Placental growth factor supports neuroendocrine tumor growth and predicts disease prognosis in patients

Georg Hilfenhaus1,2, Andreas Göhrig1,2, Ulrich-Frank Pape2, Tabea Neumann2, Henning Jann2, Dietmar Zdunek3, Georg Hess3, Jean Marie Stassen4, Bertram Wiedenmann2, Katharina Detjen1,2, Marianne Pavel2,* and Christian Fischer1,2,*

1Experimental and Clinical Research Center, a joint cooperation between the Charité Medical Faculty and the Max-Delbrück Center for Molecular Medicine, Lindenberger Weg 80, 13125 Berlin, Germany
2Medizinische Klinik mit Schwerpunkt Hepatologie und Gastroenterologie, Charité-Universitätsmedizin Berlin, Campus Virchow-Klinikum, Augustenburger Platz 1, 13353 Berlin, Germany
3Roche Diagnostics, Nonnenwald 2, 82377 Penzberg, Germany
4ThromboGenics NV, Gaston Geenslaan 1, 3001 Leuven, Belgium
*(M Pavel and C Fischer contributed equally to this work)

Correspondence should be addressed to C Fischer
Email christian.fischer@charite.de

Abstract
Placental growth factor (PIGF), a VEGF-homolog implicated in tumor angiogenesis and adaptation to antiangiogenic therapy, is emerging as candidate target in malignancies. Here, we addressed the expression, function, and prognostic value of PIGF in neuroendocrine tumors (NETs). PIGF was determined in NET patients’ sera collected retrospectively (n = 88) and prospectively (n = 87) using Roche-Elecsys and correlated with clinicopathological data. Tumoral PIGF was evaluated by immunohistochemistry, effects of PIGF on proliferation and migration in vitro were assessed using different NET cell lines and effects on tumor growth in vivo in orthotopic xenografts. Circulating and tumoral PIGF was elevated in patients with pancreatic NETs (pNETs) compared with control sera and respective healthy tissue. De novo PIGF expression occurred primarily in the tumor stroma, suggesting paracrine stimulatory circuits. Indeed, PIGF enhanced NET proliferation and migration in vitro and, conversely, neutralizing antibodies to PIGF reduced tumor growth in vivo. Elevated circulating PIGF levels in NET patients correlated with advanced tumor grading and were associated with reduced tumor-related survival in pNETs. Subsequent determinations confirmed and extended our observation of elevated PIGF levels in a prospective cohort of grade 1 and grade 2 pNETs (n = 30) and intestinal NETs (n = 57). In low-grade pNETs, normal circulating PIGF levels were associated with better survival. In intestinal NETs, circulating PIGF above median emerged as an independent prognostic factor for shorter time-to-progression in multivariate analyses. These data assign to PIGF a novel function in the pathobiology of NETs and propose PIGF as a prognostic parameter and therapeutic target.

Key Words
- neuroendocrine tumor
- NET
- placental growth factor
- PIGF
- prognostic biomarker

Endocrine-Related Cancer
(2013) 20, 305–319

http://erc.endocrinology-journals.org
DOI: 10.1530/ERC-12-0223
© 2013 Society for Endocrinology
Copyright © 2013 Society for Endocrinology
Published by Bioscientifica Ltd.
Endocrine-Related Cancer
(2013) 20, 305–319

Downloaded from Bioscientifica.com at 08/01/2019 02:22:34AM
via free access
Introduction

Neuroendocrine tumors (NETs) are rare neoplasms with an estimated annual incidence of 2–5/100 000, and are typically diagnosed at an advanced/metastatic stage of the disease. Despite dissemination however, the clinical course is relatively variable, including periods of stable disease (Baudin 2007, Modlin 2008). Pancreatic NETs (pNETs) and intestinal NETs constitute distinct molecular, pathologic, and clinical subgroups, and consequently different diagnostic and therapeutic algorithms have been developed. Parameters such as tumor size, locoregional or distant metastasis, and Ki67-based grading are currently used to separate prognostic subgroups (Pape et al. 2008, Panzuto et al. 2008). Hence, advanced NETs constitute a malignant disease with unsolved diagnostic and therapeutic medical needs. Recently, two targeted therapies, the tyrosine-kinase inhibitor sunitinib and the mTOR inhibitor everolimus, have been shown to be effective in prolonging progression-free survival in both pNETs and intestinal NETs, and have been approved for treatment of pNETs (Pavel et al. 2011, Raymond et al. 2011, Yao et al. 2011b). Both drugs act at least in part via inhibition of tumor angiogenesis, promising interest in the vascular features of NETs (Alexandraki & Kaltsas 2012). Well-established angiogenic growth factors, such as VEGF-A (Terris et al. 1998) or angiopeptins (Srirajaskanthan et al. 2009, Detjen et al. 2010), are present in NETs. In addition, characteristic neuroendocrine secretion products, including chromogranin A fragments (Corti 2010) and serotonin (Asada et al. 2009), affect angiogenesis and vascular permeability, altogether creating a unique stromal microenvironment.

Placental growth factor (PIGF) is an angiogenic growth factor of the VEGF family, which might be important in NETs for several reasons: PIGF selectively supports pathological angiogenesis, thereby promoting tumor growth, progression, and dissemination (Carmeliet et al. 2001, Fischer et al. 2007). Thus, PIGF expression in tumor tissue and/or circulating PIGF levels correlated with tumor stage, vascularity, metastasis, survival, and recurrence in several human malignancies (Part et al. 2005, Wei et al. 2005, 2009, Fischer et al. 2008, Maae et al. 2012). Moreover, PIGF induction occurred as a result of antiangiogenic therapies in human cancer patients and in mouse models, and (in the latter situation) constituted a functionally relevant mechanism of resistance development (Batchelor et al. 2007, Fischer et al. 2007, 2008, Rini et al. 2008, Willett et al. 2009). Finally, changes in circulating PIGF following treatment initiation are emerging as predictors of therapy response for selected antiangiogenic treatment modalities in clinical trials (Bass et al. 2010). With regard to NETs, preliminary data released from biomarker determinations in the RADIANT-3 trial revealed a small transient reduction of pretreatment circulating PIGF in patients receiving everolimus (ESMO abstract Yao et al. (2011c)). Though preliminary, this observation highlights the demand for a systematic study of PIGF function in NETs.

Mechanistically, PIGF functions as a pleiotropic cytokine that is expressed by and affects a wide range of different cell types within the tumor microenvironment. PIGF binds to Flt1 (VEGFR1), and to neuropilin-1 (NRP1) and -2 (NRP2), but not to the prototype VEGF receptor Flk1 (VEGFR2) (Fischer et al. 2008). Thereby, PIGF stimulates endothelial cell migration, growth and survival, and increases the proliferation of cancer-associated fibroblasts and smooth-muscle cells (Fischer et al. 2008). Moreover, PIGF recruits endothelial and other angiogenesis-competent bone-marrow progenitors as well as tumor-associated macrophages, which promote tumor angiogenesis, growth, and metastasis (Lyden et al. 2001, Fischer et al. 2008). Besides, PIGF also directly affects tumor cells, which express the PIGF receptors Flt1, NRP1, or NRP2 (Fischer et al. 2008, Bagri et al. 2009). Thereby, PIGF can activate AKT and ERK-mediated canonical signaling pathways, ultimately leading to enhanced tumor cell survival, proliferation, migration, and invasiveness (Fischer et al. 2008). Conversely, blocking PIGF using anti-PIGF antibodies emerged as therapeutic strategy to inhibit growth and metastasis in various preclinical tumor models (Fischer et al. 2007, Coenegrachts et al. 2010, Van de Veire et al. 2010, Schmidt et al. 2011, Yao et al. 2011a, Heindryckx et al. 2012).

Here, we addressed the role of PIGF in NETs. The study characterizes PIGF as a growth factor that promotes proliferation and migration of NET cells in vitro and tumor growth in vivo, and furthermore proposes circulating PIGF as a prognostic biomarker.

Materials and methods

Patients and samples

Sera and tissue samples were obtained from individuals with NETs treated at Charité-Universitätsmedizin Berlin,
Department of Gastroenterology, from 1998 to 2012. Tumor staging with TNM classification performed at the time of blood sampling, as well as histopathological diagnosis and grading, obtained from pathology reports, were established according to ENETS guidelines (Rindi et al. 2006, 2007). Clinical parameters were obtained from systematic review of the medical records. Patient informed consent and local ethics committee approval was obtained.

A retrospective cohort of patients with pNETs (n=88) and a prospective cohort including also intestinal NETs (57 intestinal NETs, 30 pNETs; Table 1) were studied. In the retrospective cohort, 65 out of 88 patients were chemotherapy-naïve and five patients were off treatment for at least 3 months before blood sampling. Out of 65, 10 chemotherapy-naïve patients received somatostatin analogs (SSA), one patient had undergone radioreceptor therapy 4 months before blood sampling, and one patient had undergone transarterial chemoembolization (TACE) within a month of blood sampling. Of the pNET patients in the validation cohort, 15 were therapy-naïve, six received SSA, seven had had chemotherapy, one patient had a combination of SSA and chemotherapy, and one patient received everolimus. Out of 57, 33 intestinal NET patients were treated with SSA and one patient with a combination of SSA and everolimus. Furthermore, one patient each received everolimus, IFNα, and the study drug oral angiogenesis inhibitor PTK787/ZK 222584 (PTK/ZK) in the intestinal NET cohort, and one patient received chemotherapy. All intestinal NET patients

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Retrospective cohort (n=88)</th>
<th>Prospective cohort (n=30)</th>
<th>Intestinal NET (n=57)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sex (female/male)</td>
<td>37/51</td>
<td>16/14</td>
<td>32/25</td>
</tr>
<tr>
<td>Age, median (range)</td>
<td>61 (24–84) years</td>
<td>54 (30–82) years</td>
<td>59 (20–78) years</td>
</tr>
<tr>
<td>pNET subtype</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Nonfunctional</td>
<td>82</td>
<td>25</td>
<td></td>
</tr>
<tr>
<td>Gastrinoma</td>
<td>3</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>Insulinoma</td>
<td>1</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>Glucagonoma</td>
<td>1</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>VIPoma</td>
<td>1</td>
<td>2</td>
<td></td>
</tr>
<tr>
<td>Carcinoid syndrome</td>
<td>0</td>
<td>0</td>
<td>41</td>
</tr>
<tr>
<td>Midgut NET subtype</td>
<td></td>
<td></td>
<td>52</td>
</tr>
<tr>
<td>Lower jejunum/ileum</td>
<td></td>
<td></td>
<td>1</td>
</tr>
<tr>
<td>Appendix</td>
<td></td>
<td></td>
<td>4</td>
</tr>
<tr>
<td>Caecum</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>TNM-stage</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Stage I</td>
<td>2</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Stage II (A/B)</td>
<td>2 (0/2)</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Stage III (A/B)</td>
<td>8 (2/6)</td>
<td>3 (0/3)</td>
<td>1 (0/1)</td>
</tr>
<tr>
<td>Stage IV</td>
<td>72</td>
<td>27</td>
<td>56</td>
</tr>
<tr>
<td>Grading</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>G1</td>
<td>11</td>
<td>7</td>
<td>30</td>
</tr>
<tr>
<td>G2</td>
<td>47</td>
<td>23</td>
<td>27</td>
</tr>
<tr>
<td>G3</td>
<td>16</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Ki67 sub-division G2</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2–5%</td>
<td>15</td>
<td>12</td>
<td>20</td>
</tr>
<tr>
<td>&gt;5–10%</td>
<td>14</td>
<td>7</td>
<td>4</td>
</tr>
<tr>
<td>&gt;10–15%</td>
<td>4</td>
<td>3</td>
<td>0</td>
</tr>
<tr>
<td>&gt;15–20%</td>
<td>7</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>PlGF levels (pg/ml)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Median</td>
<td>19.51</td>
<td>16.53</td>
<td>17.57</td>
</tr>
<tr>
<td>Range</td>
<td>7.27–75.09</td>
<td>5.79–44.05</td>
<td>9.80–100.91</td>
</tr>
<tr>
<td>sFlt1 levels (pg/ml)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Median</td>
<td>83.62</td>
<td>85.40</td>
<td>80.40</td>
</tr>
<tr>
<td>Range</td>
<td>32.60–245.60</td>
<td>13.6–159.00</td>
<td>48.10–155.60</td>
</tr>
<tr>
<td>PlGF:sFlt1 ratio</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Median</td>
<td>0.22</td>
<td>0.19</td>
<td>0.23</td>
</tr>
<tr>
<td>Range</td>
<td>0.07–0.99</td>
<td>0.07–1.00</td>
<td>0.13–0.66</td>
</tr>
</tbody>
</table>

Unless otherwise stated values indicate numbers (n) of subjects.
included in time-to-progression (TTP) analyses were chemotherapy-naïve. The time frame of the prospective cohort was May 2009–December 2012, and blood sampling and follow-up visits with imaging studies were performed every 3–6 months. Tumor progression was determined based on multi-phasic computed tomography or magnetic resonance imaging. Healthy controls were blood donors without medical history of malignant disease and consisted of 58 women and 29 men, with a median age of 45 years (range 41–56 years).

Materials
Antibodies were from ReliaTech (Braunschweig, Germany; to PlGF). Secondary antibodies were from Dianova (Hamburg, Germany). Recombinant human PlGF was from R&D Systems (Minneapolis, MN, USA). Everolimus and sunitinib were obtained from Sigma–Aldrich. Neutralizing antibodies to murine PlGF (5D11D4) and to human PlGF (16D3), as well as the IgG1 control antibody (1C8) for use in vivo were supplied by ThromboGenics NV (Leuven, Belgium) (Fischer et al. 2007).

Cell lines and culture
BON cells were a generous gift from C M Townsend (Department of Surgery, University of Texas Medical Branch, Galveston). KRJ-I intestinal NET cells were generated by R Pfargner (Institute of Pathophysiology and Immunology, Medical University of Graz, Austria) (Pfargner et al. 1996) and kindly provided by I Modlin (Department of Surgery, Yale University School of Medicine, New Haven). BON and KRJ-I cells were authenticated by short tandem repeat DNA-typing and confirmed as unique cell lines. QGP-1 cells were from the Health Science Research Resources Bank (Osaka, Japan); H727 cells were from Banca Biologica, Istituto Nazionale per la Ricerca sul Cancro (Genova, Italy).

Determination of PlGF and sFlt1 levels in serum and culture supernatants
Concentrations of PlGF and sFlt1 were determined from frozen serum samples using Elecsys PlGF and sFlt1 immunoassays (Schiettecatte et al. 2010). As circulating PlGF might be increased in patients with ischemic cardiomyopathy (Nakamura et al. 2009), parallel proBNP determinations served as a surrogate marker of myocardial function. Patients with elevated proBNP (>500 pg/ml) were not excluded from the analysis. However, the outcome of the analyses of grading, survival, and TTP in the overall cohorts and subgroups remained unchanged, if these patients were excluded. PlGF levels from cell culture supernatants were measured using Quantikine ELISA kits (R&D Systems) according to the manufacturer.

Immunohistochemical analyses
Cryostat-sections were fixed in 4% paraformaldehyde (PFA). Immunoperoxidase-staining was performed using Vectastain Elite ABC-kit (Vector Laboratories, Wertheim-Bettingen, Germany) and AEC as substrate chromogen (DAKO, Hamburg, Germany). The antibody to PlGF was diluted 1:100, and was omitted in negative controls. For a semi-quantitative evaluation of immunoreactivity, the immunoreactive area was determined relative to the total field measured using AxioVision (Zeiss, Jena, Germany).

Growth assays
In 24-well dishes, 10⁵ cells/well were plated and allowed to attach for 6 h. Following stimulation with PlGF for 48 h, cell numbers were counted using a Neubauer hemocytometer.

Migration assays
A number of 2 × 10⁵ cells/insert were placed in serum-free medium in the upper well of a chemotaxis chamber and allowed to migrate for 20 h toward PlGF added to the bottom well. Migrated cells were stained and quantified by counting 12 standardized fields at 200× magnification (refer to Supplementary Methods, see section on supplementary data given at the end of this article for details).

BON NET orthotopic tumor model
Female NMRI⁰⁰⁰ mice (21–25 g) were from Taconics (Bomholt, Denmark). Animal care followed institutional guidelines and experiments were approved by local animal research authorities. Mice were anesthetized by i.p. administration of Ketanest (100 mg/kg) and Rompun (10 mg/kg). For tumor induction, the pancreas was exposed and 1 × 10⁶ BON cells were injected into the head of the pancreas as previously described (Detjen et al. 2010). Treatment of engrafted tumors was initiated at week 3 following tumor cell implantation. Antibodies to PlGF and isotype-matched IgG1 control antibodies were administered by i.p. injection (25 mg/kg; twice weekly).
Mice were sacrificed after 9 weeks, and primary tumors were removed and weighted.

**Statistical analysis**

Data are presented as mean ± S.E.M., circulating levels of PlGF and sFlt1 as median with interquartile ranges. Statistical significance was determined by t-test, Fisher’s exact test, and Mann–Whitney U test using SPSS (v18.0) and GraphPad Prism (v5.0; San-Diego, CA, USA). Tumor-related survival and TTP were calculated based on the date of blood sampling and analyzed using the Kaplan–Meier method and Log-rank test. Cox proportional hazards regression model was used for multivariate analysis. *P < 0.05 values were considered significant; all tests were two-sided.

**Results**

**PlGF serum levels are elevated in patients with pNETs**

Given that PlGF is released into systemic circulation, we initially determined circulating PlGF in sera of patients with pNETs and healthy controls (Table 1). Median circulating PlGF concentration was found elevated to 19.5 pg/ml in pNET patients compared with 12.3 pg/ml in healthy controls (Fig. 1A). Receiver operating characteristic (ROC) analysis displayed 79.5% sensitivity and 82.8% specificity for the discrimination between patients with pNETs and controls at a cut-off level of 15.35 pg/ml (Fig. 1B), indicating that pNETs are associated with an increase in circulating PlGF.

As binding of PlGF to its soluble receptor Flt1 (sFlt1) may diminish its biological activity, we also determined sFlt1 levels. Though we found circulating sFlt1 increased in pNET patients compared with controls (Fig. 1C and D), the ratio PlGF:sFlt1 remained significantly elevated in patients with pNETs (Fig. 1E and F).

**Elevated PlGF serum levels correlate with tumor grading of pNETs**

We next hypothesized that circulating PlGF levels in patients with pNETs reflect histopathological properties and/or the clinical course of the disease. No correlation of PlGF with TNM classification was obtained. In particular, circulating PlGF did not discriminate between localized vs metastatic disease (Fig. 2A). However median PlGF levels progressively increased from grade 1 (G1) to 3 (G3) tumors (Fig. 2C).

**Levels of circulating sFlt1 correlate with metastatic disease in pNETs**

As sFlt1 has been proposed as a surrogate marker of tumor progression, we tested the association of sFlt1 levels with clinical parameters. Different from PlGF, sFlt1 levels correlated with metastatic disease (Fig. 2B). Median sFlt1 levels also progressively increased with grading (Fig. 2D).

**Elevated PlGF serum levels predict reduced tumor-related survival in pNETs**

The association of circulating PlGF and sFlt1 with tumor grade raised the question, whether these serum parameters reflected the prognosis. Accordingly, patients were grouped by high (>50th percentile) or low (≤50th percentile) levels of circulating PlGF, respectively, and Kaplan–Meier curves based on tumor-related survival were generated. Circulating PlGF levels > 50th percentile were associated with poor prognosis and predicted shorter tumor-related survival with a median survival of 4.52 years, whereas median survival in patients with low-PlGF serum concentrations had not yet been reached (hazard ratio: 2.35; 95% CI: 1.08–5.10; Log-rank P = 0.031; Fig. 2E). Subsequent multivariate analysis confirmed tumor grade, but not circulating PlGF as an independent prognostic parameter in the overall cohort (not shown), which is in accordance with the correlation of PlGF levels and grading described above. In contrast to PlGF, neither sFlt1 nor the PlGF:sFlt1 ratio correlated with tumor-related survival (Fig. 2F, and not shown).

**Prospective evaluation of PlGF as prognostic biomarker in NETs**

As our retrospective analysis suggested circulating PlGF as prognostic biomarker, we next sought to determine PlGF and sFlt1 in an independent validation cohort. Clinically, the availability of a prognostic biomarker would benefit the management of patients with lower grade tumors (G1 and G2), which often require a personalized therapeutic strategy to accompany them for decades. In order to be able to focus on lower grade tumors, yet recruit a prospective cohort within a reasonable time frame, we opened our analyses to intestinal NETs. Accordingly, patients with either low-grade pNETs or intestinal NETs were enrolled in a prospective study (Table 1). PlGF levels were found elevated in the overall cohort (n = 87), as well as in the pNET and intestinal subgroups (Fig. 3A), confirming our initial observation. We furthermore
confirmed increased sFlt1 concentrations as well as an elevated PlGF:sFlt1 ratio in NETs compared with healthy controls (Supplementary Figure 1, see section on supplementary data given at the end of this article).

Due to their slow progression, we were unable to determine tumor-related survival in the subgroup of intestinal NETs, and hence had to restrict survival analyses to pNET patients. Different from the retrospective analysis, a distinct cut-off value rather than the cohort-dependent median was required. As PlGF values from low-grade pNETs revealed considerable overlap with healthy controls, we choose the cut-off that best separated NET patients from healthy controls in our ROC analyses (15.35 pg/ml; Fig. 1B). Kaplan–Meier analyses using this cut-off indicated favorable tumor-related survival for individuals with non-pathologic circulating PlGF when

![Figure 1](image-url)

**Figure 1**
Elevated circulating PlGF and sFlt1 levels in patients with pNETs. (A) PlGF serum concentrations in healthy controls (n=87) and patients with pNET (n=88). Shown is the scatter dot plot with the median and interquartile range. *P<0.0001, Mann–Whitney U test. (B) Panel depicts corresponding ROC curve of circulating PlGF. AUC=0.85, 95% CI: 0.80–0.91, *P<0.0001. (C) sFlt1 serum concentrations in healthy controls (n=87) and patients with pNET (n=88). Shown is the scatter dot plot with a median of 73.9 pg/ml in healthy controls vs 83.6 pg/ml in pNET patients and interquartile range, *P<0.0001, Mann–Whitney U test. (D) Panel depicts corresponding ROC curve of circulating sFlt1. AUC=0.70, 95% CI: 0.62–0.78, *P<0.0001. (E) PlGF:sFlt1 ratios in healthy controls (n=87) and patients with pNET (n=88). Shown is the scatter dot plot with the median of 0.17 in healthy controls vs 0.23 in pNET patients and interquartile range, *P<0.0001, Mann–Whitney U test. (F) Panel depicts corresponding ROC curve of PlGF:sFlt1 ratio. AUC=0.76, 95% CI: 0.69–0.84, *P<0.0001.
compared with those with circulating PlGF > 15.35 pg/ml (Log-rank \( P = 0.025 \); Fig. 3B). Likewise, circulating PlGF levels furthermore allowed to separate prognostic subgroups within the group of G2 pNETs (Log-rank \( P = 0.032 \); Fig. 3C).

As was expected for the subgroup of intestinal NETs, survival is still pending in the prospective cohort and therefore we evaluated TTP instead. TTP could be determined in 47 out of 57 patients with G1 and G2 intestinal NETs, which were enrolled with stable disease. Kaplan–Meier analysis separating patients based on normal vs pathologic PlGF levels did not reveal significant differences. However, TTP was significantly reduced in patients with circulating PlGF above median (hazard ratio: 2.35; 95% CI: 1.08–5.10; Log-rank \( P = 0.031 \)).

Figure 2
Elevated circulating PlGF levels in patients with pNETs correlate to tumor grading and reduced survival. Correlation of circulating PlGF and sFlt1 levels with TNM stage and grading was tested. (A) No significant correlation (NS) was observed between serum PlGF levels in patients with localized (TNM stage I–III) vs metastatic disease (TNM stage IV). Shown is the scatter dot blot with the median and interquartile range; *\( P = 0.482 \) for TNM stage IV vs stage I–III, Mann–Whitney U test. (B) Serum levels of sFlt1 are significantly elevated in pNET patients with metastatic disease (TNM stage IV) compared with localized disease (TNM stage I–III). Shown is the scatter dot plot with the median and interquartile range; *\( P < 0.005 \) for TNM stage IV vs stage I–III, Mann–Whitney U test. (C) Correlation between serum PlGF levels and histological tumor grading as determined by Ki67 labeling index; G1 < 2%, \( n = 11 \); G2 2–20%, \( n = 47 \); G3 > 20%, \( n = 16 \). Shown is the scatter dot plot with median PlGF values of 16.6, 20.9, and 22.5 pg/ml for grade 1, 2, and 3, and interquartile range; *\( P = 0.036 \) for G1 vs. G2, \*\( P = 0.002 \) for G1 vs G3, Mann–Whitney U test. (D) Correlation between serum sFlt1 levels and histological tumor grading (groups as in C). Median sFlt1 was 74.8 pg/ml for G1, 84.6 pg/ml for G2, and 96.7 pg/ml for G3, respectively. Shown is the scatter dot plot with the median and interquartile range; *\( P = 0.013 \) for G1 vs G3, *\( P = 0.032 \) for G2 vs G3, Mann–Whitney U test. (E) Kaplan–Meier survival curve of pNET patients with high vs low levels of circulating PlGF. Patients were allocated to a group with high (> 50th percentile, corresponds to > 19.3 pg/ml, \( n = 43 \)) or low (≤ 50th percentile, \( n = 44 \)) PlGF serum levels, and Kaplan–Meier analyses were conducted; hazard ratio: 2.35; 95% CI: 1.08–5.10; Log-rank \( P = 0.031 \). (F) Kaplan–Meier analysis of tumor-related survival of pNET patients with high vs low levels of circulating sFlt1. Patients were allocated to a group with high (> 50th percentile, corresponds to > 83.6 pg/ml, \( n = 43 \)) or low (≤ 50th percentile, \( n = 44 \)) sFlt1 serum levels, and Kaplan–Meier analyses were conducted; hazard ratio: 1.25; 95% CI: 0.57–2.71; Log-rank \( P = 0.576 \).
Conversely, patients experiencing disease progression within 6 months presented higher median baseline PlGF levels than patients with stable disease throughout this period (26.7 vs 16.6 pg/ml; $P=0.009$). Moreover, median PlGF levels further stratified the clinically heterogeneous group of G2 intestinal NETs into prognostic subgroups (hazard ratio: 4.95; 95% CI: 1.25–19.60; Log-rank $P=0.023$; Fig. 3E). Finally, PlGF, but not grading or biotherapy with SSAs (29 out of 47 patients), constituted an independent prognostic parameter in the multivariate analysis of the intestinal NET cohort (Table 2). As grading is expected to emerge as an independent prognostic parameter in intestinal NETs, we inspected Ki67 values (hazard ratio: 4.95; 95% CI: 1.25–19.60; Log-rank $P=0.023$; Fig. 3E). Finally, PlGF, but not grading or biotherapy with SSAs (29 out of 47 patients), constituted an independent prognostic parameter in the multivariate analysis of the intestinal NET cohort (Table 2). As grading is expected to emerge as an independent prognostic parameter in intestinal NETs, we inspected Ki67 values

Figure 3
Prospective confirmation of elevated circulating PlGF levels and its prognostic relevance in patients with pancreatic and intestinal NETs. (A) PlGF serum concentrations in healthy controls ($n=87$) and a prospective cohort of patients with pNET ($n=30$) and intestinal NETs ($n=57$). Shown is the scatter dot plot with a median of 12.3 pg/ml in healthy controls vs 16.5 pg/ml in pNETs and 17.6 pg/ml in intestinal NETs, and the respective interquartile range, $^*P=0.001$ and $^*P<0.0001$, Mann–Whitney $U$ test. (B) Pathologic PlGF levels were associated with reduced tumor-related survival in low-grade pNETs of the prospective cohort. Patients were allocated to a group with pathological ($>15.35$ pg/ml, $n=52$) or normal ($<15.35$ pg/ml, $n=18$) serum PlGF levels, and Kaplan–Meier analyses were performed to compare tumor-related survival; Log-rank $P=0.025$. (C) Similar results were obtained with the subgroup of G2 pNETs when using the cut-off value of 15.35 pg/ml; Log-rank $P=0.032$. (D) Circulating PlGF values above median identify patients with rapid disease progression in intestinal NETs. Shown is a Kaplan–Meier analysis of time-to-progression in patients with intestinal NETs allocated to a group with high (>50th percentile, corresponds to $>17.1$ pg/ml, $n=23$) or low ($\leq$50th percentile, $n=24$) PlGF serum levels; TTP 15 months vs undefined; hazard ratio: 4.01; 95% CI: 1.38–11.65; Log-rank $P=0.011$. (E) Elevated circulating PlGF levels discriminate prognostic subgroups within the group of patients with grade 2 intestinal NETs. Patients were allocated to a group with high (>50th percentile, corresponds to $>17.8$ pg/ml, $n=12$) or low ($\leq$50th percentile, $n=13$) serum PlGF levels, and Kaplan–Meier analyses were performed to compare time-to-progression; TTP 10 months vs undefined; hazard ratio: 4.95; 95% CI: 1.25–19.60; Log-rank $P=0.023$. 

http://erc.endocrinology-journals.org
DOI: 10.1530/ERC-12-0223
© 2013 Society for Endocrinology
Printed in Great Britain
Published by Bioscientifica Ltd.
in our cohort in more detail. Indeed, 16 of the 21 G2 intestinal NETs had Ki67 values that were lower or equal to 5%, four had Ki67 values between 5 and 10%, and only one tumor had more than 10% Ki67 positive cells. We therefore speculate that this low-proliferative activity accounted for the comparable TTP in G1 and G2 subgroups.

Taken together, our prospective approach confirmed elevated circulating PlGF levels in pNETs and intestinal NETs and suggested PlGF levels as a prognostic parameter.

**pNETs are associated with de novo expression of PlGF**

As PlGF may be expressed and bound by cells from various tumor compartments, we aimed to further specify the abundance and location of PlGF in pNETs. Based on histomorphological criteria, pNET samples \( n = 23 \) consistently displayed strong PlGF expression in stromal cells, such as endothelial and inflammatory cells (Fig. 4A, B, C and E), and occasionally weak immunoreactivity for PlGF in the tumor cell compartment. In contrast, no staining was observed in non-transformed endocrine or exocrine pancreatic tissues, including their stroma (Fig. 4D and E), indicating de novo expression of PlGF in pNETs.

**PlGF enhances proliferation and motility of NET cells and is induced by targeted therapies in vitro**

To functionally address the role of PlGF, we initially determined effects of PlGF stimulation on NET cell biology in vitro. A panel of pancreatic (BON and QGP-1), ileal (KRJ-I), and bronchial (H727) NET cell lines was used. PlGF determination from supernatants of these tumor cell cultures was at (BON) or below (QGP-1, H727) the detection limit (not shown), and thus in agreement with the sparse immunohistochemical reactivity of epithelial tumor cells. However, expression analysis readily revealed the presence of at least one of the known PlGF receptors, Flt1, NRP1, and NRP2 in these NET cell lines (Supplementary Figure 2, see section on supplementary data given at the end of this article), thereby enabling responsiveness to paracrine PlGF stimulation. Indeed, recombinant PlGF enhanced proliferation of all four NET cell lines tested in clinical specimens \( n = 23 \). Immunoreactivity for PlGF was determined from the percentage of immunoreactive area/total tumor area, without additional assessment of staining intensity. Data represent mean ± S.E.M. *P < 0.05; t-test.

**Table 2**  Multivariate analysis for time-to-progression (TTP) in intestinal NETs \( n = 47 \).

<table>
<thead>
<tr>
<th>Covariate</th>
<th>P value</th>
<th>HR (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>0.601</td>
<td>0.99 (0.93–1.04)</td>
</tr>
<tr>
<td>Sex (female/male)</td>
<td>0.450</td>
<td>0.64 (0.20–2.02)</td>
</tr>
<tr>
<td>TNM-stage</td>
<td>0.988</td>
<td>0.00*</td>
</tr>
<tr>
<td>Grading</td>
<td>0.737</td>
<td>1.21 (0.40–3.71)</td>
</tr>
<tr>
<td>Biotherapy (SSA)</td>
<td>0.349</td>
<td>1.76 (0.54–5.78)</td>
</tr>
<tr>
<td>Serum PlGF levels</td>
<td>0.029</td>
<td>4.87 (1.18–20.13)</td>
</tr>
</tbody>
</table>

HR, hazard ratio; SSA, somatostatin analogs.

*Out of 47 subjects 46 were TNM-stage IV.

**Figure 4**

*De novo* expression of PlGF occurs in human pNETs. (A, B, C and D) Representative immunohistochemistry for PlGF on cryosections of clinical specimens from patients with pNETs (A, B and C) and respective adjacent non-transformed tissue (D). Scale bars = 100 μm. (E) Semicquantitative analysis of overall PlGF expression in tumor epithelial and stromal cells of
In view of the biological action of PlGF on NET cells, we next tested, whether recently approved targeted therapies affect PlGF production of NET cells in vitro. To allow for paracrine tumor cell/stroma interactions, we determined PlGF production from different endothelial cell and fibroblast preparations, including NET-derived tumor fibroblasts. PlGF was consistently detected in supernatants from endothelial cells, but not from fibroblast cultures (not shown), prompting us to choose mixed spheroid cultures of BON cells with endothelial cells as an in vitro approach (see Supplementary Methods). Incubation of spheroid co-cultures with the tyrosine-kinase inhibitor sunitinib, but not with the mTOR-inhibitor everolimus over 72 h elevated PlGF levels in the supernatant (264.3 ± 38.7% of control; P = 0.02; and 122.3 ± 10.5% of control; P = 0.13). Thus, PlGF release increased in response to sunitinib, suggesting that PlGF induction in NETs may occur following antiangiogenic treatment.

### Blocking PlGF inhibits growth of orthotopic NETs

Finally, we decided to investigate the function of PlGF in vivo, using an established preclinical NET model. Accordingly, BON orthotopic pNET xenografts were grown in NMRI nu/nu mice and engrafted tumors treated with either isotype-matched control IgG1 or a combination of neutralizing antibodies to mouse and human PlGF from week 3 following tumor cell implantation. By the end of the experiment in week 9, functional inactivation of tumor cell and stroma-derived PlGF by the neutralizing antibodies had resulted in a significant reduction of tumor weight, indicating that PlGF supports NET growth in vivo and constitutes a therapeutic target (Fig. 5H, I and J).

### Discussion

The current study highlights PlGF as a novel, stroma-derived growth factor in pancreatic and intestinal NETs. In vitro, PlGF enhanced proliferation and migration of NET cell lines, suggesting that PlGF directly affects NET cell biology. In vivo, blocking PlGF using neutralizing antibodies reduced growth of orthotopic BON NET xenograft tumors, indicating that PlGF represents a therapeutic target in preclinical NET models. In the clinical situation, elevated circulating PlGF levels were associated with reduced tumor-related survival and/or shorter TTP in NETs, suggesting circulating PlGF as an easily accessible candidate prognostic biomarker.

The therapeutic concept of antiangiogenesis has created much interest in tumor-type specific angioregulatory growth factors. Studies conducted in NETs found that VEGF-A, the prototype angiogenic factor in cancer, is highly expressed in intestinal NETs (Terris et al. 1998) but was found less abundant in pNETs, inversely correlated to microvascular density, and did not convey prognostic information in pNETs (Terris et al. 1998, Marion-Audibert et al. 2003, Couvelard et al. 2005). Here, we find that PlGF, a member of the VEGF family, is prominently induced in the stroma of pNETs. Although de novo expression of PlGF reportedly may occur in the tumor cell compartment (Fischer et al. 2008), our analysis suggests that this was rarely the case in pNETs. Rather, the tumor stroma of pNETs uniformly revealed abundant PlGF immunoreactivity. Genetic and pharmacological evidence attests to the functional relevance of stroma-derived PlGF in that inactivation of stromal PlGF impaired (Carmeliet et al. 2001, Schmidt et al. 2011), whereas transgene overexpression of stromal PlGF stimulated tumor growth, angiogenesis, and metastasis in mouse models (Marcellini et al. 2006, Fischer et al. 2007, 2008). Given that the known PlGF receptors, Flt1, and neuropilins were shown expressed in NETs (Terris et al. 1998, von Marschall et al. 2003), our current observation of stromal PlGF expression suggests the presence of auto-/paracrine signaling loops. Functionally, PlGF stimulated proliferation and migration of NET cells in vitro, which is in line with published results revealing stimulatory actions of PlGF on proliferation of pancreatic tumor cells (Fischer et al. 2007), as well as on migration of glioma, breast, and lung cancer cells (Fischer et al. 2008, Taylor et al. 2010). More importantly, we were able to reduce the growth of BON NET xenograft tumors using neutralizing antibodies to functionally inactivate tumor cell- and stroma-derived PlGF.

A second line of evidence strongly implicates PlGF in the pathology of NETs: circulating PlGF levels were elevated in pNET patients, correlated with tumor grade, and predicted poor clinical outcome. In contrast, PlGF did not correlate with TNM stages and did not differ between localized (TNM stages I–III) and metastatic disease (TNM stage IV). We therefore favor the concept that high circulating PlGF levels reflected a more aggressive tumor biology rather than a more extensive tumor load.
Although expression of PlGF in tumor tissues was proposed as prognostic marker for progression and survival in gastric, colorectal, NSCLC, hepatocellular, and breast cancer (Parr et al. 2005, Wei et al. 2005, Fischer et al. 2008), relatively few studies evaluated circulating levels of PlGF. Results linked high circulating PlGF to adverse prognosis in renal (Matsumoto et al. 2003) and oral squamous cell carcinoma (Cheng et al. 2012) but not in hepatocellular...
cancer (Nagaoka et al. 2010), and high preoperative PlGF levels predicted reduced recurrence-free and overall survival in colorectal cancer patients undergoing curative surgery (Wei et al. 2009, Rahbari et al. 2011). The latter data fit with our observation that PlGF levels >50th percentile correlated with shorter TTP in the prospective intestinal NET cohort. Indeed, we identified circulating PlGF as a prognostic indicator of TTP in an intestinal NET collective with low overall proliferative activity, in which grading was unable to separate prognostic subgroups. Furthermore, pathologic PlGF levels identified distinct subgroups regarding tumor-related survival within the heterogeneous group of G2 pNETs in our prospective approach. Currently, we cannot pinpoint the cause of the more robust separation of prognostic subgroups of G2 intestinal NETs by median PlGF levels compared with the separation of G2 pNET prognostic subgroups by pathologic PlGF levels. Difference in end points (tumor-related survival vs TTP), differences in NET types (pNET vs intestinal NET), or differences in treatment modalities offer potential explanations that will have to be thoroughly evaluated in future prospective studies.

Irrespective of such adjustments, our current data established the link from circulating PlGF to disease prognosis, which could have considerable translational impact on the management of NETs based on three considerations:

First, advanced G2 tumors comprise a large and clinically heterogeneous group of patients, in which the course of disease and resultant treatment decisions are variable, with choices currently based on empirical knowledge rather than objective parameters. In the current study, high PlGF levels in patients with G2 tumors specifically characterized individuals with an unfavorable prognosis. Notably, absolute Ki67 values within the G2 subgroup did not correlate to PlGF levels \( r=0.156; P=0.215 \), suggesting that PlGF conveyed prognostic information that is distinct from Ki67-based grading. Hence, circulating PlGF may serve to further stratify patients with G2 tumors and help to optimize clinical management, as it could identify patients that benefit from immediate/more aggressive treatment and/or shorter monitoring intervals.


In NET disease, which often requires long-term treatment regimens, prevention of co-morbidity, which might be of greater risk to patients than the malignancy itself, is of crucial importance. So far, a phase I clinical trial with the humanized anti-PlGF antibody TB-403 has proven lack of toxicity (Martinsson-Niskanen et al. 2011, Lassen et al. 2012). Moreover the recent approval of sunitinib for the treatment of pNETs (Raymond et al. 2011) might have substantial implications on considering PlGF as therapeutic target in these tumors, as sunitinib efficiently blocks Flt1 signaling (Fischer et al. 2008).

Third, changes in circulating PlGF may occur as a consequence of treatment. Patients with NETs may undergo decades of therapy. Apart from side effects of long-term treatments, resistance development therefore constitutes a major concern, especially when considering antiangiogenic treatments (Paez-Ribes et al. 2009). In this context, mouse models revealed that circulating and tumoral PlGF levels increase upon VEGF inhibition and conventional chemotherapies, resulting in recruitment of proangiogenic macrophages and angiogenic escape (Fischer et al. 2007). Similarly, chronic exposure of colorectal cancer cells \textit{in vitro} to bevacizumab increased the expression of PlGF and consequently enhanced tumor cell migration and invasion, and metastatic potential \textit{in vivo} (Fan et al. 2011). Of clinical relevance, induction of circulating PlGF occurred in patients receiving VEGF inhibitors for treatment of colorectal and renal cell cancers and glioblastoma (Motzer et al. 2006, Batchelor et al. 2007, Fischer et al. 2007, Willett et al. 2009, de Groot et al. 2011). Thus, it is tempting to speculate that early and/or transient increases in circulating PlGF may predict therapy response, while long-term induction of circulating PlGF might constitute a functional mechanism and/or indicator of resistance development. Given the recent approvals of sunitinib and everolimus for treatment of pNETs, circulating PlGF may represent an easily accessible candidate biomarker for monitoring therapy response and/or disease progression. While sunitinib-dependent effects on circulating PlGF levels have not yet been determined in NET patients, PlGF measurements from the RADIANT-3 trial reported a transient reduction in response to everolimus, which is difficult to interpret. At a first glance, the decrease of circulating PlGF levels seems counterintuitive in view of the antiangiogenic activity attributed to everolimus, which might be expected to induce a compensatory rise of PlGF. However, reported
values give a snapshot of circulating PlGF levels obtained at 28 days of treatment, and the moderate antiangiogenic action of everolimus may require a longer treatment period for efficient induction of an angiogenic escape. Furthermore, PlGF reduction may have resulted from reduced tumor burden in treatment responders.

With the advent of novel therapies for NETs, a personalized therapy may have come into reach, but a definition of biomarker profiles that are useful for the stratification of subgroups is mandatory. PlGF holds promise for NETs in multiple ways: as a prognostic marker, as potential therapeutic target, and – possibly – as a response marker for selected systemic therapies. Our current analyses provide an intriguing link from high PlGF serum levels to poor outcome that deserves an urgent prospective follow up in larger cohorts of pancreatic and intestinal NETs.

Thus, our data provide a compelling basis for a more extensive prospective evaluation of PlGF in larger and more diverse NET cohorts. Finally, our observations strongly advocate an inclusion of PlGF into the portfolio of biomarkers in ongoing and/or further prospective NET trials.

Supplementary data
This is linked to the online version of the paper at http://dx.doi.org/10.1530/ERC-12-0223.

Declaration of interest
G Hess was previously an employee of Roche Diagnostics and is currently a consultant to Roche Diagnostics. JM Stassen was previously an employee of ThromboGenics NV and is currently a scientific advisor to ThromboGenics NV. The other authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

Funding
G Hilfenhaus was supported by a Gunther Speidel scholarship. C Fischer was supported by a grant from Experimental and Clinical Research Center, Berlin, Germany. This work was supported by a grant from Berliner Krebsgesellschaft e.V., Berlin, Germany.

Acknowledgements
We thank M Welzel for outstanding technical assistance, and R Pfagner and I Modlin for the opportunity to use the KRJ-I cells.

References


Received in final form 6 February 2013
Accepted 4 March 2013
Made available online as an Accepted Preprint 5 March 2013