Biallelic inactivation of the SDHC gene in renal carcinoma associated with paraganglioma syndrome type 3

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Abstract

The etiology and pathogenesis of renal cell carcinoma (RCC) are only partially understood. Key findings in hereditary RCC, which may be site specific or a component of a syndrome, have contributed to our current understanding. Important heritable syndromes of RCC are those associated with pheochromocytoma, especially von Hippel–Lindau disease (VHL) associated with germline VHL mutations, and pheochromocytoma and paraganglioma syndrome (PGL) associated with mutations in one of the four genes (SDHA–D) encoding succinate dehydrogenase. A subset of individuals with SDHB and SDHD germline DNA mutations and variants develop RCC. RCC has never been described as a component of SDHC-associated PGL3. The European–American Pheochromocytoma and Paraganglioma Registry comprises 35 registrants with germline SDHC mutations. A new registrant had carotid body tumor (CBT) and his mother had CBT and bilateral RCC. Blood DNA, paragangliomas, and RCCs were analyzed for mutations and loss-of-heterozygosity (LOH) in/flanking SDHC and VHL. The proband with unilateral CBT had a germline SDHC c.3G > A (p.M1I) mutation. His mutation-positive mother had CBT at age 42, clear cell RCC (ccRCC) at age 68, and papillary RCC (pRCC) at age 69. Both paraganglial tumors showed somatic LOH of the SDHC locus. Both ccRCC and pRCC did not have a somatic SDHC mutation but showed LOH for intragenic and flanking markers of the SDHC locus. LOH was also present for the VHL locus. Our findings suggest that RCC is a component of PGL3. Biallelic inactivation of the SDHC gene may represent a new pathway of pathogenesis of syndromic and nonsyndromic RCC, perhaps of both clear cell and papillary histologies.

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Introduction

Pheochromocytoma-related syndromes have contributed to our understanding of the pathogenesis of renal cell carcinomas (RCCs: Pavlovich & Schmidt 2004, Gill et al. 2011b). In particular, two heritable syndromes have been recognized to confer a risk of RCC. von Hippel–Lindau disease (VHL) is associated with clear cell RCC (ccRCC) in about 30% of the affected patients. The other, pheochromocytoma/paraganglioma syndrome (PGL), associated with germline succinate dehydrogenase (SDHB) mutations has been particularly associated with increased risk of RCC.
Renal parenchymal neoplasias are classified as RCC and benign tumors. RCCs include ccRCC, papillary RCC (pRCC), chromophobe RCC, and RCC of unclassified cell type. The most prevalent are ccRCCs occurring in about 65% of parenchymal malignancies followed by pRCCs accounting for 15% of all RCCs (Delahunt & Eble 2004, Grignon et al. 2004, Cohen & McGovern 2005).

The most well-described disorder comprising both ccRCC and pheochromocytoma is VHL, a relatively common autosomal dominant heritable endocrine neoplasia syndrome, caused by germline mutations in the VHL tumor suppressor gene on 3p25–p26 (Lonser et al. 2003). Renal tumors in VHL are always ccRCC and almost always associated with germline-truncating mutations in the VHL gene.

Recently, heritable ccRCC associated with pheochromocytoma and paraganglioma has been described as a consequence of germline heterozygous mutations of the gene-encoding subunit B of the enzyme SDHB by at least two independent groups (Vanharanta et al. 2004, Ricketts et al. 2008). Interestingly, germline variation in SDHB/D has been found to account for 10% of the Cowden heritable cancer syndrome without mutations in its major susceptibility gene PTEN and that those with SDHB/D variants have an increased prevalence of RCC over those with germline PTEN mutations (Ni et al. 2008, 2012).

Succinate dehydrogenase lies at the crossroads of the Krebs tricarboxylic acid cycle and the electron transport chain. SDH catalyzes succinate to fumarate, which is the substrate of fumarate hydratase (FH). Germline FH mutations are associated with hereditary type 2 pRCC (Tomlinson et al. 2002). Site-specific pRCC has been described as a familial disease associated with germline gain-of-function mutations of the MET proto-oncogene encoding a receptor tyrosine kinase (Schmidt et al. 2004). In contrast to FH-associated pRCCs, which are type 2 papillary histology, pRCCs associated with MET mutations are type 2. Chromophobe (sometimes papillary) RCC is a component of Birt–Hogg–Dube syndrome associated with mutations of the BHD (FLCN) gene located on chromosome 17 (Schmidt et al. 2001, Khoo et al. 2002). Other component phenotypes of these latter patients include fibrofolliculomas, lung cysts, and pneumothorax.

As we and others have described that patients with germline mutations of the SDHB gene develop RCC in the setting of hereditary pheochromocytoma/PGL as well as Cowden syndrome, we sought to address the hypothesis that germline mutations in SDHC may also be associated with RCC by interrogating the population-based European–American Pheochromocytoma and Paraganglioma Registry for kidney tumors in patients and family members with germline mutations in the SDHC gene (Neumann et al. 2004, Vanharanta et al. 2004, Ni et al. 2008).

Patients and methods

Patients

As of October 1, 2011, the population-based European–American Pheochromocytoma and Paraganglioma Registry comprised a total of more than 2000 registrants, of which 35 had germline mutations of the SDHC gene, mainly head and neck paragangliomas. Originally, SDHC mutations were only related to head and neck PGL (Schiavi et al. 2005). When it became evident that SDHC mutation carriers might also have a risk of pheochromocytomas and paragangliomas of the abdomen and thorax (Peczkowska et al. 2008), we informed all 35 SDHC mutation-positive patients to undergo clinical screening by magnetic resonance imaging (MRI) of the thorax and abdomen. We were also cognizant to inspect other organs, such as the kidneys.

We obtained written informed consent from both the research participants, the original index registrant (proband), and his mother, in accordance with an approved protocol deriving from our institutions’ Institutional Review Board for Human Research Subjects’ Protection.

Genetic analysis

For purposes of mutation analysis, we obtained from both the registants 10 ml each EDTA-anticoagulated blood and paraffin blocks of the carotid body tumors (CBTs) from both the patients as well as paraffin blocks containing both the mother’s kidney tumors. Genomic DNA was extracted from blood leukocytes according to standard methods. Germline genomic DNA was subjected to PCR-based Sanger sequencing for the genes VHL, RET, SDHB, SDHC, and SDHD as previously performed (Neumann et al. 2002, Schiavi et al. 2005).

Corresponding normal and tumor tissues from the proband’s and mother’s CBTs and both kidney blocks from the mother were dissected using laser capture microdissection so that somatic DNA from the tumor and the germline DNA from the corresponding normal tissue could be isolated. Somatic loss-of-heterozygosity (LOH)/allelic imbalance (AI) analyses were performed for the SDHC locus and the VHL locus on paired normal tumor DNA samples. LOH/AI analysis was performed using eight polymorphic loci.

(Vanharanta et al. 2004, Ricketts et al. 2008, Gill et al. 2011a,b). Renal parenchymal neoplasias are classified as RCC and benign tumors. RCCs include ccRCC, papillary RCC (pRCC), chromophobe RCC, and RCC of unclassified cell type. The most prevalent are ccRCCs occurring in about 65% of parenchymal malignancies followed by pRCCs accounting for 15% of all RCCs (Delahunt & Eble 2004, Grignon et al. 2004, Cohen & McGovern 2005).
markers flanking the gene and one marker inside the SDHC gene (Table 1, Fig. 1A). The nine markers covered a total genetic distance of 10.25 Mb. As a negative control, seven markers flanking the SDHB gene were analyzed as well. Each of the markers has been amplified in a PCR reaction using the Qiagen Multiplex PCR Kit. One of the two primers of each microsatellite marker was labeled with fluorescent dyes at its 5'-end. The PCR amplification was performed for 40 cycles after an initial 15 min denaturation at 95 °C. Each cycle comprised 30 s at 94 °C, 90 s at an annealing temperature of 60 °C, and 60 s at 72 °C for extension. The reaction was terminated with a final 30 min extension period. PCR products were size separated on the MegaBACE 500 DNA Analysis System (Amersham Biosciences) and analyzed using MegaBACE Genetic Profiler v2.2 Software (Amersham Biosciences). LOH was scored as positive when the degree of reduction in allelic signal intensity was > 50% in one of the two alleles of the tumor DNA compared with the peaks from blood DNA. All PCR reactions showing LOH were independently repeated for confirmation.

For VHL, LOH analyses were performed using seven markers, all flanking the gene, two telomeric (D3S3691 and D3S1597) and five centromeric (D3SVHL3, D3S1335, D3SVHL7, D3SVHL8, and D3S3611; Table 2). All these markers together cover 1.66 Mb of DNA (Fig. 1B).

In addition to LOH/AI analysis, we analyzed the tumor-derived DNA from the two paragangliomas and the two kidney carcinomas for potential somatic mutations in all exons of the SDHC gene and the clear cell carcinoma for VHL mutations.

Results

Screening for renal neoplasias in SDHC mutation-positive registrants

Of the 35 total registrants with germline SDHC mutations, all were strictly recommended to undergo

<table>
<thead>
<tr>
<th>Marker</th>
<th>Carotid body tumor (Patient 1 (son))</th>
<th>Carotid body tumor (Patient 2 (mother))</th>
<th>Renal cell carcinoma (patient 2) ccRCC</th>
<th>Renal cell carcinoma (patient 2) pRCC</th>
</tr>
</thead>
<tbody>
<tr>
<td>D1S2707</td>
<td>×</td>
<td>×</td>
<td>√</td>
<td>×</td>
</tr>
<tr>
<td>D1S2771</td>
<td>×</td>
<td>√</td>
<td>√</td>
<td>√</td>
</tr>
<tr>
<td>D1S484</td>
<td>√</td>
<td>√</td>
<td>√</td>
<td>√</td>
</tr>
<tr>
<td>SDHC-CA-2</td>
<td>–</td>
<td>√</td>
<td>×</td>
<td>√</td>
</tr>
<tr>
<td>SDHC-CA-3</td>
<td>√</td>
<td>×</td>
<td>√</td>
<td>√</td>
</tr>
<tr>
<td>SDHC-TETRA</td>
<td>√</td>
<td>√</td>
<td>√</td>
<td>√</td>
</tr>
<tr>
<td>D1S2675</td>
<td>×</td>
<td>×</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>D1S2844</td>
<td>√</td>
<td>×</td>
<td>×</td>
<td>√</td>
</tr>
<tr>
<td>D1S2851</td>
<td>×</td>
<td>–</td>
<td>×</td>
<td>×</td>
</tr>
</tbody>
</table>

√, LOH was detected at the marker; ×, the marker is heterozygous in the germline and somatic LOH was not detected; –, the marker is homozygous in the germline of that patient.

Figure 1 Physical maps and markers of the regions of the genes SDHC and VHL. (A) Physical map of polymorphic markers within and flanking the SDHC gene. Approximate distances between markers are shown in kilobases. (B) Physical map polymorphic markers of the VHL gene. Approximate distances between them are shown in kilobases.
clinical screening by MRI of the thorax and abdomen. Ten of the 35 patients followed these recommendations, and one of the ten was found to have renal tumors. The latter was the registered mother of an index case, who was a 46-year-old male operated at age 40 for a right CBT. Postoperatively, the index case was screened for an underlying mutation, in the setting of genetic counseling; a germline c.3G>A (p.M1I) mutation of SDHC was found. Subsequently, he underwent clinical screening for other paraganglial system tumors, but MRI of the thorax, abdomen, and pelvis did not show abnormalities. Like the proband, the SDHC p.M1I mutation-positive mother was operated on for a CBT at age 42. At age 68, a 6 cm (diameter) right renal mass was detected by MRI (Fig. 2A and B) and removed by nephrectomy. Histopathology revealed a Fuhrman grade 2 clear cell carcinoma (Fig. 2C), stage pT1b, RN0, M0. At age 69, a 2 cm left renal mass was detected by MRI (Fig. 3A and B) and removed by partial nephrectomy. Histopathology revealed a Fuhrman grade 2, type 1 pRCC (Fig. 3C), stage pT1a, RN0, M0.

Germline mutation analysis of other RCC-related predisposition genes

In addition to SDHC, germline genomic DNA of the index case’s mother was subjected to PCR-based Sanger sequencing for the genes VHL, SDHB, SDHC, SDHD, and MET. No other gene, except for SDHC, was found to be mutated in the germline.

Table 2 Loss-of-heterozygosity (LOH) findings at the von Hippel–Lindau disease (VHL) locus in the renal cell carcinomas of the mother VHL

<table>
<thead>
<tr>
<th>Marker</th>
<th>ccRCC</th>
<th>pRCC</th>
</tr>
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<tbody>
<tr>
<td>D3S3691</td>
<td>✔</td>
<td>✔</td>
</tr>
<tr>
<td>D3S1597</td>
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<td>✔</td>
</tr>
<tr>
<td>D3SVHL3</td>
<td>✔</td>
<td>✔</td>
</tr>
<tr>
<td>D3S1335</td>
<td>×</td>
<td>✔</td>
</tr>
<tr>
<td>D3SVHL7</td>
<td>×</td>
<td>✔</td>
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<tr>
<td>d3SVHL8</td>
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<td>–</td>
</tr>
<tr>
<td>D3S3611</td>
<td>✔</td>
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</table>

✔, LOH was detected at the marker; ×, the marker is heterozygous in the germline and somatic LOH was not detected; –, the marker is homozygous in the germline of that patient.

![Figure 2](image1.png)

Figure 2 Clear cell carcinoma of the right kidney from the proband’s mother aged 68 years. (A) Frontal view from MRI. (B) Transverse view from MRI. (C) Histology of the right kidney tumor showing clear cell carcinoma.
LOH/AI analysis

**Carotid body tumors**

The analysis of the nine SDHC-defining polymorphic markers revealed LOH of centromeric and telomeric markers of the SDHC gene in the paragangliomas from the proband and his mother (Fig. 1A). In addition, the proband’s CBT showed clear LOH in the marker within the SDHC gene (SDHC-CA-3; Supplementary Figure 1, see section on supplementary data given at the end of this article). In the CBT of the proband’s mother, LOH was demonstrated in all centromeric markers and in two of three informative telomeric markers. Of note, the intragenic marker SDHC-CA-3 (within intron 4), which underwent LOH in the son’s tumor, retained heterozygosity in the mother’s tumor (Supplementary Figure 2A, see section on supplementary data given at the end of this article). It should be noted that SDHC-CA-2 is only 3 kb upstream of exon 1 and SDHC-TETRA is only 68 kb downstream of exon 6 and both underwent LOH in the mother’s CBT (Fig. 1A).

**Kidney tumors**

We analyzed both renal carcinomas from the proband’s mother, who had a clear cell cancer, followed by a type I papillary carcinoma of the contralateral kidney a year later. Because of the known role of VHL and the suspected role of SDHC in renal carcinomas, we performed LOH analysis of markers within and flanking both SDHC and VHL (Fig. 1), and we also inspected the sequencing chromatogram using tumor DNA as template (Fig. 4). In addition, the pRCC was shown to have somatic LOH of SDHC minimally ranging from D1S2771 to D1S2844, a distance of 9.7 Mb. The ccRCC showed LOH of all three informative SDHC centromeric markers (D1S2707, D1S2771, and D1S484), the intragenic marker (SDHC-CA-3; Supplementary Figure 2B and C, see section on supplementary data given at the end of this article), and one telomeric marker (SDHC-TETRA). LOH of the VHL gene, spanning D3S3691 to D3S3611 (and potentially distally), was evident in the pRCC (Table 2). For the clear cell renal carcinoma, LOH was found in two telomeric markers (D3S3691 and D3S1597) and in two centromeric markers (D3SVHL3 and D3S3611). As a negative control, we showed that the seven markers defining SDHB retained heterozygosity in both renal tumors.

Immunochemistry for SDHB has been performed for both kidney carcinomas and for the paragangliomas of the mother and the son. The paragangliomas of the son and the mother showed negative SDHB.
immunostaining as expected. SDHB immunostaining was negative for the ccRCC as expected but clearly positive in the pRCC, which is so far unexplained. The normal tissue components in the slides of the kidney tumors were positive as expected.

Discussion

RCC is a relatively uncommon disease, but it is the most common type of kidney neoplasia in adults and, recently, has served as a paradigmatic cancer for molecular targeted therapies. Paradoxically, however, the pathogenesis and etiology of RCC are at best partially understood. Major gains of knowledge regarding etiology and pathogenesis have come from hereditary RCC predisposition syndromes, chief of which is VHL disease caused by germline mutations of the VHL gene (Latif et al. 1993, Zbar et al. 1996, Neumann & Zbar 1997, Neumann et al. 1998, Delahunt & Eble 2004, Grignon et al. 2004, Maher et al. 2011). Remarkably, germline mutations of SDHB have been found in patients with ccRCC and RCC with a distinct solid histology (Vanharanta et al. 2004, Ricketts et al. 2008, Gill et al. 2011a,b). Germline mutations of MET or FH have been reported in patients with pRCC, the former type 1 and the latter type 2 (Schmidt et al. 2001, Tomlinson et al. 2002). In the setting of Cowden and Cowden-like syndromes, germline SDHB/SDHD mutations/variants were found to associate with higher prevalence of breast, thyroid, and renal cell cancers (Ni et al. 2008, 2012). Similarly, germline epimutation of KLLN, which shares a bidirectional promoter with PTEN, was associated with Cowden and Cowden-like syndrome and confers a higher risk of breast and renal carcinomas (Bennett et al. 2010). Subsequently, germline KLLN epimutation was shown to be associated with a subset of apparently sporadic ccRCC, with increased somatic KLLN methylation noted in the matched ccRCC tumors (Bennett et al. 2011). Somatic LOH has been identified in the VHL gene in VHL-associated ccRCC and somatic LOH of the SDHB gene in ccRCC from patients with germline SDHB mutations (Vanharanta et al. 2004, Ricketts et al. 2008). Somatic mutations with biallelic inactivation of the VHL gene in ccRCC has been shown by Foster et al. (1994) and Shuin et al. (1994) and somatic gain-of-function mutations of the MET gene in pRCC has also been reported (Schmidt et al. 1997). Thus, we believe that uncovering the molecular pathogenesis of RCC associated with heritable disease will lend useful clues to RCC pathogenesis in general.

In this study, we systematically interrogated our population-based registry for SDHC mutation-positive participants who also had RCC. Of those who took up our surveillance recommendations, one registrant (10%) was found to have metachronous bilateral RCC of clear cell and papillary histologies. Bilateral involvement is a clinical red flag suggesting heritable disease. However, because the RCCs occurred at an age that is no different from the general population, we proceeded to show that biallelic inactivation by somatic deletion of the remaining wild-type allele occurred in both the RCCs. In contrast, SDHB LOH was absent in both RCC tumors. Neither somatic intragenic mutations of SDHC or VHL were found. Only somatic LOH of the VHL locus was found in the pRCC but surprisingly not present in the ccRCC.

Neither the mother’s right kidney ccRCC nor the left-sided type 1 pRCC showed the morphology described for germline SDHB mutation-associated renal carcinomas described by Gill et al. (2011a,b). The SDHB-related renal tumors showed formation into solid nests or in tubules with cuboidal cells, eosinophilic cytoplasm, cytoplasmic inclusions, and indistinct borders, all of which did not appear to be present in these SDHC-related kidney cancers.

Because germline SDHC mutations are relatively uncommon in our population-based registry compared.
with SDSH and SDSHD mutation-positive participants, it is unclear whether further RCC will develop in the registrants with longer follow-up. For example, it was believed that carriers of mutations of the SDHC gene have an exclusive risk for head and neck paragangliomas but also subsequently shown to be at a risk for adrenal pheochromocytoma (Schiavi et al. 2005). Therefore, the number of patients who have been subjected to a complete clinical screening of the paraganglial system including MRI of the abdomen is limited. In fact, only ten out of 35 identified SDHC mutation carriers in the European–American Pheochromocytoma and Paraganglioma Registry had thus radiological imaging of the kidneys.

In summary, we have found, despite small sample sizes in our population-based registry, that 10% of individuals with SDHC gene mutation develop RCC, at least of clear cell and papillary histology, both of which show second allele somatic inactivation by deletion. If this observation can be replicated in other studies, they suggest that RCC may be a new component of SDHC-related PGL. Further, our hypothesis-generating observation suggest that biallelic inactivation of the SDHC gene could be a new molecular pathway for the pathogenesis of RCC and this should be explored in both heritable and sporadic settings.

Supplementary data
This is linked to the online version of the paper at http://dx.doi.org/10.1530/ERC-11-0324.

Declaration of interest
The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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References


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