification, has allowed the clinical use of radioiodine to detect and/or ablate residual thyroid cancer tissue in patients who have undergone thyroidectomy (see review in Shen et al. (2001)).

Both iodide uptake and organification are mainly stimulated by TSH. To selectively induce radioiodine accumulation in thyroid tissues for effective radioiodine therapy, TSH is elevated either by T₄ withdrawal to induce endogenous hypothyroidism or by exogenous administration of recombinant human TSH (rhTSH) prior to radioiodine administration (see guidelines published by Luster et al. (2008) and Cooper et al. (2009)). However, about 20–30% patients with metastatic thyroid cancer do not benefit from radioiodine therapy due to reduced or absent NIS expression/function (Caillou et al. 1998, Castro et al. 2001, Patel et al. 2002, Ward et al. 2003, Oler & Cerutti 2009). Thus, it is of clinical importance to identify/develop pharmacological reagents that could increase and/or restore radioiodine accumulation in these tumors. As radioiodine accumulation is the net outcome of iodide influx and efflux, strategies to increase radioiodine accumulation include selectively increase thyroidal iodide influx and/or decrease iodide efflux.

Much effort has been made to identify reagents that increase NIS mRNA levels and/or radioiodine accumulation in cultured thyroid cells. Among all agents tested, LY294002, a phosphatidylinositol 3-kinase (PI3K) inhibitor, has been reproducibly shown to increase thyroidal NIS expression (Garcia & Santisteban 2002, Zaballos et al. 2008, Liu et al. unpublished observations 2010) as well as NIS-mediated radioiodide uptake activity (Kogai et al. 2008, Liu et al. unpublished observations 2010). Although thyroidal NIS expression was increased by mitogen-activated protein kinase (MAPK) inhibitors (Knauf et al. 2003, Vadysirisack et al. 2007), it was not accompanied by comparable degree of increase in NIS-mediated radioiodide uptake activity (Vadysirisack et al. 2007, Liu et al. unpublished observations 2010). Histone deacetylase (H-DAC) inhibitor depsipeptide appeared to modestly increase NIS mRNA level as well as radioiodine accumulation in both thyroid and non-thyroid cancer cells (Kitazono et al. 2001). A recent study showed that combination treatment of inhibitors for H-DAC, MAPK, and PI3K increased radioiodine accumulation in several human thyroid cancer cell lines (Hou et al. 2010), but this finding is yet to be confirmed by others. Most importantly, to our knowledge, none of these findings have been validated using immune-competent genetically engineered mice carrying thyroid tumors surrounded by physiologically/pathologically relevant microenvironment.

Our laboratory has previously reported that 17-allylamino-17-demethoxygeldanamycin (17-AAG) increases NIS-mediated radioiodine accumulation by decreasing iodide efflux in NIS-expressing PCCl3 rat thyroid cells and RET/PTC1-transformed PC Cl3 cells (Marsee et al. 2004). This finding was subsequently confirmed by another study showing that 17-AAG increases iodide retention in cultured human anaplastic thyroid cancer cells stably expressing human NIS (Elisei et al. 2006). In this study, we have used noninvasive radionuclide imaging with micro-single photon emission computed tomography (micro-SPECT) to validate the effect of 17-AAG on increasing thyroidal, but not salivary, radioiodine accumulation in wild-type (WT) mice and in our thyroid-targeted RET/PTC1 (Tg-PTC1) thyroid cancer mouse model. Our findings encourage the investigation of 17-AAG derivative pharmacological agents to increase the efficacy of radioiodine therapy in thyroid cancer patients, in addition to their potential roles as novel chemotherapeutic agents.

Materials and methods

Animals and treatment

The study protocol (2009A0118) was approved by our Institutional Animal Care and Use Committee that oversees the responsible use of animals in university research and instructional activities. All research activities were conformed to the statutes of the Animal Welfare Act and the guidelines of the Public Health Service as issued in the Guide for the Care and Use of Laboratory Animals (revised 1996). Mice received Harlan Teklad LM-485 Mouse/Rat sterilizable diet (Harlan Teklad Diets, Madison, WI, USA) containing 2.61 mg iodine/kg diet. Tg-PTC1 mice were generated in the FVB/N background, and all Tg-PTC1 mice developed thyroid tumors by 1 month of age (Jhiang et al. 1996, Cho et al. 1999). For T₃-bovine TSH (bTSH) experiments, mice were supplemented with 12 μg/ml T₃ in their drinking water daily. On day 5, mice received a single i.p. injection of bTSH in 200 μl 1% BSA/PBS and after 16 h the mice were subjected to SPECT imaging or killed for ex vivo γ counting, as described below. The bTSH dosages utilized are reported in the section ‘Results’.
Two different experimental plans were used for 17-AAG (National Cancer Institute: NCI 330507) experiments. We started with the protocol of using T$_3$–bTSH-treated mice i.p. injected with a single dose of 40 mg/kg 17-AAG (the highest dose used in mice reported in NCI Investigator Brochure, 2004) and performed SPECT imaging at 2 h post 17-AAG injection. We later modified the protocol using mice neither treated with T$_3$ nor bTSH (nontreated mice) i.p. injected with a single dose of 30 mg/kg 17-AAG and performed SPECT imaging at 3 h post 17-AAG injection.

In vivo multi-pinhole SPECT imaging

Small animal SPECT imaging was performed by the X-SPECT preclinical platform (Gamma Medica, Northridge, CA, USA) equipped with two gamma cameras, each mounted with a multiple pinhole collimator with 1 mm aperture. After i.p. injection of 100–150 µCi Na$^{123}$I ($t_{1/2}=13.3$ h) or Na$^{99m}$TcO$_4$ ($t_{1/2}=6.01$ h) in saline, mice were anesthetized using isofluorane inhalation and immobilized on the animal bed. For subsequent calibration of radioactivity in target tissues at various times, a 0.25 ml Eppendorf tube containing a known amount of the same radionuclide was included as a decay control (DC). SPECT imaging was acquired with 32 projections, each for 50 s on a 30 mm radius of rotation. SPECT images were acquired at 1, 6, and 24 h post Na$^{123}$I injection or 1 h post Na$^{99m}$TcO$_4$ injection.

Ex vivo $\gamma$ counting

Following T$_3$–bTSH treatment, animals were i.p. injected with 20–40 µCi Na$^{123}$I in saline. Animals were killed at 6 h post Na$^{123}$I injection, and thyroid gland, salivary gland, and liver were harvested and their radioactivity was measured with a $\gamma$ counter (PerkinElmer, Waltham, MA, USA) along with a known activity of Na$^{123}$I as a standard for quantification.

Calculation of thyroidal and salivary radioiodine accumulation

The total injected dose was calculated by subtracting the postinjected syringe counts from the preinjected syringes counts. Using the Amira 3.1 software (Gamma Medica), region of interest (ROI) was manually drawn over the thyroid gland, salivary gland, or DC on $xy$, $yx$, and $xz$ planes, and the total radioactivity in the 3D region defined by the three selected planes was automatically determined. Thyroidal and salivary radioiodine accumulation were calibrated by DC counts at each time point and reported as the percentage of injected dose (%ID).

**Statistical analysis**

As many of the experiments measured thyroidal radioiodine accumulation in mice over time, linear mixed models were used to take account of the dependency of observations within the same animals. To ensure unbiased hypothesis tests, we used a covariance structure estimation method for small samples, which avoids underestimation of experimental error. Two sample $t$-tests were used for simple two group comparisons and Holm’s step down procedure (Holm 1979) was used to adjust for multiple comparisons. $P$ values <0.05 for single comparisons or after adjustment for multiple comparisons were considered as significant.

**Results**

Temporal dynamics of radioiodine accumulation in thyroid and salivary glands evaluated by 3D SPECT functional imaging

Thyroidal iodide accumulation is largely the sum of NIS-mediated iodide uptake along with iodide retention by iodide organification. In comparison, salivary glands accumulate iodide through NIS-mediated iodide uptake but they lack iodide organification. The temporal dynamics of $^{123}$I accumulation in thyroid and salivary glands in WT mice was examined by noninvasive SPECT nuclear imaging (Fig. 1). At 1 h post Na$^{123}$I injection (t1), $^{123}$I accumulation was evident in both thyroid and salivary glands. At 6 h post Na$^{123}$I injection (t6), $^{123}$I accumulated in the thyroid gland was further increased due to continued iodide uptake and organification. However, $^{123}$I accumulated in the salivary gland was decreased due to lack of iodide organification and decreased $^{123}$I in the blood circulation. At 24 h post Na$^{123}$I injection (t24), $^{123}$I accumulation in the thyroid gland continued to increase when normalized by physical decay. Conversely, no $^{123}$I accumulation was detected in the salivary gland at t24, suggesting that $^{123}$I was completely cleared from the blood circulation by excretion or tissue uptake in other tissues such as the thyroid. Consequently, thyroidal radioiodine accumulation at t24 was mainly contributed by iodide retention.

Several mice were subjected to SPECT imaging in the same experimental settings as stated above. The amount of $^{123}$I accumulation in the thyroid (%ID) varied extensively among mice and the degree of $^{123}$I accumulation at t6 was not always near maximum, i.e. to the similar extent seen at t24 (Fig. 2). This variability may be contributed by various sizes of...
folicular lumen, as radioactivity equilibrium is reached faster in follicular lumen with smaller radius (Cantraine & Dewandre 1975). Thyroidal %ID at t1, t6, and t24 for each mice studied is shown in Table 1 (t1: 0.63–6.45%, mean ± 2.07%; t6: 1.59–23.21%, mean ± 8.88%; and t24: 1.48–24.66%, mean ± 11.55%). Albeit our small sample number, mice of 14–15 months of age tended to have lower iodide influx as indicated by thyroidal %ID at t1. Mice of 2–3-months old appeared to have lower iodide retention ability as indicated by lower ratios of t6 or t24 over t1 thyroidal %ID. This may be at least in part contributed by their faster renal clearance rate as shown by a sharper decrease in ¹²³I organification/retention in salivary gland at t6 versus t1 (t6/t1 %ID: 2–3 months: 0.1–0.3, mean=0.22, N=3; > 6 months: 0.73–1.01, mean=0.86, N=4). These findings warrant further study using a larger cohort.

The extent of single dose bTSH-induced thyroidal radioiodine accumulation in T₃-supplemented mice is less than that induced by continuous stimulation of endogenous mouse TSH in nontreated mice

To examine the effect of exogenous bTSH on temporal dynamics of thyroidal radioiodine accumulation in the absence of endogenous mouse TSH (mTSH), endogenous mTSH was suppressed by T₃ supplementation in WT mice. As shown in Fig. 2, thyroidal radioiodine accumulation was diminished in all mice that underwent T₃ supplementation (t1: 0.18–1.06%, mean ± 0.49%; t6: 0.26–0.59%, mean ± 0.41%; t24: 0–0.2%, mean ± 0.06%, N=4), despite extensive variability of thyroidal radioiodine accumulation seen in nontreated mice (N=11). These data confirm that our T₃ supplementation protocol is sufficient to suppress endogenous mTSH-induced thyroidal radioiodine accumulation, regardless of age and gender.

In mice treated with 6, 30, or 60 μg of bTSH at day 5 of T₃ supplementation, thyroidal radioiodine accumulation was increased in a dose-dependent manner (data not shown). Further increase of bTSH to 90 μg did not further increase (P value=0.71) thyroidal radioiodine accumulation as determined by ex vivo γ counting of thyroids at 6 h post Na¹²³I injection (Fig. 3A). As shown in Fig. 2, thyroidal radioiodine accumulation average across time was significantly increased (P value=0.0007) by a single dose of 60 μg bTSH in T₃-supplemented mice (t1: 0.34–1.99%, mean ± 0.93%; t6: 1.22–2.84%, mean ± 2.01%; t24: 1.97–3.36%, mean ± 2.46%, N=5). However, thyroidal %ID in T₃-bTSH-treated mice was much lower (P value=0.027) than that in nontreated mice. This is most likely due to the short half-life of bTSH (<3 h, Guyot et al. 2007), such that a single dose of 60 μg bTSH in T₃-supplemented mice is not sufficient to induce thyroidal radioiodine accumulation comparable to nontreated mice, which had continuous low-level stimulation of endogenous mTSH.
were subjected to SPECT imaging as well. In T3 mice, thyroidal radioiodine accumulation was far less (P value < 0.0017), confirming that T3 supplementation for 5 consecutive days is sufficient to suppress endogenous TSH-induced thyroidal radioiodine accumulation. While one dose of 60 µg bTSH was sufficient to induce some extent of thyroidal radioiodine accumulation, the extent of 60 µg bTSH-induced radioiodine accumulation was almost undetected (P value = 0.001) than that in WT mice (8.9–16.3%, mean = 12.44%, N = 4) without normalization for thyroid gland volume or weight. However, the size of thyroid glands in Tg-PTC1 mice was consistently larger than that in WT mice.

To compare the duration of bTSH-induced NIS activity between WT mice and Tg-PTC1 mice, Na$^{99m}$TcO$_4$, which cannot be organified, was used to evaluate NIS-mediated radionuclide influx at 17 or 41 h post bTSH administration. As shown in Fig. 3B, thyroid %ID of Na$^{99m}$TcO$_4$ was modestly decreased (P value = 0.28) at 41 h compared to 17 h post bTSH administration in WT mice (17 h: 1.27–1.54%, mean = 1.41%; 41 h: 0.23–1.50%, mean = 0.93%, N=3). In comparison, thyroid %ID of Na$^{99m}$TcO$_4$ was significantly decreased (P value = 0.0076) at 41 h in all three Tg-PTC1 mice examined (17 h: 1.7–2.60%, mean = 2.01%; 41 h: 0.00–0.57%, mean = 0.32%, N=3). Indeed, statistical analysis by linear mixed model showed that the difference of NIS activity between 17 and 41 h post bTSH administration for Tg-PTC1 mice was much more evident (P value = 0.014) than that for WT mice. Taken together, thyroid

The extent and duration of thyroidal radioiodine accumulation induced by bTSH is less in Tg-PTC1 mice than that in WT mice

Tg-PTC1 mice can serve as a preclinical animal model to examine factors that modulate radioiodine accumulation in thyroid tumors. We previously reported that thyroidal radioiodine accumulation in Tg-PTC1 mice is decreased compared with WT mice at 6 h post Na$^{123}$I injection when normalized with tissue weight despite of elevated serum TSH level in Tg-PTC1 mice (Cho et al. 1999). This is consistent with the clinical findings that malignant thyroid tumors are typically ‘cold’ nodules. Our pilot study showed that thyroidal radioiodine accumulation was also suppressed by T$_3$ supplementation in Tg-PTC1 mice (data not shown). To compare the responsiveness of WT and Tg-PTC1 mice to exogenous bTSH toward induction of thyroidal radioiodine accumulation, $^{123}$I accumulation at t6 was evaluated ex vivo in T3-supplemented mice injected with a single dose of bTSH. As shown in Fig. 3A, the extent of bTSH-induced thyroidal radioiodine accumulation at t6 in Tg-PTC1 mice (1.6–2.5%, mean = 2.04%, N = 4) was significantly lower (P value = 0.001) than that in WT mice (8.9–16.3%, mean = 12.44%, N = 4) without normalization for thyroid gland volume or weight. However, the size of thyroid glands in Tg-PTC1 mice was consistently larger than that in WT mice.

Table 1 Temporal dynamics of thyroidal and salivary radioiodine accumulation in nontreated wild-type mice

<table>
<thead>
<tr>
<th>Gender</th>
<th>Age (months)</th>
<th>Thyroidal %ID t1</th>
<th>Thyroidal iodide retention ability t6/t1</th>
<th>Salivary %ID t1</th>
<th>Salivary iodide retention ability t6/t1</th>
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<td>M</td>
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<tr>
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<td>6.47</td>
<td>1.7</td>
</tr>
<tr>
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<td>13.62</td>
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<tr>
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<tr>
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<tr>
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<td>1.98</td>
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</tr>
<tr>
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</tr>
<tr>
<td>F</td>
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<td>2.14</td>
<td>10.85</td>
<td>4.89</td>
<td>4.13</td>
</tr>
</tbody>
</table>

Figure 2 Temporal dynamics of thyroidal radioiodine accumulation by one dose of 60 µg bTSH in T3 mice is much less than that induced by continuous endogenous mTSH in nontreated mice. WT mice were given 12 µg/ml T$_3$ in drinking water daily. On day 5, mice were injected with one dose of 60 µg bTSH (T$_3$-60 µg bTSH; N = 5). Na$^{123}$I (100–150 µCi) was administered into mice at 16 h post bTSH injection. SPECT images were taken at 1(t1), 6(t6), and 24 h (t24) post Na$^{123}$I injection. For comparison, T$_3$ mice (N = 4) and nontreated mice (N = 11) were subjected to SPECT imaging as well. In T$_3$ mice, thyroidal radioiodine accumulation was almost undetected (P value = 0.0017), confirming that T$_3$ supplementation for 5 consecutive days is sufficient to suppress endogenous TSH-induced thyroidal radioiodine accumulation. While one dose of 60 µg bTSH was sufficient to induce some extent of thyroidal radioiodine accumulation, the extent of 60 µg bTSH-induced radioiodine accumulation was far less (P value = 0.027) than that of nontreated mice.

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tumor-bearing thyroid glands of Tg-PTC1 mice not only were less responsive to bTSH in the induction of radioiodine accumulation but the duration of NIS induction was also much shorter than normal thyroids in WT mice. It is interesting to note that thyroidal %ID of Na$^{99m}$TcO$_4$ at 17 h post bTSH injection in Tg-PTC1 mice was higher compared to WT mice, suggesting that the number of thyroid follicular cells responsive to bTSH at an early time point in Tg-PTC1 mice is much more than that in WT mice. This phenomenon can be explained by increased proliferation of thyroid follicular cells (Cho et al. 1999) and the much larger size of thyroids observed in Tg-PTC1 mice.

The area under time–activity curve of thyroid in WT mice is markedly increased by 30 mg/kg 17-AAG

None of the T$_3$–bTSH mice injected with a single dose of 40 mg/kg 17-AAG survived during SPECT imaging acquisition (N=3, data not shown), indicating the experimental condition was not tolerable by the mice. We then examined the effect of 30 mg/kg 17-AAG in nontreated mice that had Na$^{123}$I injection at 3 h post 17-AAG injection (see the top panel of Fig. 4). Three experimental trials with identical design were performed to compare temporal radioiodine accumulation between dimethyl sulfoxide (DMSO)- and 17-AAG-treated mice. As shown in Fig. 4, thyroidal radioiodine accumulation at t24 in 17-AAG-treated mice was much higher (P value = 0.15) than DMSO-treated mice in each experimental trial (DMSO versus 17-AAG: 5.42 vs 9.22%; 20.66 vs 65.88%; 8.07 vs 55.05%), despite extensive variations of thyroidal %ID among different experimental trials. Conversely, thyroidal radioiodine accumulation at t1 was not increased (P value = 0.98) by 17-AAG treatment (DMSO versus 17-AAG: 3.58 vs 4.02%; 1.18 vs 0.63%; 3.24 vs 3.44%). These results indicate that 17-AAG markedly increases iodide retention although it has little effect on NIS-mediated iodide influx.

The average of thyroidal %ID at t1, t6, and t24 among three experimental trials as well as the average...
17-AAG had minimal effect on thyroidal iodide influx. However, between DMSO versus 17-AAG-treated mice, suggesting that thyroidal radioiodine accumulation was quite comparable in wild-type (WT) mice is markedly increased by 30 mg/kg 17-AAG as defined by the AUC. Indeed, while the value of AUC varied greatly among experimental trials (DMSO: 224.4 ± 43.3, N = 2; 17-AAG: 442.6 ± 62.7, N = 2), and the fold increase in AUC by 17-AAG in the two experimental trials was 2.21 and 4.14. Thyroidal iodide retention ability in 17-AAG-treated Tg-PTC1 mice (t24/t1 %ID = 2.6 ± 0.6) was much lower than that in WT mice (t24/t1 %ID = 15.7 ± 0.9) at t24, yet the fold increase of iodide retention ability by 17-AAG was comparable between Tg-PTC1 mice and WT mice at t24. Taken together, the effect of 17-AAG on increasing radioiodine accumulation in cultured thyroid cells is validated in WT mice using temporal SPECT imaging. Furthermore, 17-AAG did not appear to have an effect on salivary %ID (data not shown).

The AUC of thyroid tumor in Tg-PTC1 mice is markedly increased by 30 mg/kg 17-AAG

We next examined the effect of 30 mg/kg 17-AAG on thyroidal radioiodine accumulation in thyroid tumor-bearing thyroid glands of Tg-PTC1 mice. While WT mice were tolerant of 30 mg/kg 17-AAG treatment, two out of four Tg-PTC1 mice did not survive through SPECT imaging acquisition. However, as shown in Fig. 5, both of the remaining Tg-PTC1 mice treated with 17-AAG had evident increase in thyroidal radioiodine accumulation compared with mice treated with DMSO at t24 (DMSO versus 17-AAG: 3.50 vs 25.89%; 2.53 vs 19.23%). Similar to WT mice, 17-AAG appeared to increase radioiodine retention rather than radioiodide influx in Tg-PTC1 mice. As shown in Table 2, iodide retention was moderately increased by 17-AAG at t6 (∼1.5-fold), yet it was further increased at t24 (∼4-fold) as calculated by the average ratios of thyroidal %ID at t6 or t24 versus t1 in each experimental trial. The value of AUC was less variable between the two experimental trials in Tg-PTC1 mice (DMSO: 148.8 ± 44.3, N = 2; 17-AAG: 442.6 ± 62.7, N = 2), and the fold increase in AUC by 17-AAG in the two experimental trials was 2.21 and 4.14.

<table>
<thead>
<tr>
<th>Mice</th>
<th>Treatment</th>
<th>t1</th>
<th>t6</th>
<th>t24</th>
<th>6 h/1 h</th>
<th>24 h/1 h</th>
<th>AUC^a</th>
</tr>
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<tbody>
<tr>
<td>WT</td>
<td>DMSO</td>
<td>2.6 ± 1.3</td>
<td>9.3 ± 4.1</td>
<td>11.2 ± 7.9</td>
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<td>17-AAG</td>
<td>DMSO</td>
<td>2.7 ± 1.8</td>
<td>13.0 ± 6.6</td>
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<td>Tg-PTC1</td>
<td>DMSO</td>
<td>7.4 ± 5.6</td>
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<tr>
<td>17-AAG</td>
<td>DMSO</td>
<td>9.2 ± 1.0</td>
<td>16.7 ± 2.9</td>
<td>22.6 ± 4.7</td>
<td>1.9 ± 0.5</td>
<td>2.6 ± 0.6</td>
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^aAUC, area under time–activity curve; it is calculated by \( \int_{t_1}^{t_{24}} f(t) dt \); f(t), thyroidal %ID; t, time (h).
In this study, we quantified temporal radioiodine accumulation in both thyroid and salivary glands modulated by T₃, bTSH, and/or 17-AAG in mice using SPECT functional imaging. By comparing temporal dynamics of radioiodine accumulation between salivary and thyroid glands, we found that administered radioiodine was almost completely cleared from blood circulation after 24 h, and that iodide organification contributes greatly to thyroidal radioiodine accumulation over time. The extent of thyroidal radioiodine accumulation stimulated by a single dose of exogenous bTSH in T₃ mice was much less than that in nontreated mice, justifying the clinical use of multiple doses of rhTSH or the merit of T₄ withdrawal in thyroid cancer patients prior to radioiodine therapy. Furthermore, the extent and duration of radioiodine accumulation stimulated by bTSH was reduced in thyroid tumor-bearing thyroid glands of Tg-PTC1 mice compared with the thyroids in WT mice. Finally, 17-AAG significantly increased thyroidal exposure of radioiodine over time in our Tg-PTC1 preclinical thyroid cancer mouse model, which encourages the study of 17-AAG derivatives not only as novel chemotherapeutic agents to impede tumor progression but also to improve radioiodine therapy in patients with thyroid cancer. Optimally, 17-AAG derivatives could permit those tumors with low radioiodine accumulation to become amenable to radioiodine treatment and allow for those tumors with normal radioiodine accumulation to be affectively treated with lower radioiodine activities.

Salivary gland dysfunction is the most frequent nonthyroidal complications of radioiodine therapy for patients with thyroid cancer (Kloos 2009). A better understanding of the temporal dynamics of radioiodine accumulation in the thyroid and salivary glands may lead to strategies to mitigate radioiodine-induced salivary gland damage without compromising the efficacy of thyroid cancer radioiodine therapy. A recent study showed that the time required to reach maximal accumulation of ⁹⁹mTcO₄⁻ uptake in thyroid glands is shorter than that in salivary glands of C57 BL/6J mice (Franken et al. 2010). This difference could be explained by higher NIS transport activity and/or smaller NIS-expressing epithelial-cell-delimiting compartments of the thyroid, i.e. thyroid colloid versus salivary acini and duct lumens. Similarly, the rate of decrease in ⁹⁹mTcO₄⁻ accumulation is faster in the thyroid than in the salivary gland after the administration of competitive inhibitors (Franken et al. 2010). In this study, we showed that salivary gland %ID was 1.02–5.75-fold higher than thyroid %ID at t₁ (see Table 1), which has been reported by others (Zuckier et al. 2004, Franken et al. 2010) and is anticipated by the larger size of the salivary gland. Indeed, voxel counts in the selected ROI of salivary were 1.93–9.38-fold higher than those of the thyroid at t₁ (data not shown), although functional voxel counts in selected ROI may not exactly reflect anatomic volume of the target tissue. Interestingly, the normalized means (total signal intensity counts divided by voxel counts) of salivary glands at t₁ were lower than those of the thyroid (salivary/thyroid = 0.47–0.94 with a mean of 0.68), indicating that NIS expression/activity per anatomic unit is lower in salivary glands compared to the thyroid gland. Salivary normalized means at t₁ were quite comparable among mice (62–80 with a mean of 72), yet thyroid normalized means at t₁ varied greatly among mice (66–163 with a mean of 113). Thus, NIS activity per anatomic unit in the salivary gland is not changed by gender or age among different mice.

Although rhTSH administration has been shown to be equally effective compared to thyroid hormone withdrawal in preparation of low-risk patients for postoperative thyroid remnant ablation (Pacini et al. 2006, Chianelli et al. 2009, Elisei et al. 2009, 2010). This difference could be explained by higher NIS transport activity and/or smaller NIS-expressing epithelial-cell-delimiting compartments of the thyroid, i.e. thyroid colloid versus salivary acini and duct lumens. Similarly, the rate of decrease in ⁹⁹mTcO₄⁻ accumulation is faster in the thyroid than in the salivary gland after the administration of competitive inhibitors (Franken et al. 2010). In this study, we showed that salivary gland %ID was 1.02–5.75-fold higher than thyroid %ID at t₁ (see Table 1), which has been reported by others (Zuckier et al. 2004, Franken et al. 2010) and is anticipated by the larger size of the salivary gland. Indeed, voxel counts in the selected ROI of salivary were 1.93–9.38-fold higher than those of the thyroid at t₁ (data not shown), although functional voxel counts in selected ROI may not exactly reflect anatomic volume of the target tissue. Interestingly, the normalized means (total signal intensity counts divided by voxel counts) of salivary glands at t₁ were lower than those of the thyroid (salivary/thyroid = 0.47–0.94 with a mean of 0.68), indicating that NIS expression/activity per anatomic unit is lower in salivary glands compared to the thyroid gland. Salivary normalized means at t₁ were quite comparable among mice (62–80 with a mean of 72), yet thyroid normalized means at t₁ varied greatly among mice (66–163 with a mean of 113). Thus, NIS activity per anatomic unit in the salivary gland is not changed by gender or age among different mice.

Although rhTSH administration has been shown to be equally effective compared to thyroid hormone withdrawal in preparation of low-risk patients for postoperative thyroid remnant ablation (Pacini et al. 2006, Chianelli et al. 2009, Elisei et al. 2009,
Tuttle et al. 2010), it remains uncertain whether rhTSH is equally effective in treating high-risk patients with known residual cancer. The level and duration of serum TSH levels required for effective ablation of thyroid remnant could be very different from that for residual cancer and may vary depending on various somatic tumor mutations that affect radioiodine accumulation. Indeed, Pötzi et al. (2006) reported that a higher radioactivity of $^{131}$I is required to achieve comparable radioiodine accumulation in thyroid cancer metastases in patients administered radioiodine after rhTSH versus thyroid hormone withdrawal preparation. This study showed that thyroid radioiodine accumulation in T$_3$-bTSH mice was much lower than that in nontreated mice suggesting that the duration of exposure to TSH is important to promote desirable thyroid radioiodine accumulation. We also showed that thyroid tumorbearing thyroid glands in Tg-PTC1 mice were less responsive to bTSH-induced thyroidal radioiodine accumulation than normal thyroids in WT mice. Taken together, thyroid cancer foci may require higher $^{131}$I activity for treatment as opposed to normal thyroid remnants and longer duration of elevated TSH level may be required to achieve the desirable $^{131}$I activity in thyroid cancer foci. Accordingly, various thyroid cancer mouse models (see review in Knostman et al. (2007) and Burniat et al. (2008)) can be invaluable to examine the concepts of magnitude and duration of serum TSH elevation required to confer sufficient radioiodine accumulation in thyroid tumors of different histologies that are initiated by various genetic events.

Findings using cultured cells do not always translate into preclinical animal models or patients. For example, leptin decreases radioiodine uptake activity in FRTL-5 immortalized rat thyroid cells (Isozaki et al. 2004) and rat thyroid slices (de Oliveira et al. 2007), yet it increases thyroidal radioiodine uptake activity in live rats (de Oliveira et al. 2007). The discordance is most likely explained by tissue microenvironments and/or systemic effects that do not exist in isolated cultured cell system. In this study, we confirmed that 17-AAG increases thyroid exposure to radioiodine over time in WT mice as well as Tg-PTC1 transgenic mice carrying thyroid tumors. The fact that 17-AAG did not increase thyroidal radioiodine accumulation at 11, but did at 16 and 24, indicates that 17-AAG has little effect on NIS-mediated radioiodine influx but mainly acts on iodide retention/organification. This result is consistent with our previous finding that 17-AAG increases radioiodine accumulation by decreasing iodide efflux in cultured cells (Marsee et al. 2004) and that 17-AAG did not increase thyroidal $^{99m}$TcO$_4$ accumulation at either 4 or 27 h post treatment (data not shown). The mechanism underlying the increase in thyroidal iodide retention by 17-AAG is currently unknown. Since heat shock protein 90 (Hsp90) is known for its role as molecule chaperone, it would be interesting to investigate whether 17-AAG acts to impair the secretion of iodinated Tg derivatives.

The following issues need to be addressed before our findings can be translated into clinical trials. Is 17-AAG’s effect on thyroidal radioiodine accumulation mainly mediated by Hsp90 inhibition? Do all Hsp90 inhibitors increase thyroidal radioiodine accumulation? Which Hsp90 inhibitor has the desired pharmacokinetic and pharmacodynamic profiles? What are the optimal dose, duration, and timing of Hsp90 inhibitor treatment to increase the efficacy of radioiodine therapy? Is the magnitude of increased radioiodine accumulation by Hsp90 inhibitors sufficient to meaningfully improve the efficacy of radioiodine therapy? Are there any differences in the effects of Hsp90 inhibitors on thyroid tumors derived from various genetic abnormalities, such as B-RAF$^{V600E}$ mutation? Finally, while not immediately needed for Hsp90 inhibitor clinical application, what are the molecular targets of this treatment in thyroid tumors, and can they be more optimally modulated by other approaches?

**Declaration of interest**

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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**References**

Y-Y Liu et al.: 17-AAG on thyroidal radioiodine accumulation in vivo


