RANK- and c-Met-mediated signal network promotes prostate cancer metastatic colonization

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Abstract

Prostate cancer (PCA) metastasis to bone is lethal and there is no adequate animal model for studying the mechanisms underlying the metastatic process. Here, we report that receptor activator of NF-κB ligand (RANKL) expressed by PCA cells consistently induced colonization or metastasis to bone in animal models. RANK-mediated signaling established a premetastatic niche through a feed-forward loop, involving the induction of RANKL and c-Met, but repression of androgen receptor (AR) expression and AR signaling pathways. Site-directed mutagenesis and transcription factor (TF) deletion/interference assays identified common TF complexes, c-Myc/Max, and AP4 as critical regulatory nodes. RANKL–RANK signaling activated a number of master regulator TFs that control the epithelial-to-mesenchymal transition (Twist1, Slug, Zeb1, and Zeb2), stem cell properties (Sox2, Myc, Oct3/4, and Nanog), neuroendocrine differentiation (Sox9, HIF1α, and FoxA2), and osteomimicry (c-Myc/Max, Sox2, Sox9, HIF1α, and Runx2). Abrogating RANK or its downstream c-Myc/Max or c-Met signaling network minimized or abolished skeletal metastasis in mice. RANKL-expressing LNCaP cells recruited and induced neighboring non metastatic LNCaP cells to express RANKL, c-Met/activated c-Met, while downregulating AR expression. These initially non-metastatic cells, once retrieved from the tumors, acquired the potential to colonize and grow in bone. These findings identify a novel mechanism of tumor growth in bone that involves tumor cell reprogramming via RANK–RANKL signaling, as well as a form of signal amplification that mediates recruitment and stable transformation of non-metastatic bystander dormant cells.

Key Words
- RANKL
- RANK
- c-Met
- prostate cancer
- metastasis
- cancer dormancy

Introduction

Bone is the most common site of prostate cancer (PCA) metastases. Metastatic lesions in bone are significantly associated with bone pain, hypercalcemia or hypocalcemia, pathological fracture, and spinal cord compression (Coleman 2001). Receptor activator of NF-κB ligand (RANKL)–RANK signaling has many crucial
physiological roles in bone and other tissues (Hanada et al. 2009), and aberrant RANK–RANK signaling in cancer and bone cells affects cancer bone colonization (Dougall 2011). We reported previously that β2-microglobulin (β2-M), a major histocompatibility protein co-receptor, promotes PCA cell osteomimicry by inducing RANKL and non-collagenous bone matrix proteins (Huang et al. 2006, Odero-Marah et al. 2008). β2-M-induced RANKL expression promoted epithelial-to-mesenchymal transition (EMT) in PCA cells (Zhau et al. 2008). Forced expression of β2-M in non-metastatic human breast, lung, and kidney cancer cells enhanced endogenous RANKL expression and induced EMT and bone and soft tissue homing (Josson et al. 2011). These findings support an important role for RANKL–RANK signaling in PCA metastasis.

Hepatocyte growth factor (HGF) and its receptor tyrosine kinase c-Met mediate cell motility, migration, increased tumor local invasion, and metastasis (Knudsen & Edlund 2004). Deregulation of the c-Met network is one of the most common mechanisms of solid tumor development (Birchmeier et al. 2003). The c-Met receptor cross talks with many membrane-localized signaling proteins, including EGFR, erbB2, erbB3, c-Src, and G-protein-coupled receptors, indicating the existence of numerous mechanisms of tumor cell escape from therapies directed at single oncogenic targets (Organ & Tsao 2011). c-Met inhibitors with differential selectivity have been developed based on their relative ATP competitive binding. Cabozantinib (XL-184), a recently FDA approved mixed c-Met, VEGFR2, KIT, and AXL inhibitor, proved highly effective against bone metastatic PCa (Smith et al. 2013). c-Met activation can be ligand-dependent or -independent and is associated with androgen receptor (AR)-deficient castration-resistant prostate cancer (CRPC; Verras et al. 2007) and AR-negative PCA stem cells (van Leenders et al. 2011).

Here, we describe a feed-forward loop, involving the induction of RANKL and c-Met signal and repression of AR, which drives RANK receptor-mediated bone metastasis. Novel RANK-mediated downstream master regulator (MR) transcription factors (TFs) were identified that coordinate PCA cell EMT, stemness, neuroendocrine, and osteomimicry phenotypes. We validated the roles of TFs and effector molecules in PCA metastasis with in vivo animal models guided by molecular imaging where abrogating RANK or its downstream c-Myc/Max or c-Met signaling network abolished skeletal metastasis in mice. Animal models also showed that RANKL-expressing PCA cells conferred bone colonizing and aggressive phenotypes to neighboring non-metastatic bystander cells by activating the RANK-mediated downstream signaling network. Significantly, RANKL and its downstream signaling network in primary human PCA tissues predict patient survival (Hu et al. 2013).

Subjects and methods

Cell culture

ARCaP_E and ARCaP_M cells established by our laboratory (Xu et al. 2006) represent stages of human PCA progression. They were maintained in T-medium (Invitrogen) supplemented with 5% FBS. LNCaP and LNCaPNeo/RANKL were maintained in RPMI-1640 supplemented with 10% FBS. RAW264.7 cells, provided by Dr Neale Weitzmann at Emory University, were maintained in DMEM supplemented by 10% FBS.

Cloning RANKL expression vectors

Human RANKL cDNA ORF (NM_003701) purchased from OriGene (Rockville, MD, USA) was amplified by PCR and subcloned into p3×FLAG-myc-CMV-25 (Sigma–Aldrich) at the NotI and Xbal restriction enzyme sites. The amplified human RANKL cDNA was subcloned into pCDH-CMV-MCS-EF1-copGFP (System Biosciences, Mountain View, CA, USA) plasmids at the Xbal and NotI restriction enzyme sites. The sequences of p3×FLAG-RANKL and pCDH-CMV-RANKL-EF1-copGFP were confirmed by DNA sequencing.

Cell transfection, transduction, and gene deletion protocols

LNCaP cells were transfected with either p3×Flag-tagged RANKL or neo-control-plasmid (p3×FLAG-myc-CMV-25) cDNA (Sigma–Aldrich) for 48 h. The stable cell clones were selected in 400 μg/ml of G418 until individual colonies containing the transfected construct were confirmed by western blot analysis. The retrovirus packaging cells, 293GPG, were maintained in DMEM with 10% heat-inactivated FBS supplemented with tetracycline, puromycin, G418, and penicillin/streptomycin. The shRANK-1 and -4 retroviruses were generated using the retroviral small interfering RNA (siRNA) constructs pPower-hRK-1 (hRK-1) and pPower-hRK-4 (hRK-4) prepared using standard molecular cloning techniques and were used to infect ARCaP_M and LNRANKL cells for 24 h in the presence of 8 μg/ml of polybrene. The oligosquences of RANK and control short hairpin RNA (shRNA) are as follows: shRANK-1, CCAGAAGATATGTGCTACCCA; shRANK-4, TGGGACGCTGTCTAGCAAAA; and shCon,
ACCACATCTTATAGGTTGTT. The transfected cells as well as parental and tumour-derived LNNeo-RFP cells were also transduced with firefly luciferase retroviruses (MSCV Luciferase PGK Hygro; Addgene, Cambridge, MA, USA) and selected with 200 μg/ml hygromycin. To evaluate the possible functions of key receptors and TFs in PCa metastasis, we conducted genetic deletion studies of c-Met and c-Myc and/or Max from LNNeo cells. c-Myc (sc-29226-V) and Max (sc-38080-V) shRNA viral particles were purchased from Santa Cruz Biotechnology, Inc. and c-Met shRNA viral particles (TRCN0000009851, TRCN0000040044, and TRCN0000121087) were purchased from Sigma–Aldrich.

In vivo experiments

All animal procedures were performed according to an approved protocol from the Institutional Animal Care and Use Committee. LNNeo, LNNeo, or LNNeo-RFP cells (1 × 10⁶ cells/50 μl PBS) were tagged with the luciferase gene and inoculated intracardially or intratibially into 5- to 7-week-old male athymic nude mice (Charles River, Wilmington, MA, USA) as described previously (Odero-Marah et al. 2008). To study their interactions and recruitment, in some studies, cells were injected together or separately at different ratios or by a separate route. All mice were imaged weekly with either bioluminescence or fluorescence (red fluorescent protein (RFP): excitation, 620 nm; a near-infrared MHI-148 organic dye: excitation, 783 nm, emission, 840 nm (Yang et al. 2010)) using Xenogen and scanned by X-ray by Luminar XR or X-ray microtomography (μCT) by Scanco vivaCT40 to examine the skeleton and determine the types of bone lesions.

RT-PCR and quantitative real-time PCR

Total RNA from cells was isolated using an RNeasy Mini Kit (Qiagen) according to the manufacturer’s instructions. cDNA was generated from 3 μg of total RNA using a SuperScript III First-Strand Synthesis System (Invitrogen), Quantitative real-time PCR was performed by the ABI 7500 Fast system in a total of 20 μl of reaction containing 1 μl cDNA, 1 μl primer pairs, 8 μl ABI SYBR Green Master Mix (Applied Biosystems), and 8 μl RNase-free water, and run at 95 °C, 3 min, followed by 40 cycles of 95 °C, 30 s and 60 °C, 30 s. Real-time PCR primer sequences are listed in Table 1.

Western blot analysis

Proteins (30 μg) were resolved on a 4–12% Bis–Tris gradient SDS–PAGE under reducing conditions and transferred onto nitrocellulose membrane. The primary antibodies were RANKL, E-cadherin, vimentin, OPG, c-Met (Santa Cruz Biotechnology, Inc.), p-c-Met (Tyr-1230/34; Invitrogen), RANK (Amgen, Thousand Oaks, CA, USA), and N-cadherin (BD Transduction Laboratories, San Jose, CA, USA). AR (441), Chr-A (H-300), synaptotherin (SYP; D4), CD44 (DFII485), Sox-2 (Y-17), Nanog (SA10), LIN-28 (H-44) (Santa Cruz Biotechnology, Inc.), FOXA2 (D56D6; Cell Signaling Technology), and PROM1 (CD133; Abnova, Taipei City, Taiwan) antibodies were used to detect neuroendocrine and stem cell differentiation. c-Myc (D84C12XP), Lamin A/C (Cell Signaling Technology), and Max (sc-197X) antibodies (Santa Cruz Biotechnology, Inc.) were used for nuclear protein detection.

### Table 1 Real-time PCR primer sequences

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<th>Genes</th>
<th>Real-time PCR primer sequences</th>
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<td>OPG</td>
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<td>c-Met</td>
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<tr>
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<tr>
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<tr>
<td>AR</td>
<td>F: 5′-GACCATGGTGGCTTGTCTGAC-3′  R: 5′-GGGTTGCTGAGCCAAATTT-3′</td>
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Immunohistochemistry

Human PCa tissue arrays in FFPE were purchased from Invitrogen and the PCa bone metastasis array (UWTMA22) was kindly provided by Dr Robert L Vessella from the University of Washington (Seattle, WA, USA). Immunohistochemical (IHC) staining followed a previously published protocol (Zhau et al. 2008) using primary antibodies against RANKL FL-317, c-Met C-12 (Santa Cruz Biotechnology, Inc.), and p-c-Met (pYpYpY1230/1234/1235; Invitrogen). IHC staining intensity was scored using the combined intensity and percentage of positive-scoring cells as previously reported (De Marzo et al. 1999). Strong intensity was scored as 3, intermediate as 2, weak as 1, and negative as 0. Each intensity score was then summed with the score of the percentage of cells that were stained, with >50% of the cells as 2, <50% of the cells as 1, and none as 0. One-way ANOVA was used to analyze the population scores between the two stages of cancer progression (bone vs benign, well-differentiated, or poorly-differentiated vs benign). The Gleason grade of the tumors and the quantification of the staining were determined by a pathologist, Dr Peizhen Hu.

Construction of RANKL and c-Met promoter-luciferase reporter plasmid and deletion mutants

The 2.5 kb human RANKL promoter was amplified from the human BAC clone RP11-86N24 using primers 5’-GTGACAGAATTCCTAGGGGCAAGTC-3’ (forward) and 5’-GGCGAAGCTTTGCGCTGGGCTAGCTCCGCTTG-3’ (reverse). The amplified fragment was subsequently cloned into a firefly luciferase expression vector pGL3-basic (Promega). The authenticity of the 2.5 kb RANKL promoter region was confirmed by DNA sequencing. A series of deletion mutants of the RANKL promoter were generated from the 2.5 kb RANKL promoter using a QuikChange II XL Site-Directed Mutagenesis Kit following the manufacturer’s instructions (Stratagene, Santa Clara, CA, USA). Human c-Met promoter-luciferase and its deletion constructs were provided by Dr Y Liu (Department of Pathology, University of Pittsburgh). Additional deletion constructs were also generated by the above protocol. Human AR promoter (6 kb) was provided by Dr Donald Tindall (Mayo Clinic).

Transient transfection and luciferase reporter assay

RANKL, c-Met, and AR promoters and deletion mutants plus β-galactosidase plasmid (for transfection efficiency control) were transiently transfected into PCa cells using Lipofectamine 2000 (Invitrogen) for 48 h. After 48 h, the transfected cells were serum-starved for 24 h before treatment with RANKL, OPG, or RANKL plus OPG for another 48 h. For c-Myc inhibition, LNCaP cells were pretreated with 40 μM 10058-F4 c-Myc inhibitor for 4 h before treatment with RANKL. The luciferase reporter assay was conducted as described previously (Huang et al. 2010).

Flow cytometry

Fluorescence-activated cell sorting (FACS) analyses were conducted as described previously (Sheridan et al. 2006). Stained cells were analyzed by BD Accuri C6 flow cytometer (BD, Franklin Lakes, NJ, USA) and results further analyzed using FlowJo Software (Ashland, OR, USA).

Microarray analysis

Human U133plus2.0 array hybridizations were performed by the UCLA Clinical Microarray Core following the standard Affymetrix GeneChip Expression Analysis protocol. The acquisition of array images was undertaken using the Affymetrix GeneChip Command Console 1.1 (AGCC). The microarray data are publically available (GEO GSE48432).

Computational analyses of global gene expression profile

The guanine cytosine robust multi-array analysis (GCMRA) method was applied to adjust the background signal. Probe intensities were normalized using the quantile normalization procedure (Wu et al. 2004). Integrated hypothesis testing included i) two independent tests, the T-test and the log2 median ratio test; ii) for each test, an empirical distribution of the null hypothesis that the means of the genes are not different was estimated by random permutations of the samples; iii) for each gene, the false discovery rate (FDR) was computed by a two-tailed test using the empirical distributions by Storey’s method (Storey & Tibshirani 2003); and iv) the two sets of FDRs from the individual tests were combined to compute the overall FDR using Stouffer’s method (Hwang et al. 2005). Finally, differentially expressed genes (DEGs) were selected for i) FDR < 0.05 and ii) absolute expression changes larger than twofold. From 3345 DEGs (FDR < 0.05, log2 fold change ≥ 1), we found that 1644 genes were upregulated and 1701 genes were significantly downregulated. A functional enrichment analysis of the list of up- and downregulated genes using the Database for Annotation, Visualization and Integrated Discovery (DAVID) Software (Bethesda, MD, USA) (Huang da et al. 2009) identified cellular processes enriched by the DEGs.
Probability of significance was transformed into enrichment score, which is $-\log_{10}$ (enrichment $P$ value).

**MR analysis for identification of key TFs**

To identify key TFs, we first collected about 780,000 items of TF target interaction data for 391 TFs in the public databases including TRED (Zhao et al. 2005), EEDB (Severin et al. 2009), mSigDB (Subramanian et al. 2005), Amadeus (Linhart et al. 2008), OregAnno (Griffith et al. 2008), PAZAR (Portales-Casamar et al. 2007), and ChEA (Lachmann et al. 2010). We used Fisher’s exact test (FET; Carro et al. 2008), OregAnno (Griffith et al. 2008), mSigDB (Subramanian et al. 2005), Amadeus (Linhart et al. 2008), and ChEA (Lachmann et al. 2010). We used Fisher’s exact test (FET; Carro et al. 2008) to compute the significance of overlap between the TF-targets and DEGs of interest including RANK receptor downstream signaling, TF modules, EMT, stemness, and neuroendocrine differentiation. Then we selected eight TFs whose targets were significantly enriched by the upregulated DEGs ($P < 0.01$ in FET).

**Identification of disease phenotypes**

Upregulated genes in RANKL-overexpressed cells compared with control cells were used to identify associated human disease phenotypes using g:Profiler (Reimand et al. 2011), providing gene annotations from the HPO, a standardized vocabulary of phenotypic abnormalities encountered in human disease. Finally, we selected a list of disease/disease phenotype associations enriched by the 1644 upregulated genes with $P < 0.05$.

**Statistical analysis**

Differences between groups were analyzed using Student’s $t$-test (two groups), one-way ANOVA (three or more groups), two-way ANOVA (two or more factors), or FET as appropriate. Where multiple groups were compared using ANOVA, a post hoc Tukey’s or Dunnett’s method was used to enable multiple comparisons between groups. Data that were not normally distributed were log-transformed before statistical tests were performed. A $P$ value $< 0.05$ was considered statistically significant. All statistical analysis was performed using R v2.15.1.

**Results**

**RANKL expression increases with human PCa progression and is capable of driving non-metastatic PCa cells to colonize bone and soft tissues in mice**

RANKL is prevalently expressed in human PCa specimens, with increased expression in higher grade and metastatic tumors compared with benign and low-grade PCa (Fig. 1A). We previously demonstrated that RANKL expression was significantly correlated with the overall survival of PCa patients (Hu et al. 2013). Moreover, RANKL expression correlates with the ability of PCa to metastasize to bone. ARCaPM and LN<sup>RANKL</sup> cells expressed respectively more intrinsic or transfected RANKL than their parental ARCaP<sub>E</sub> and LN<sup>Neo</sup> cells as determined by IHC (Fig. 1B). ARCaPM cells were observed to express high levels of endogenous RANKL. Silencing of RANKL in ARCaPM cells resulted in attenuation of the cells’ mesenchymal phenotype (Fig. 1C) and their migratory and invasive potential (Fig. 1D). Consistent with this result, silencing of RANK, the cognate receptor for RANKL, abolished the cells’ metastatic potential (Fig. 1E). Because androgen-responsive, non-metastatic LNCaP cells expressed low levels of RANKL, we tested whether stable, forced RANKL expression in LNCaP (LN<sup>RANKL</sup>) cells promoted RANKL–RANK signaling and bone metastasis. Relative expression levels of RANKL, RANK, and OPG in LN<sup>RANKL</sup> and control non-transfected LN<sup>Neo</sup> cells, ARCaP<sub>E</sub>, and ARCaPM are shown in Fig. 2A. To test the possibility that RANKL expression in PCa cells confers bone metastasis in mice, $1 \times 10^6$ LN<sup>RANKL-Luc</sup> or control LN<sup>Neo-Luc</sup> cells were injected intracardially into mice. In marked contrast to the indolent LN<sup>Neo-Luc</sup> (0/15), LN<sup>RANKL-Luc</sup> cells metastasized primarily to bone (20/20) and, less uniformly, to lymph node (18/20), lung (8/20), and adrenal gland (17/20) as evaluated by bioluminescence (Fig. 2B). The predominant metastatic lesions in mouse skeleton were osteolytic, with a minor osteoblastic component as evaluated by μCT (Fig. 2C). The presence of tumor cells and the differentiation of pre-osteoclasts to mature osteoclasts in PCa bone lesions was validated by H/E and positive TRAP staining (Fig. 2D). Because LN<sup>RANKL</sup> cells produced secreted RANKL (~3 ng/ml; data not shown), we tested the effect of i.p. injection of recombinant soluble RANKL (sRANKL, 50 μg/kg, twice a week) on bone colonization of intratibially inoculated, non-metastatic LN<sup>Neo-BFP-Luc</sup> cells. Mice that had received injections of RANKL exhibited bone metastases while no tumors formed in the skeleton in the animals that received injections of vehicle only (Fig. 2E). sRANKL was shown previously to induce bone turnover/resorption in rat bone; however, spontaneous tumor formation in response to this cytokine does not occur (McHugh et al. 2003).
This analysis showed that increased expression of RANKL promoted the expression of i) development-related genes regulating stem cells, neuronal differentiation, and morphogenesis; ii) genes controlling cell migration, angiogenesis, chemotaxis, and EMT; iii) genes associated with increased cell proliferation and decreased apoptosis; and iv) genes controlling bone development, bone morphogenesis, ossification, and tissue renewal (Supplementary Fig. 1, see section on supplementary data given at the end of this article). Increased RANKL expression significantly activated a number of cell signaling networks downstream from RANK that support cell growth, survival, and cell cycle.
RANKL overexpression in PCa cells confers increased lethal and osteolytic bone metastasis in nude mice. (A) RANKL, RANK, and OPG expression was assessed in human PCa ARCaPc and ARCaPM EMT cell model and human LNneo and LNRANKL cells by qRT-PCR (\(*P<0.05; **P<0.01; \text{and} ***P<0.001\)). (B) Representative bioluminescent images showed that luciferase-tagged LNNeo cells, but not luciferase-tagged LNneo control cells, induced bone and soft tissue metastases in nude mice (total flux (photons/s): LNNeo-Luc, 1.88 \(\times\) 10^6, 1.53 \(\times\) 10^6, and 1.05 \(\times\) 10^6; LNRANKL-Luc, 1.92 \(\times\) 10^5, 3.79 \(\times\) 10^5, and 4.14 \(\times\) 10^5). (C) Representative 3D \(\mu\)CT images of mice bearing LNNeo and LNRANKL cells showed that RANKL overexpression induced predominantly osteolytic lesions in mouse skeleton. (D) Representative H/E and positive TRAP staining in tumors in mouse bone induced by LNRANKL cells. (E) Representative red fluorescent or bioluminescent images showed sRANKL (50 \(\mu\)g/kg i.p. twice a week, 2 weeks after intraosseous inoculation of 1 \(\times\) 10^6 PCa cells) induced tumor growth in mouse tibia inoculated with LNneo-RFP-Luc cells (\(n=4\)). No tumor was observed in vehicle-treated mice (average total radiant efficiency of RFP (photons/(s)/W/cm^2)) or total flux (photons) of bioluminescence: vehicle, 3.68 \(\times\) 10^6 (RFP) and 8.96 \(\times\) 10^6 (bioluminescence); sRANKL, 2.34 \(\times\) 10^7 (RFP) and 8.63 \(\times\) 10^7 (bioluminescence)).
c-Met activation, as indicated by increased phosphorylation (at tyrosine 1230/1234/1235 sites), was also increased in LNRANKL cells. Expression of RANKL, c-Met, and activated c-Met was attenuated by a RANKL decoy receptor, OPG, or an anti-RANKL antibody, denosumab (Fig. 4B). These findings indicate that RANK–RANKL signaling significantly alters an oncogenic signaling program. To approach the mechanism underlying these changes, we used RANKL-, c-Met-, and AR-promoter luciferase constructs to identify cis-elements and TFs that may control them. Both the RANKL- and c-Met promoter regions contain E boxes (CACGTG) at −1384 or −1250 bp respectively. These are consensus regions for c-Myc/Max binding (Fig. 4D and E). Upregulation of RANKL and c-Met expression by enhanced RANK–RANKL signaling was mediated through direct interaction of the c-Myc/Max heterodimer to the E-box region within RANKL and c-Met promoters as assessed by mutation (Fig. 4D and E), chromatin immunoprecipitation (ChIP) (Supplementary Fig. 2A and C, see section on supplementary data and stemness; NE, neuroendocrine differentiation). (B) Independent analysis to identify master regulators predicted by master regulator analysis (MRA) score, which is $-\log_{10}$ transformed value of significance (P values). (C) Bioinformatics analysis using g:Profiler revealed that bone-related diseases are representative features of RANKL-overexpressing LNCaP cells.
Figure 4

RANKL induces expression of RANKL and c-Met but suppresses AR expression/activation through c-Myc/Max and AP4 transcription factors in the LNCaP cell background. (A) RANKL overexpression upregulates endogenous RANKL and c-Met expression and c-Met phosphorylation but suppresses AR expression. (B) Exogenous RANKL treatment or intrinsic RANKL expression induces endogenous RANKL expression, c-Met expression, and c-Met phosphorylation. Inductions are partly sensitive to OPG or anti-RANKL antibody (denosumab) treatments. (C) RANKL-c-Met signaling increases the level of c-Myc and Max expression. Increased c-Myc and Max nuclear proteins were observed in LNCaP cells treated with RANKL or in LNRANKL cells as evaluated by western blot analysis. Lamin A/C was used as internal control. (D) Activities of human RANKL, (E) c-Met, and (F) AR promoter-luciferase reporter constructs were compared between LNRANKL and LNNeo cells. Site-directed mutagenesis was used to remove the E-box, the cis-element required for c-Myc/Max interaction. Complementary experiments confirmed the results by assaying promoter reporter activities under 10058-F4 (40 µM) exposure, known to inhibit c-Myc function by interference with its dimerization with Max, and by shRNA knockdown of the c-Myc, Max, or AP4 to suppress c-Myc/Max or AP4 binding to its E-box cis-element (*P < 0.05; **P < 0.01; and ***P < 0.001).
has been shown to be a conserved and direct target gene of c-Myc (Cole & McMahon 1999, Jung et al. 2008). The interaction site was identified at the E-box/AP4 binding site (CAGCTG) on the AR promoter region at +374 bp. Deletion of the AP4 site significantly upregulated AR promoter activity in LN\textsuperscript{RANKL} cells. Similarly, interference with c-Myc and Max dimerization with 10058-F4 or genetic ablation of c-Myc, c-Myc and Max, or AP4 using shRNA or siRNA also increased AR promoter activity in LN\textsuperscript{RANKL} cells (Fig. 4F). ChIP analysis further demonstrated direct interaction of c-Myc, Max, and AP4 with the AP4 site within the AR promoter (Supplementary Fig. 2D). AR expression at the RNA and protein level was restored by knocking down c-Myc, c-Myc and Max, or AP4 in LN\textsuperscript{RANKL} cells (Supplementary Fig. 3, see section on supplementary data given at the end of this article).

**RANKL–RANK signaling promotes EMT and stem and neuroendocrine phenotypes**

Consistent with our previous report using ARCaP cells (Zhou et al. 2008), RANKL–RANK signaling also promoted the expression of EMT markers in LNCaP cells. Increased expression of RANKL induced the expression of mesenchymal features, including subtle morphological changes to a more spreading morphology (Supplementary Fig. 4A, see section on supplementary data given at the end of this article), mesenchymal gene expression profiles (increased N-cadherin, and vimentin expression but decreased E-cadherin expression, Supplementary Fig. 4B), and more aggressive cell behaviors (increased cell motility, migration, invasion, and anchorage-independent growth, Supplementary Fig. 4C and D). LNCaP cell-derived RANKL or exogenously administered recombinant RANKL are biologically functional and induce pre-osteoclast maturation as measured by increased TRAP+ staining, when co-cultured with mouse RAW264.7 pre-osteoclasts (Supplementary Fig. 5, see section on supplementary data given at the end of this article). This activity was attenuated by a RANKL decoy receptor, OPG.

RANKL–RANK signaling also promoted stem cell and neuroendocrine differentiation in LNCaP cells. Figure 5A shows increased expression of the stem cell markers CD44, CD133, SOX2, OCT3/4, CD49f, Lin28b, and Nanog. Figure 5B shows elevated expression of the neuroendocrine markers FOXA2, SVP, and chromogranin A. Induced expression of stem cell and neuroendocrine markers was partly blocked by denosumab and OPG, indicating that the maintenance of these phenotypes requires continuous RANKL–RANK signaling (Fig. 5C and D). Consistent with higher intrinsic RANKL expressed by ARCaP\textsubscript{M} cells, we also observed that this cell line expressed higher levels of TFs that regulate EMT, stem, and neuroendocrine phenotypes (Supplementary Fig. 6, see section on supplementary data given at the end of this article). Together, these results are consistent with previous reports showing that cancer cells undergoing EMT expressed stem cell characteristics (Kong et al. 2011). The emergence of an neuroendocrine phenotype in the LNCaP as well as ARCaP\textsubscript{M} cell backgrounds in response to RANKL–RANK signaling has not been reported and is probably due to the activation of TFs such as FoxA2, SOX9, and HIF1\textalpha that regulate the neuroendocrine phenotype of PCa cells (Qi et al. 2010).

**RANK, c-Met, and c-Myc/Max are required for bone colonization by LN\textsuperscript{RANKL} cells**

RANKL–RANK signaling seems obligatory for bone colonization. LN\textsuperscript{RANKL} cells with RANK knockdown (Fig. 6A) formed very small tumors in mice tibias, undetectable by NIR dye-based fluorescence imaging (Yang et al. 2010) but detectable by X-ray (Fig. 6B); moreover, the tumors were less osteolytic and much less bone resorptive compared with bone tumors derived from control shRNA-transduced LN\textsuperscript{RANKL} cells (Fig. 6C). Administration of sRANKL failed to induce bone tumor formation in mice bearing intratibially injected RANK-silenced LN\textsuperscript{RANKL} cells (Fig. 6D). Knockdown of the RANK downstream target TFs, c-Myc/Max, or its effector/target, c-Met, completely abolished the ability of LN\textsuperscript{RANKL} cells to form skeletal or soft tissue metastasis in mice (Fig. 6E). These results support the obligatory role of RANKL–RANK, c-Myc/Max, and c-Met signaling in cancer bone colonization.

**Metastatic LN\textsuperscript{RANKL} cells transform neighboring non-metastatic LN\textsuperscript{Neo} cells**

We tested the ability of $1 \times 10^3$ LN\textsuperscript{RANKL} cells combined with $1 \times 10^6$ non-metastatic $1 \times 10^6$ LN\textsuperscript{Neo} cells to form tumors in mouse skeleton after intratibial injection. While neither LN\textsuperscript{RANKL} nor LN\textsuperscript{Neo} cells when injected individually in these amounts form tumors during a 3-month observation period, tumors arose from the mixed cell populations in <2 months (Fig. 7A). A similar result was observed when LN\textsuperscript{RANKL} cells were injected into the tibia and LN\textsuperscript{Neo-RFP} cells were injected by the intracardiac route (Fig. 7B). IHC analysis revealed that tumors were chimeras consisting of RFP-positive and -negative cells that abundantly expressed RANKL, c-Met, and p-c-Met (Fig. 7C), although parental LN\textsuperscript{Neo-RFP} cells expressed undetectable levels of RANKL, c-Met, and p-c-Met in culture.
We cloned LNNeo-RFP cells from harvested chimeric tumors using the RFP marker and characterized the gene expression profiles of the cultured sublines. Figure 7D shows that tumor-derived LNNeo-RFP cells in culture exhibited patterns of gene expression resembling LNRANKL cells, patterns distinctly different from those of the parental LNNeo-RFP cells. We subsequently labeled both parental and tumor-derived LNNeo-RFP cells with luciferase, and when injected intratibiaally, only the tumor-derived LNNeo-RFP-Luc cells but not the parental LNNeo-RFP-Luc cells colonized the mouse skeleton (Fig. 7E). These results, in aggregate, indicate that LNRANKL cells are able to transform or reprogram neighboring bystander LNNeo-RFP cells to generate tumors in bone.

In summary, these experiments demonstrate that a RANK-mediated signaling network can drive PCa tumor growth in the mouse skeleton. This mechanism involves a feed-forward process in which RANKL and c-Met are induced, and AR is downregulated, as well as a transcriptional network involving c-Myc/Max as an essential node. In this model system, downregulation of RANK, c-Myc/Max, or c-Met expression abolished the bone-homing potential in mice. RANKL-expressing PCa cells were also shown to exhibit transforming potential by altering the gene expression of neighboring bystander cells and coopting them to participate in bone colonization.

Discussion

Hormone-refractory, bone metastatic human prostate and breast cancers are typically lethal; however, little is known...
Abrogation of RANK or its downstream target transcription factors, c-Myc/Max, or its effector/target, c-Met abolishes the metastatic potential of LN^RANKL^ cells. (A) Partial RANK knockdown in LN^RANKL^ cells was confirmed by western blot analysis. (B) Representative NIR fluorescent images showed that RANK-knocked-down LN^RANKL^ cells (n = 10) form no or very small tumors undetectable by NIR fluorescent imaging, but LN^RANKL^ cells transduced with control shRNA (shCon) (n = 10) form obvious intratibial tumors (average total radiant efficiency of NIR ((pph/s)/μW/cm²)): shCon (intraosseous), 4.86 × 10^19 and 3.37 × 10^19; shRANK (intraosseous), 2.63 × 10^19 and 3.84 × 10^19). (C) X-ray and μCT scans showed that LN^RANKL^ cells induced osteolytic lesions in mouse tibia, unlike RANK-knocked-down LN^RANKL^ cells where mouse tibia were intact with minimal detectable bone lesions. (D) Representative in vivo bioluminescent images further demonstrated that administration of recombinant sRANKL (50 μg/kg i.c. twice a week, 2 weeks after intrasosseous inoculation of test cell line) fails to induce intratibial tumor formation of luciferase-tagged RANK-knocked down LN^RANKL^ cells in mice (n = 10) (total flux (photons/s)): shRANK-1 (intraosseous) vehicle, 2.53 × 10^8 and shRANK-1 (intraosseous) + sRANKL, 1.80 × 10^9). (E) Representative in vivo bioluminescent images further demonstrated that luciferase-tagged LN^RANKL^ shCon cells, but not luciferase-tagged LN^RANKL^ cells with RANK (n = 10), c-Met (n = 10), c-Myc (n = 8), or c-Myc/Max (n = 8) knockdown, induce bone and soft tissue metastases after intracardiac injection (average total flux (photons/s)): shCon (intracardiac), 4.60 × 10^8, 9.71 × 10^8, and 4.01 × 10^8; shRANK (intracardiac), 4.27 × 10^8 and 6.13 × 10^8; shMet (intracardiac), 4.52 × 10^8 and 2.25 × 10^8; shMyc (intracardiac), 8.51 × 10^7 and 1.83 × 10^7; and shMyc + shMax (intracardiac), 8.65 × 10^6 and 1.64 × 10^6).
Figure 7
Metastatic LNRANKL cells can transform neighboring non-metastatic LNNeo cells to undergo EMT, express neuroendocrine (NE) and stem cell biomarkers, and form tumors in mouse skeleton. (A) A small population of LNRANKL cells was capable of recruiting non-metastatic LNNeo-RFP cells, which were incapable of forming tumors by themselves, to participate in bone colonization in mice. One thousand, ten thousand, and one hundred thousand LNNeo-RFP cells were co-inoculated with one million LNRANKL-Luc cells in both tibia of nude mice. Tibial tumors and tumors disseminated from bone to soft tissues were harvested and subjected to fluorescent imaging to detect the red fluorescent signal from non-metastatic LNNeo-RFP cells using the Xenogen imaging system at an excitation of 535 nm and emission of 620 nm. Spleens were also harvested and used as negative controls. Total radiant efficiency (%/4μW/cm²) of the RFP in tumors was labeled in white in the corresponding tumors. (B) Representative in vivo bioluminescent and red fluorescent images were also demonstrated with mice bearing intratibial inoculation of either luciferase-tagged LNRANKL cells followed by intracardiac inoculation of LNNeo-RFP cells to test the homing potential of RFP-tagged LNCapRFP cells. (C) IHC and fluorescence images were obtained from chimeric tumors induced in mouse skeleton by inoculating 1000 LNRANKL cells plus 1×10⁶ LNNeo-RFP cells. Representative IHC and fluorescence images of the tumors were merged (200× magnification). Data show co-localization of RFP cells with RANKL, c-Met, and p-c-Met expression in prostate tumors from mouse skeleton. (D) LNNeo-RFP cells harvested from chimeric tumors (tumor LNNeo-RFP) acquired EMT, neuroendocrine, and stem cell properties demonstrated by relative expression of markers detected by qRT-PCR (*P<0.05; **P<0.01; and ***P<0.001). (E) Representative bioluminescent images demonstrated tibial tumor formation induced by luciferase-tagged tumor-derived LNNeo-RFP cells but not by luciferase-tagged parental LNNeo-RFP cells (total flux (photons/s): parental LNNeo-RFP-Luc: 5.77×10⁶; 2.09×10⁷; and 1.56×10⁸ and tumor LNNeo-RFP-Luc: 5.35×10⁷; 9.94×10⁷; and 8.52×10⁷).

phenotypes that promote cancer cell growth in bone (Fig. 3). ii) The effects of RANKL-expressing cells on bystander cells appear permanent, based on our ex vivo culture and profiling of RFP-labeled cells derived from bone metastases. These results support our previous observation that human PCa cells inoculated into immunodeficient mice recruited bystander cells, which underwent genetic modification, as exhibited by their cytogenetic profiles (Pathak et al. 1997). iii) Interaction with pre-osteoclasts promotes osteoclastogenesis, increases bone turnover, and releases soluble and insoluble factors from the skeleton, supporting cancer growth and survival in bone. Our data collectively indicate that autocrine and paracrine RANKL–RANK signaling is critical for the development of bone and soft tissue metastases in mice.

RANKL is expressed by inflammatory T and B cells, osteoblasts, marrow stromal cells, mesenchymal stem cells, and stromal fibroblasts (Thomas et al. 2001). RANKL expression can be regulated by endocrine factors and dietary cholesterol (Sanbe et al. 2009). The functional roles of RANKL depend on the triad relationship of RANKL, RANK, and OPG where RANKL activity is modulated by the presence of RANK receptor and OPG (Ando et al. 2008). The pathophysiological significance of our findings is that RANKL expression is affected by inflammation and significantly augmented by hormonal deprivation (Li et al. 2009) or increased dietary cholesterol (Nuche-Berenguer et al. 2011). Interestingly, cancer bone and soft tissue dissemination is also known to be increased by inflammatory infiltrates in tumors. Increased cancer metastases were also observed in patients with higher dietary cholesterol and hosts undergoing androgen-deprivation therapy (Tan et al. 2011).

Coordinated cell signal network expansion downstream from RANKL–RANK in PCa cells is associated with the invasive phenotype that commonly develops in CRPC patients upon disease progression or under hormonal or chemotherapeutic intervention (Mulholland et al. 2012). Our current model suggests that the RANKL–RANK-mediated signal network downregulates AR and renders the PCa cells more aggressive and metastatic, including the ability to recruit and transform non-metastatic
bystander cells. While the exact mechanisms by which AR downregulation promotes PCa aggressiveness is unknown, it has however been observed that approximately 25% of CRPC patients actually lose AR expression and develop rapid progressive bone metastasis. These patients express low levels of AR and PSA but have been observed to exhibit explosive metastatic disease clinically. Our study raises several important questions that require further study in clinical settings. Can LN\(^{\text{RANKL}}\) cells be identified in the primary, at metastatic sites, or in the systemic circulation and be selectively or more effectively targeted based on their unique phenotype? As androgen deprivation can induce PCa stem cell and neuroendocrine differentiation, can androgen deprivation also induce the resurgence of PCa cells with a phenotype resembling LN\(^{\text{RANKL}}\)? Can the dietary cholesterol, hormonal or inflammatory status of patients be modulated to reduce overall levels of RANKL or RANKL–RANK signaling, to minimize the tumor cell population that initiates PCa metastasis, thus reducing tumor recurrence and distant dissemination? A better understanding of RANKL–RANK signaling in clinical cancer metastasis will improve cancer bone targeting and patient survival.

In summary, our results indicate that RANKL, either derived from the tumor or from the host, plays a key role in cancer bone metastasis. A small population of LN\(^{\text{RANKL}}\) cells was observed to initiate and promote cancer bone and soft tissue metastases by recruiting bystander cells to form tumors in bone. The mechanism underlying this recruitment appears to involve a feed-forward mechanism in which RANKL, RANK, and c-Met expression is increased and AR is downregulated. RANKL alters a large transcriptional program that appears to govern formation of the premetastatic niche as well as emergence of osteomimetic, EMT, and stem and neuroendocrine differentiation. Our results raise a new paradigm beyond the clonal expansion and evolution of cancer cells, where a population of metastasis-initiating PCa cells recruits and activates bystander cells, including ‘dormant’ cancer cells, through RANK-mediated signal amplification.

Supplementary data
This is linked to the online version of the paper at http://dx.doi.org/10.1530/ERC-13-0548.

Declaration of interest
The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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