Pasireotide and octreotide antiproliferative effects and sst\textsubscript{2} trafficking in human pancreatic neuroendocrine tumor cultures

Amira Mohamed\textsuperscript{1,2}, Marie-Pierre Blanchard\textsuperscript{3}, Manuela Albertelli\textsuperscript{4}, Federica Barbieri\textsuperscript{4}, Thierry Brue\textsuperscript{1,5}, Patricia Niccoli\textsuperscript{6}, Jean-Robert Delpero\textsuperscript{7}, Genevieve Monges\textsuperscript{8}, Stephane Garcia\textsuperscript{9}, Diego Ferone\textsuperscript{4}, Tullio Florio\textsuperscript{4}, Alain Enjalbert\textsuperscript{1,2}, Vincent Moutardier\textsuperscript{10}, Agnes Schonbrunn\textsuperscript{11}, Corinne Gerard\textsuperscript{1}, Anne Barlier\textsuperscript{1,2,\*} and Alexandru Saveanu\textsuperscript{1,2,\*}

\textsuperscript{1}Aix-Marseille Université, CNRS, CRN2M-UMR 7286, Faculté de Médecine, Secteur Nord – CS80011, 51, Bd Pierre Dramard, 13444 Marseille Cedex 15, France
\textsuperscript{2}Molecular Biology Laboratory, AP-HM, Conception Hospital, 13385 Marseille, France
\textsuperscript{3}Aix-Marseille Université, CNRS, Plate-Forme de Recherche en Neurosciences PFRN, 13444 Marseille Cedex 15, France
\textsuperscript{4}Department of Internal Medicine and Center of Excellence for Biomedical Research, University of Genova, Genova, Italy
\textsuperscript{5}Endocrinology Department, AP-HM, Timone Hospital, 13385 Marseille, France
\textsuperscript{6}Oncology Department, \textsuperscript{7}Surgery Department, and \textsuperscript{8}Biopathology Department, Paoli Calmettes Cancer Institute, 13009 Marseille, France
\textsuperscript{9}Pathology Laboratory, and \textsuperscript{10}Surgery Department, AP-HM, Nord Hospital, 13015 Marseille, France
\textsuperscript{11}Department of Integrative Biology and Pharmacology, University of Texas, Texas 77225, Houston, USA
\textsuperscript{\*}(A Saveanu and A Barlier contributed equally to this work)

Correspondence should be addressed to A Saveanu or A Barlier
Email alexandru.saveanu@univ-amu.fr or anne.barlier@univ-amu.fr

Abstract

Gastroenteropancreatic neuroendocrine tumors (GEP–NETs) raise difficult therapeutic problems despite the emergence of targeted therapies. Somatostatin analogs (SSA) remain pivotal therapeutic drugs. However, the tachyphylaxis and the limited antitumoral effects observed with the classical somatostatin 2 (sst\textsubscript{2}) agonists (octreotide and lanreotide) led to the development of new SSA, such as the pan sst receptor agonist pasireotide. Our aim was to compare the effects of pasireotide and octreotide on cell survival, chromogranin A (CgA) secretion, and sst\textsubscript{2} phosphorylation/trafficking in pancreatic NET (pNET) primary cells from 15 tumors. We established and characterized the primary cultures of human pancreatic tumors (pNETs) as powerful preclinical models for understanding the biological effects of SSA. At clinically relevant concentrations (1–10 nM), pasireotide was at least as efficient as octreotide in inhibiting CgA secretion and cell viability through caspase-dependent apoptosis during short treatments, irrespective of the expression levels of the different sst receptors or the WHO grade of the parental tumor. Interestingly, unlike octreotide, which induces a rapid and persistent partial internalization of sst\textsubscript{2}, pasireotide did not phosphorylate sst\textsubscript{2} and induced a...
rapid and transient internalization of the receptor followed by a persistent recycling at the cell surface. These results provide the first evidence, to our knowledge, of striking differences in the dynamics of sst2 trafficking in pNET cells treated with the two SSAs, but with similar efficiency in the control of CgA secretion and cell viability.

Introduction

Gastroenteropancreatic neuroendocrine tumors (GEP-NETS) constitute a rare group of tumors, with a significant increase in incidence and prevalence in the past few decades (Oberg 2011). GEP-NETS still raise difficult therapeutic problems despite the emergence of targeted therapies such as rapalogs (inhibitors of the mammalian target of rapamycin; Yao et al. 2011) or tyrosine kinase inhibitors (Raymond et al. 2011). Despite numerous clinical trials testing various strategies, progression-free survival or overall survival remain limited. In this context, it would be of interest to develop relevant preclinical models.

Among all pharmacological tools, the somatostatin analogs (SSA), octreotide and lanreotide, remain the pivotal therapeutic molecules for controlling hormone-related symptoms of functioning GEP-NETS (Rubin et al. 1999, O’Toole et al. 2000, Modlin et al. 2010). However, the initial antisecretory potency of SSA decreases with long-term treatment. The mechanisms potentially involved in this adaptation (or tachyphylaxis) to continuous exposure to SSA are not yet clearly understood (Hofland & Lamberts 2003). These drugs also have antitumor activity, as demonstrated in the PROMID and CLARINET studies (Rinke et al. 2009, Caplin et al. 2014). However, less than 5% of patients have objective radiological tumor regression (Rinke et al. 2009). Moreover, 50% of patients display stabilization of tumor size, but this antitumoral effect concerns NET classified as a ‘good prognosis’ (Baudin et al. 2012).

GEP-NETS express multiple somatostatin (sst) receptors: sst2 (70–100% of tumors; Reubi et al. 2001, Papotti et al. 2002, O’Toole et al. 2006), but also sst1 and/or sst5 and/or sst3. The two commercial SSA, octreotide and lanreotide, are octapeptides that bind with high affinity to sst2 and with moderate affinity to sst5. The failures of these classical SSA have led to the development of new SSA such as pasireotide. This molecule i) binds avidly to sst5 and sst2 and more mildly to sst3 and sst1 (Schmid & Silva 2005), ii) is able to suppress adrenocorticotropic hormone secretion in Cushing’s disease, mainly through sst5 (Boscaro et al. 2009), and iii) induces differential sst2 trafficking, as shown in the heterologous model HEK 293 cells (Pöll et al. 2010) as well as in rat pancreatic Ar42J tumor cells (Kao et al. 2011). Together, these data raise the question of its use in GEP-NETS. In a phase II clinical trial, pasireotide has been shown to control symptoms related to carcinoid syndrome in 27% of patients with NETs who were resistant to octreotide (Kvols et al. 2012). However, the effects of pasireotide in comparison to octreotide on tumor secretion and proliferation and its relationship to WHO grades, sst expression profile, and sst trafficking in GEP-NETS remain to be explored.

For somatotroph adenomas, our group previously demonstrated that sensitivities to SSA are strongly correlated in vivo (in patient) and in vitro (in culture of tumor primary cells from the same patient) (Jaquet et al. 2000). Under these conditions, the use of human tumoral cells in primary culture might be a relevant strategy for evaluating the therapeutic effect of different drugs, particularly of SSA.

In this study, we established and characterized, for the first time, to our knowledge, primary cell cultures of human pancreatic NETs (pNETs). We show that pNET primary cells in culture retained neuroendocrine characteristics such as expression and secretion of chromogranin A (CgA), expression of the different subtypes of sst receptors, and sensitivity to octreotide at concentrations used in vivo. We then used this model to evaluate the effects of pasireotide on cell viability, CgA secretion, and on sst2 phosphorylation and trafficking in comparison with octreotide.

Patients and methods

Patients

This study was approved by the Ethics Committee of the Aix-Marseille University (Aix-Marseille, France) and informed consent was obtained from each patient. Fifteen patients with GEP–NET tumors were included in the study (Table 1). Patients underwent surgery. Fourteen tumors were located in the pancreas (pNETs); one was a hepatic metastasis of a pNET. In 13 cases, there was no clinical or biochemical evidence of hormonal hypersecretion, thus
they were considered as nonfunctional pNETs. In two cases, the tumors were secreting and symptomatic: insulinoma (T4) and gastrinoma (T10), confirmed by pathology report. Determination of Ki67 and proliferation index allowed grade classification (WHO 2010 classification; Table 1).

Pharmacological compounds

The SSAs, octreotide and pasireotide, were provided by Novartis AG. Forskolin and isobutylmethylxanthine (IBMX) were from Sigma–Aldrich and calyculin A from Ozyme (St Quentin les Yvelines, France).

Cell culture

Tumor fragments of human pNETs, obtained after surgery, were dissociated mechanically and enzymatically (Jaquet et al. 1985). The tumoral cells were seeded on 24-, 12-, or four-well plates (according to the experiments) coated with extracellular matrix (ECM) (from bovine endothelial corneal cells as previously described (Jaquet et al. 1985)) required for cell adhesion. The cells were maintained in a culture in d-valine-depleted DMEM (d-valine was replaced by L-valine to block fibroblast proliferation), supplemented with 10% FCS, penicillin (100 U/ml), streptomycin (100 μg/ml), and glutamine (100 U/ml) at 37 °C in a water-saturated atmosphere containing 7% CO2.

Determination of the sst mRNA levels, cell viability, and CgA secretion was carried out on cell cultures of all the tumors. Other experiments were performed according to the quantity of tumoral cells available after dissociation of each tumor. All the experiments were done in d-valine DMEM containing 5% FCS.

Real-time quantitative PCR

Total mRNAs were extracted from 3×10^5 cells using the RNeasy Microkit (Qigene). The receptor mRNAs, sst1, sst2, sst3, and sst5, were detected by real-time quantitative PCR using specific primers and probes as described previously (Saveanu et al. 2001). To produce standard curves, cDNA constructs were produced for each variable, verified by sequencing (Beckman Coulter Ceq 8000 (Beckman Coulter, Fullerton, CA, USA)), and linearized. The mRNA levels of each receptor subtype were normalized to the glucuronidase β (GUSB) mRNA level, as described previously (Saveanu et al. 2001). The results were expressed as copy of mRNA for the sst gene/copy of mRNA for GUSB.

Cell viability

The cells from each tumor were seeded into 24-well plates (4×10^4 cells/well). After 24 h in culture, the cells were treated without or with octreotide or pasireotide at the indicated concentrations for 72 h. The medium was recovered for CgA determination, and cell viability was assayed by a luminescent cell viability assay (CellTiter Glo; Promega) according to the manufacturer’s protocol. Results were expressed as percentages of the value for the control cells (in the absence of the analogs).

CgA secretion

CgA secretion was determined on the medium recovered from cell viability experiments after 72 h of treatment in the presence or absence of the analogs (see above). The recovered culture media were centrifuged at 400 g and stored at −20 °C. CgA was measured using a commercial ELISA Kit (Chromogranin A ELISA Kit, Dako, Les Ulis, France) according to the manufacturer’s protocol. Basal tumoral secretion of the cells from each tumor in culture was reported as U/l CgA per 72 h (Table 2). The effects of drugs on CgA secretion were expressed as percentages of the value for the control group.

---

Table 1 Clinical and immunohistochemical characteristics of 15 human pNETs

<table>
<thead>
<tr>
<th>Tumor (number)</th>
<th>Age (years)</th>
<th>Sex</th>
<th>Tumor (mm)</th>
<th>Ki67 (%)</th>
<th>Grade</th>
</tr>
</thead>
<tbody>
<tr>
<td>T1</td>
<td>53</td>
<td>F</td>
<td>15</td>
<td>&lt;2</td>
<td>G1</td>
</tr>
<tr>
<td>T2</td>
<td>59</td>
<td>M</td>
<td>35^b</td>
<td>3</td>
<td>G2</td>
</tr>
<tr>
<td>T3</td>
<td>67</td>
<td>M</td>
<td>25</td>
<td>90</td>
<td>G3</td>
</tr>
<tr>
<td>T4</td>
<td>27</td>
<td>F</td>
<td>17</td>
<td>1</td>
<td>G1</td>
</tr>
<tr>
<td>T5</td>
<td>69</td>
<td>M</td>
<td>55^b</td>
<td>&lt;1</td>
<td>G1</td>
</tr>
<tr>
<td>T6</td>
<td>65</td>
<td>M</td>
<td>12</td>
<td>4</td>
<td>G2</td>
</tr>
<tr>
<td>T7</td>
<td>64</td>
<td>M</td>
<td>26^b</td>
<td>1</td>
<td>G1</td>
</tr>
<tr>
<td>T8</td>
<td>49</td>
<td>F</td>
<td>45^b</td>
<td>13</td>
<td>G2</td>
</tr>
<tr>
<td>T9</td>
<td>66</td>
<td>F</td>
<td>50</td>
<td>6</td>
<td>G2</td>
</tr>
<tr>
<td>T10</td>
<td>58</td>
<td>F</td>
<td>70^c</td>
<td>&lt;2</td>
<td>G1</td>
</tr>
<tr>
<td>T11</td>
<td>53</td>
<td>F</td>
<td>15</td>
<td>5</td>
<td>G2</td>
</tr>
<tr>
<td>T12</td>
<td>45</td>
<td>M</td>
<td>27</td>
<td>10</td>
<td>G2</td>
</tr>
<tr>
<td>T13</td>
<td>26</td>
<td>F</td>
<td>37^a</td>
<td>1.2</td>
<td>G1</td>
</tr>
<tr>
<td>T14</td>
<td>36</td>
<td>F</td>
<td>49^a</td>
<td>5</td>
<td>G2</td>
</tr>
<tr>
<td>T15</td>
<td>67</td>
<td>F</td>
<td>44</td>
<td>3</td>
<td>G2</td>
</tr>
</tbody>
</table>

Tumor size is expressed as maximal diameter (mm). For primary tumors associated with (a) multiple localization, (b) positive lymph node, (c) hepatic metastasis and positive lymph node, tumor size is the sum of the diameters for each localization. Grade is determined according to WHO classification.
Apoptosis measurement

DNA fragmentation measurement The cells from each tumor were seeded on 14 mm ECM-coated coverslips in four-well tissue culture plates (10^5 cells/well). After 24 h in culture, the cells were treated without or with octreotide or pasireotide (1 nM) for 24 h. The cells were fixed with 1% paraformaldehyde in PBS for 15 min at room temperature. DNA fragmentation was detected by TUNEL with the Apopotag Red In Situ Apoptosis Detection Kit (Millipore, Molsheim, France), according to the manufacturer's protocol. The apoptotic cells were then viewed with a Leica/Leitz DMRB microscope using a PL fluotar 100 objective and scored manually. Results were expressed as the number of TUNEL-positive cells/total cells in 40 successive fields per condition for each cell culture.

Determination of caspase activity The cells from each tumor were seeded into 24-well plates (4 \times 10^4 cells/well). After 24 h in culture, the cells were treated without or with octreotide or pasireotide (1 nM) for 24 h. The activity of the executioner caspases 3 and 7 was measured using the Caspase Glo 3/7 Assay (Promega) according to the manufacturer's protocol. Results were expressed as a percentage of the value for the control conditions.

CAMP assay

The cells from each tumor were seeded into 24-well plates (4 \times 10^4 cells/well). After 24 h in culture, the cAMP concentration was determined using the HTRF-cAMP Femto Kit (CisBio International, Bagnols-sur-Cèze, France), as described previously (Cuny et al. 2012). According to the manufacturer's protocol, the cells were detached using trypsin, seeded onto 96-well plates in suspension, and incubated at 37 °C for 3 h in the presence of 1 μM forskolin and 0.1 μM IBMX (control condition) or combined with 1 nM octreotide or pasireotide. The HTRF assay reagents were then added directly into the 96-well plates. The results were expressed as a percentage of the value for the control conditions.

Immunocytochemistry

The cells from each tumor were seeded on 14 mm ECM-coated coverslips in four-well tissue culture plates (10^5 cells/well). After 24 h in culture, the cells were treated without or with 10 nM octreotide for 72 h and CgA was determined as described in the 'Patients and methods' section. sst receptor mRNAs were determined as described in the 'Patients and methods' section by real-time PCR. –, undetectable. *P<0.05, †P<0.01, and ‡P<0.001.

<table>
<thead>
<tr>
<th>Tumor (number)</th>
<th>Control (U/l)</th>
<th>Octreotide (% inhibition)</th>
<th>CgA secretion</th>
<th>mRNA expression (copies/copy GUSB)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>sst_1</td>
<td>sst_2</td>
</tr>
<tr>
<td>T1</td>
<td>170±5</td>
<td>51±3^‡</td>
<td>0.34</td>
<td>4.94</td>
</tr>
<tr>
<td>T2</td>
<td>49±0</td>
<td>76±5^‡</td>
<td>0.13</td>
<td>0.96</td>
</tr>
<tr>
<td>T3</td>
<td>10±1</td>
<td>60±3^‡</td>
<td>0.04</td>
<td>0.32</td>
</tr>
<tr>
<td>T4</td>
<td>76±1</td>
<td>75±2^‡</td>
<td>0.40</td>
<td>0.16</td>
</tr>
<tr>
<td>T5</td>
<td>16±2</td>
<td>55±1^‡</td>
<td>4.21</td>
<td>3.23</td>
</tr>
<tr>
<td>T6</td>
<td>870±125</td>
<td>73±1^‡</td>
<td>0.28</td>
<td>0.88</td>
</tr>
<tr>
<td>T7</td>
<td>735±33</td>
<td>69±2^‡</td>
<td>1.55</td>
<td>2.35</td>
</tr>
<tr>
<td>T8</td>
<td>21±2</td>
<td>47±1^‡</td>
<td>0.01</td>
<td>0.54</td>
</tr>
<tr>
<td>T9</td>
<td>93±19</td>
<td>86±3^‡</td>
<td>0.40</td>
<td>3.65</td>
</tr>
<tr>
<td>T10</td>
<td>568±21</td>
<td>63±10^‡</td>
<td>0.78</td>
<td>2.67</td>
</tr>
<tr>
<td>T11</td>
<td>569±48</td>
<td>33±1^‡</td>
<td>0.77</td>
<td>2.75</td>
</tr>
<tr>
<td>T12</td>
<td>488±10</td>
<td>88±2^‡</td>
<td>3.51</td>
<td>0.5</td>
</tr>
<tr>
<td>T13</td>
<td>115±7</td>
<td>84±1^‡</td>
<td>3.53</td>
<td>5.45</td>
</tr>
<tr>
<td>T14</td>
<td>691±87</td>
<td>81±4^‡</td>
<td>0.63</td>
<td>0.96</td>
</tr>
<tr>
<td>T15</td>
<td>325±9</td>
<td>29±0^‡</td>
<td>0.74</td>
<td>2.80</td>
</tr>
</tbody>
</table>

The cells were cultures without or with 10 nM octreotide for 72 h and CgA was determined as described in the 'Patients and methods' section. sst receptor mRNAs were determined as described in the 'Patients and methods' section by real-time PCR. –, undetectable. *P<0.05, †P<0.01, and ‡P<0.001.
Probes, Life Technologies, Saint Aubin, France, 1:800 in PBS containing 10% non-immune goat serum) was used as secondary antibody. The coverslips were mounted in ‘ProlonGold’ including DAPI for nuclei staining (Life Technologies).

Confocal images were acquired on a Zeiss LSM780 laser-scanning microscope equipped with a 63× (1, 4 n.a) oil immersion lens with a zoom factor of 1 or 4. The images of sst-Alexa 488 and DAPI-stained cells were respectively obtained using the 488-nm band of an Argon laser and 405 nm diode. Spectral detection of emitted fluorescence was set as follows: 495–535 nm for Alexa 488 and 410–475 nm for DAPI. Images were collected automatically as frame-by-frame z-series. The pinhole was adjusted to the first Airy disk. The pictures presented are single optical slices. Immunocytochemistry experiments were performed on primary cultures from nine pNETs (T2, T3, T4, T6, T7, T10, T13, T14, and T15).

**Image analysis and quantification of the sst2 receptor**

Gray-level confocal images (8-bit) of sst2 staining were analyzed and quantified using ImageJ Software (http://imagej.nih.gov/ij/list.html). For each cell, mean gray values were measured along a five pixel-width line drawn at the cell periphery, considered to be membrane immunostaining. For cytoplasmic staining, gray values of three distinct regions covering most of cytoplasm were determined. Mean gray values were measured on four successive optical slices. Twelve cells were selected at random from each cell culture for each experimental condition. The results were expressed as the mean ± S.E.M. of triplicate determinations as a percentage of the control value.

**Statistical analysis**

Each assay was performed in triplicate. Results are presented as the mean ± S.E.M. of triplicate determinations of a representative tumor (Fig. 1D) or mean ± S.E.M. of at least three tumors. The statistical significance between two paired groups was determined by the Wilcoxon’s non-parametric test and those between two unpaired groups by the Mann–Whitney U nonparametric test. To measure the strength of association between pairs of variables without specifying dependency, Spearman’s rank order correlations were run. The differences were considered statistically significant at P≤0.05.

**Results**

**Establishment and characterization of pNET primary cells in culture**

Fifteen pNETs were successfully dissociated to obtain primary cells in culture. Under our culture conditions, cells from all tumors secreted CgA in a range of 10–870 U/l per 72 h (Table 2). As illustrated in Fig. 1A (T13), positive CgA immunolabeling was observed in all cells, showing that they were neuroendocrine cells.

The sst2 and sst1 mRNAs were expressed in the cells from all tumors. The sst2 expression level was higher than the sst1 level in 80% of tumors (mean mRNA levels, 2.07 ± 0.44 and 1.15 ± 0.36 copy/copy GUSB respectively, Fig. 1B and Table 2). sst3 and sst5 mRNAs were expressed in 73 and 67% of cases respectively (Table 2). The expression levels of the sst receptors did not differ significantly with respect to the WHO grade of the tumor, except for sst5, which was expressed in 100% of grade 1 tumors and only in 37% of
grade 2 tumors (Tables 1 and 2). Despite differences in the quantitative levels of the sst receptors due to the presence of other sst-expressing cell types in the tissue, sst expression profiles were similar to those of the primary tumor tissues (data not shown).

Given that sst2 is the main target of SSA and is not expressed in fibroblasts, expression of the sst2 protein was analyzed by immunocytochemistry in cells from eight pNET tumors (T2, T3, T4, T6, T7, T10, T14, T13, none of which had been pretreated with SSA before surgery). Immunoreactivity of sst2 was found in almost 100% of cells in all of the tumors investigated, as illustrated in Fig. 1C (T2). In the absence of SSA, sst2 immunoreactivity was mainly located on the plasma membrane and to a lesser extent in the cytoplasm (Fig. 1C). This confirmed that, under our culture conditions, an overwhelming majority of cells were neuroendocrine cells, and that fibroblast proliferation was contained.

Finally, the functionality of sst2 was explored by testing the effect of octreotide on CgA secretion. As illustrated in Fig. 1D (T6), octreotide displayed a dose-dependent inhibition of CgA secretion. An octreotide-dependent decrease in CgA secretion was significantly observed in all investigated tumors in a range of 29–88% for 10 nM octreotide (Table 2). There was no correlation between maximal inhibition of CgA secretion and the sst expression levels or the WHO grade of the tumor.

**Effect of pasireotide and octreotide on cell viability**

The effects of octreotide and pasireotide on cell viability were determined for the characterized pNET primary cultures. Both analogs reduced viable cell number in the primary cultures from all 15 tumors (Fig. 2A). Maximal inhibition, observed at 1 or 10 nM for each drug, ranged from 23 to 86% with octreotide and from 16 to 79% with pasireotide.
A significant difference between the effects of octreotide and pasireotide on maximal cell viability inhibition was observed in only two cases: T1 and T3 (Fig. 2A). A strong correlation was observed between the effects of both analogs (Fig. 2B). There was no correlation between the maximal inhibition of cell viability with octreotide or pasireotide and the mRNA levels of the different subtypes of sst. Moreover, pasireotide did not induce better inhibition in tumor cells expressing high levels of sst1, sst5, or sst3.

For two patients, one bearing tumor T2 and the other T13, it was possible to analyze the effect of analogs on two tumoral sites, the primary tumor and a metastatic lymph node for the T2 patient, and two different pancreatic tumors for the T13 patient. In both cases, there were no significant differences in cell viability inhibition for the two tumoral sites, either with octreotide or pasireotide (data not shown).

To determine the mechanism underlying the reduction in the number of viable cells induced by octreotide and pasireotide, DNA fragmentation was investigated using a TUNEL assay on primary cultures from three tumors (T9, T11, and T13). Both octreotide and pasireotide increased the number of apoptotic events (Fig. 2C), indicating an apoptotic mechanism. The apoptotic effect of octreotide and pasireotide was further investigated by measuring the activity of the executioner caspases. Basal caspase activity in untreated cells was significantly increased by octreotide and pasireotide in all the primary cultures from six tumors (201.3±17.7 and 182.1±36.3%, respectively, P<0.03; Fig. 2D; T1, T3, T7, T8, T13, and T15). So both analogs induced caspase-dependent apoptotic cell death in pNET primary cells.

**Effect of pasireotide and octreotide on CgA secretion**

The antisecretory effect of pasireotide was then determined and compared with that of octreotide. Pasireotide dose-dependently decreased CgA secretion, similarly to octreotide, in the primary cultures from all the tumors (Fig. 3A). Although the decrease in cell number observed under SSA treatment, as described above, potentially has an effect on CgA secretion, inhibition of CgA secretion by octreotide and pasireotide was still observed after ‘normalization’ of CgA secretion data against cell viability. Under these conditions, CgA secretion represents 36±7 and 41±5% (n=15) of the value for controls for 1 nM SSA treatment respectively, indicating direct effects of SSA on CgA secretion mechanisms. A good correlation was observed between maximal inhibitions of CgA secretion under...
pasireotide and octreotide (Fig. 3B). No correlation was observed between maximal inhibitions of CgA and the mRNA levels of the different subtypes of sst. Moreover, there was no correlation between the inhibitions observed on cell viability and on CgA secretion for each analog (not shown).

Given that cAMP is one of the main molecules involved in the inhibition of hormonal secretion induced by SSA (Hofland & Lamberts 2003), the effect of octreotide and pasireotide on the intracellular cAMP levels was investigated using primary cultures from three tumors (T5, T9, and T10). Octreotide and pasireotide similarly reduced the cAMP level (by 49 ± 2 and 50 ± 6%, respectively, Fig. 3C), in agreement with the similar antisecretory effect observed for the two analogs on the pNET primary cells.

### Time course of sst2 internalization and recycling under pasireotide and octreotide

The time course of sst2 trafficking under pasireotide and octreotide treatment was then investigated by sst2 immunolabeling in pNET primary cells. In primary cultures from seven tumors (T2, T3, T6, T7, T10, T14, and T13), sst2 internalization was rapidly observed after 5 min of

**Figure 4**

Differential time course of sst2 internalization with pasireotide and octreotide. Primary cells in culture (T2) were incubated without (A) and with 1 nM octreotide (B, C and D) or 1 nM pasireotide (E, F and G) for the time indicated. sst2 is shown in green and nuclei are stained in blue (DAPI), zoom factor 4, Scale bars: 5 μM. (H) Quantification of membrane sst2 immunostaining in T2 and T14, as described in the ‘Patients and methods’ section, in the presence of octreotide (white bars) or pasireotide (black bars) for the time indicated. Results are expressed as mean ± S.E.M. as a percentage of the level of control immunostaining in each experimental condition. *P < 0.05; **P < 0.01; ***P < 0.001; NS, not significant. Membrane sst2 immunostaining under control conditions is 71 ± 3% of total sst2 immunostaining.
octreotide treatment, as illustrated in Fig. 4B (T2), and was maintained in the cytosol up to 30 min (Fig. 4C and D; T2). The quantification of sst2’s subcellular localization showed that sst2 labeling at the cell surface progressively decreased and reached 66 ± 2% of control sst2 immunostaining level after 30 min exposure to octreotide (Fig. 4H; T2 and T14).

Five minutes of pasireotide treatment also induced sst2 internalization, as illustrated in Fig. 4E (T2). In contrast, after 15 min of pasireotide treatment, sst2 immunolabeling was mainly observed at the cell surface (Fig. 4F and G; T2). The quantification of sst2’s subcellular localization in the presence of pasireotide showed that sst2 labeling at the cell surface represented 67 ± 3% of the control level of sst2 immunostaining after 5 min of pasireotide exposure and progressively recovered to the control level (Fig. 4H; T2 and T14), indicating membrane recycling of the receptor.

To investigate whether the differential sst2 localization observed under octreotide/pasireotide persisted after long-term exposure, immunocytochemical analysis of sst2 was carried out after 1 and 24 h SSA treatment. After 1 and 24 h of treatment with pasireotide, the sst2 receptor was mainly located at the cell surface (Fig. 5F and G; T10). In contrast, with octreotide treatment, sst2 distribution was similar to that observed at 5–30 min, still mainly in the cytosol (Fig. 5C and D; T10).

**Time course of sst2 phosphorylation with pasireotide and octreotide**

Finally, sst2 phosphorylation, which is strongly involved in receptor trafficking, was investigated in pNET primary cells, using an antibody specific for the sst2 receptor phosphorylated at Ser341/343 (psst2). In untreated cells, no psst2 immunostaining was detected (Fig. 6A; T15) whereas sst2 receptor immunostaining was present at the cell surface (Fig. 6B; T15). After 5 and 30 min exposure to 1 nM octreotide, clusters of psst2 were detected, both at the cell surface and in the cytosol (Fig. 6C and D; T15). In contrast, no phosphorylation of the sst2 receptor was detected after 5 and 30 min exposure to 1 nM pasireotide (Fig. 6E and F; T15).

**Discussion**

For the first time, we have established and characterized primary cell cultures of human GEP–NETs. Tumoral cells of pNETs have a slow growth rate *in vitro* and low adhesion to plastic or the available commercial matrix (A Mohamed, 2011 unpublished observations). Using bovine ECM, we obtained monolayers of neuroendocrine cells that retain CgA expression and secretion, expression of the different subtypes of sst receptors, and sst2.
functionality. The sst expression patterns in the primary cells of pNETs were similar to those previously found in GEP–NET tumor tissues by our group (O’Toole et al. 2006) and by others (Reubi et al. 2001, Papotti et al. 2002, Ono et al. 2007, Srirajaskanthan et al. 2009, Kim et al. 2011): sst2 remains the primary sst expressed, sst1 is the secondary subtype expressed, both in terms of percentage of positive tumors and of quantitative mRNA level.

In the human pNET BON1 cell line, which expresses sst2, the antiproliferative effect of octreotide is not clearly observed in vitro (BON cells in culture) or in vivo (BON cell xenografts) (Moreno et al. 2008). Concentrations of octreotide above 1 μM are required to produce small amounts of growth inhibition (Georgieva et al. 2010). In contrast, in the pNET primary cultures described herein, the concentrations of octreotide (1 nM) needed to decrease cell viability and CgA secretion were similar to those found in the plasma of treated patients (from 1 to 10 nM for octreotide (Woltering et al. 2008) and about 8 nM for pasireotide (Kvols et al. 2012)). Note that pasireotide induces growth inhibition with an IC50 1000-fold superior in the BON1 cell line (Somnay et al. 2013). In our study, both analogs decreased cell viability by caspase-dependent apoptosis and repressed cAMP levels, which could be involved in the inhibition of CgA secretion.

In all the primary cultures from the 15 human pNETs studied, pasireotide decreased cell viability and CgA secretion. Pasireotide has a 40-, 30-, and fivefold higher binding affinity than octreotide for sst5, sst1, and sst3, respectively and a 2.5-fold lower affinity for sst2 (Schmid 2008). In pNET cells, despite the expression of sst1, sst3, and sst5, pasireotide was not more efficient than octreotide. Similar inhibitory effects on cell viability and CgA secretion were observed for pasireotide and octreotide at concentrations found in the plasma of treated patients (from 1 to 10 nM for octreotide (Woltering et al. 2008) and about 8 nM for pasireotide (Kvols et al. 2012)). Note that pasireotide induces growth inhibition with an IC50 1000-fold superior in the BON1 cell line (Somnay et al. 2013). In our study, both analogs decreased cell viability by caspase-dependent apoptosis and repressed cAMP levels, which could be involved in the inhibition of CgA secretion.
indicate that under our experimental conditions, pasireotide acts mainly through sst2 (which is the most expressed receptor) in pNET primary cells in culture. To our knowledge, no comparison has been made of the effects of pasireotide and octreotide in GEP–NET patients naïve to SSA treatments.

We used pNET cells in culture to explore the dynamics of sst2 trafficking and phosphorylation under octreotide and pasireotide treatment. In untreated cells, sst2 was not phosphorylated and was mainly located at the cell surface. A clinically relevant concentration of octreotide (1 nM) induced a rapid and persistent partial sst2 internalization associated with its phosphorylation on Ser341/343. These results are consistent with observations showing that excised GEP–NET tumors of untreated patients display exclusively nonphosphorylated expression of sst2 on the cell surface, whereas tumors of octreotide-treated patients contain Ser341/343 phosphorylated sst2, predominantly in the cell surface, whereas tumors of octreotide-treated patients contain Ser341/343 phosphorylated sst2, predominantly internalized (Reubi et al. 2010, Waser et al. 2012). In the same way, we show for the first time, to our knowledge, in pNET cells in culture, striking differences in sst2 trafficking under a clinically relevant concentration of pasireotide (1 nM). Pasireotide induced a rapid and transient internalization of sst2 followed by a persistent re-expression of the receptor at the cell surface. Surprisingly Ser341/343 phosphorylation was not observed under pasireotide even at 5 min. This could be explained by massive and rapid recycling of the receptor, which requires its dephosphorylation during internalization before reinsertion into the plasma membrane. The lack of Ser phosphorylation (critical for sst2 desensitization/uncoupling; Liu et al. 2008, Kao et al. 2011) indicates that the level of active receptors at the cell surface is greater with chronic pasireotide than with octreotide.

The differential patterns of sst2 internalization induced by pasireotide and octreotide have been shown in the HEK 293 cell line transfected either with human sst2 (hsst2; Lesche et al. 2009) or with rat sst2 (rsts2; Pöll et al. 2010). In the study of Lesche and colleagues, pasireotide was less potent than octreotide in inducing hsst2 internalization and required 30 min exposure to 1 μM pasireotide. In the study of Pöll and colleagues, rsts2 internalization is not observed after 5-min exposure to 10 μM pasireotide. Moreover, in contrast to octreotide, a lack of sst2 internalization with pasireotide has also been observed in the rat pituitary and pancreas in vivo after 90-min exposure (Pöll et al. 2010). A close correlation between the phosphorylation levels of a cluster of four threonine residues, within the cytoplasmic C-terminal region of sst2, and sst2 internalization has been established under pasireotide and octreotide treatments in rsts2-transfected HEK 293 cells: octreotide but not pasireotide is able to increase threonine phosphorylation and induces sst2 internalization (Pöll et al. 2010). It has similarly been shown that pasireotide is less effective than sst (ss14) in inducing endogenous sst2 internalization in pancreatic Ar42J cells or in rsts2 receptor-transfected CHOK1, and this lesser internalization is associated with less phosphorylation of both Ser341/343 and Thr353/354 (Kao et al. 2011). In addition, in that study, arrestins were less recruited to the membrane and formed more unstable complexes with sst2 under pasireotide than under ss14. Moreover sst2 was more rapidly dephosphorylated and recycled after pasireotide treatment than ss14 treatment (Kao et al. 2011). It has also been shown that GRK2 is required for ss14-stimulated sst2 phosphorylation on Ser341/343 (Liu et al. 2009) and that GRK2/GRK3 are required for ss14-dependent phosphorylation of sst2 on Thr353/354 (Pöll et al. 2010). Furthermore, overexpression of GRK2 increases pasireotide-dependent phosphorylation and internalization of sst2 (Pöll et al. 2010). Recently, GRK2 and β-arrestin 1 mRNA levels were found to be correlated with octreotide response in human GH adenomas (Gatto et al. 2013). Therefore besides the determination of the ss14 expression profiles, assessment of GRK and arrestin levels would be of interest for predicting the extent of sst2 internalization/recycling and biological effects with SSA therapy.

Taken together, our results on primary cultures of pNET cells provide strong evidence for the existence of differential patterns of sst2 phosphorylation and internalization/recycling with clinically relevant concentrations of octreotide and pasireotide, as previously demonstrated in sst2-transfected heterologous cell lines and in a rat pancreatic tumor cell line endogenously expressing the sst2 receptor (recently reviewed by Reubi & Schonbrunn (2013)).

The differential sst2 trafficking in pNET primary cells during treatment with pasireotide and octreotide observed in this study, does not initiate different biological effects. Although sst2 internalization/desensitization is greater in the presence of octreotide, the decreased number of active receptors at the cell surface might be compensated for by the different affinity of octreotide for sst2 and for the other ss1 receptors, relative to the affinity of pasireotide. Moreover, independently of the differential sst2 trafficking, both analogs could recruit the same downstream signaling cascades. These might account for the similar biological effects we observed for the two analogs over the timescale studied.

Octreotide can control hormonal-related symptoms in patients initially. However, some responsive tumors escape after prolonged treatment and become resistant
to octreotide (Modlin et al. 2010). Although clinical observations of tachyphylaxis in GEP–NET during octreotide treatment are well established, the underlying mechanisms are still elusive. Preliminary results of the phase III study conducted to compare pasireotide LAR with octreotide LAR in patients inadequately controlled by conventional doses of SSA indicate that long-term GEP–NET treatment with pasireotide LAR may be of benefit (Clinical Trial number NCT00690430, unpublished results (Wolin et al. 2013)). Under these conditions, it would be of interest to explore the localization and the phosphorylation status of sst2 in tumors of pasireotide-treated patients, as has been previously performed for octreotide (Reubi et al. 2010, Waser et al. 2012), to confirm or rebut our observations on pasireotide-persistent membrane expression of sst2 under continuous treatment.

Escape from response to octreotide observed over months or years is unlikely to be due to rapid uncoupling or internalization of the receptor (Hofland & Lamberts 2003, Reubi & Schonbrunn 2013). It is rather more likely that it could be due to changes in the expression or recruitment of crucial signaling partners downstream of the sst2 receptor or change in proteins that regulate receptor function (Reubi & Schonbrunn 2013). Recently, Li et al. (2012) have shown that 16 months of continuous exposure to octreotide still decreased cell growth of CNDT2.5 (a small intestine NET-derived cell line) and progressively upregulated several genes involved in growth/differentiation and cell signaling. Therefore, tachyphylaxis might be associated with the downregulation of such genes. How the dynamics of the receptor, encoded by chronic agonist binding, could be involved in such processes requires further investigations.

In conclusion, the experiments described in this study are the first investigation, to our knowledge, of the role of pasireotide and octreotide in primary culture of human pNET cells. It is shown that at clinically relevant concentrations, pasireotide is at least as efficient as octreotide for repressing hormonal secretion and cell viability despite the striking difference in sst2 trafficking between pasireotide and octreotide treatment in pNET cells. Finally, pNET cells in primary cultures could be considered as a relevant preclinical model i) to improve our understanding of sst receptor pharmacology and ii) to study biological effects in development of new drugs.

Declaration of interest
The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

Acknowledgements
The authors thank Anne Laure Germanetti for technical advice on qRT-PCR studies and Gary Burkhart for his help with correcting the English.

References

Funding
This work was supported by Centre National pour la Recherche Scientifique (UMR 7286), Aix-Marseille Université, Association pour le Développement des Recherches Biologiques et Médicales au Centre Hospitalier Régional de Marseille (A.D.R.E.M.), Protisvalor Méditerranée (Aix-Marseille Université), and Novartis Pharma SAS. A Mohamed was a recipient of a fellowship from the A.D.R.E.M. (2010/2011). This study was partially supported by Novartis Pharma SAS.

Author contribution statement
The authors have made the following declarations about their contributions: A Mohamed: set-up of GEP–NETs primary cultures, pharmacological studies, chromogranin A, cell viability assessment, and immunocytochemistry studies; M-P Blanchard: immunocytochemistry studies by confocal microscopy; M Albertelli and F Barbieri: set-up of primary culture of GEP–NETs; T Brue, P Nicoli, and J-R Delpero: correlation between clinical data and in vitro functional data, scientific discussions; G Monges and S Garcia: immunohistochemical analysis of the tumor and WHO grade determination; D Ferone and T Florio: set-up of primary culture of GEP–NETs, establishment of the project and scientific discussions; V Moutardier: correlation between clinical data and in vitro functional data and scientific discussions; A Enjalbert: data analysis and scientific discussions, A Schonbrunn: set-up of sst2, antibody experiments and scientific discussions; C Gerard: data analysis, scientific discussions, and drafting of the paper; A Barlier: scientific direction of the project, data analysis, and drafting of the paper; and A Saveanu: scientific direction of the project, real-time PCR of sst, data analysis, scientific discussions, and drafting of the paper.


Received in final form 10 June 2014
Accepted 8 July 2014
Made available online as an Accepted Preprint
10 July 2014