Procalcitonin for detecting medullary thyroid carcinoma: a systematic review

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Abstract

The aim of the present study was to perform a systematic review of published studies to provide a robust estimation of the use of procalcitonin (ProCT) as a diagnostic marker of medullary thyroid carcinoma (MTC), with particular focus on its specificity and negative predictive value in excluding MTC. A comprehensive computer literature search was conducted to find relevant published articles on the topic. We used a search algorithm based on a combination of the terms ‘medullary,’ ‘thyroid,’ and ‘ProCT.’ The search was updated until February 2015. To expand our search, references of the retrieved articles were also screened. A total of 39 articles were retrieved, of which nine original papers published from 2003 to 2014 were selected for the review. Some of these studies used ProCT in the preoperative diagnosis of MTC, whereas others measured ProCT during the follow-up of patients who had been previously treated for MTC. Other laboratory measurements were performed in some of the included studies. The results of the majority of the studies indicate that ProCT measurement appears to be a very promising and reliable serum marker for the diagnosis of MTC, and it is not inferior to calcitonin (CT). The sample handling is less laborious, and in the few CT-negative cases reviewed, the assay had even greater sensitivity. It would be worthwhile to establish cutoff levels using larger patient series, because we speculate that this assay could potentially replace CT measurement in the future.

Key Words
- medullary thyroid cancer
- calcitonin
- procalcitonin
- thyroid nodule

Introduction

Medullary thyroid carcinoma (MTC) is a malignancy that originates from thyroid parafollicular C cells and accounts for about 5% of thyroid cancers (Kloos et al. 2009). In the majority of cases (i.e., four out of every five), this cancer occurs as a sporadic tumor, whereas the remaining ones are part of familial disorders (Kloos et al. 2009). Diagnosing MTC is still a challenge in clinical practice. Cytological examination by fine needle aspiration (FNAC) has a very low detection rate for MTC (Pusztaszeri et al. 2014, Trimboli et al. 2015); ultrasonography does not achieve high reliability in this context (Trimboli et al. 2014); and finally, the measurement of serum calcitonin (CT) should represent the most accurate tool for detecting MTC, but its use in patients with thyroid lesions is still a matter of...
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Routine determination of CT is highly debated (Kloos et al. 2015). For these reasons, a non-negligible quantity of MTC cases have been described (Trimboli & Giovanella 2009). Thus, more tools for preoperatively diagnosing MTC are required in order to better plan the initial treatment of patients; furthermore, there is a great need for accurate markers with high specificity and negative predictive value to exclude MTC in patients with thyroid nodules and to carefully manage MTC patients during follow-up.

Recently, the determination of ProCT, a precursor of CT, has been investigated in the diagnosis and management of MTC. ProCT is a 116 amino acid peptide that originates from the C cells of the thyroid gland, and it is a very stable protein that features a concentration-independent in vivo half-life of 20–24 h (Trimboli & Giovanella 2015). In previous studies better performance of ProCT in comparison with CT have been recorded. Specifically, when the same antibodies were used, all of the commercial ProCT assays yielded similar results. In addition, ProCT samples do not need to be kept cold on ice during the entire process chain and are easier to manage in clinical practice. These characteristics give ProCT great potential to replace CT as a marker for detecting and managing MTC patients. To date, no review on this topic has, to our knowledge, been published.

The aim of the present study was to perform a systematic review of published studies to provide a robust estimate of using ProCT as a diagnostic marker of MTC for both preoperative diagnosis and recurrence detection. In particular, the present study focused on ProCT specificity and negative predictive value in excluding MTC.

Materials and methods

We searched studies that reported the measurement of ProCT in preoperative and/or postoperative diagnoses of MTC. A comprehensive computer literature search of the PubMed/MEDLINE, Embase, and Scopus databases was conducted to find published articles on this topic. The search algorithm was based on the combinations of the terms ‘medullary’ AND ‘thyroid’ AND ‘ProCT.’ We did not use a beginning date limit, and the search was updated until February 18, 2015. To identify additional studies and expand our search, references of the retrieved articles were also screened. All original articles that described serum ProCT in MTC were eligible for inclusion. Case reports were initially excluded. The authors of the present study independently reviewed the retrieved articles. For each included study, information was abstracted concerning the study data (including authors, year of publication, journal of publication, type of article, and study design).

Results

Eligible articles

The comprehensive computer literature search revealed 39 articles. The titles and abstracts of these articles were reviewed, and nine original papers were selected; their full-text versions were then retrieved, and their references lists were scanned (Bihan et al. 2003, Bolko et al. 2003, Algeciras-Schimnich et al. 2009, Kaczka et al. 2010, 2012, Walter et al. 2010, Kratzsch et al. 2011, Giovanna et al. 2013, Machens et al. 2014). These papers were published from 2003 to 2014, and they were all original articles (Table 1). Almost all of the studies were from European researchers (Bihan et al. 2003, Bolko et al. 2003, Kaczka et al. 2010, 2012, Walter et al. 2010, Kratzsch et al. 2011, Giovanna et al. 2013, Machens et al. 2014), and one was conducted by US investigators (Algeciras-Schimnich et al. 2009). Some of these studies used ProCT (Bihan et al. 2003, Giovanna et al. 2013, Machens et al. 2014) in the preoperative diagnosis of MTC, whereas the remaining ones (Bihan et al. 2003, Bolko et al. 2003, Kaczka et al. 2010, 2012, Walter et al. 2010, Kratzsch et al. 2011) measured ProCT during follow-up of patients who had been previously treated for MTC. Additional laboratory investigations were performed in some of the included studies. For CT measurement, different assays were employed and different reference values were adopted. Preoperative ProCT stimulation by pentagastrin was performed in two studies (Kratzsch et al. 2011, Giovanna et al. 2013). The different characteristics between the studies (i.e., objective, ProCT cutoff level, etc.) did not allow a meta-analysis.

Use of ProCT for the postoperative detection of recurrent MTC

Seven papers (Bihan et al. 2003, Bolko et al. 2003, Algeciras-Schimnich et al. 2009, Kaczka et al. 2010, 2012, Walter et al. 2010, Kratzsch et al. 2011) reported ProCT measurement in MTC patients during follow-up after surgery. A total of 265 MTC cases, of which 114 involved cured patients and 151 were recurrent/persistent cases,
were reported. The most relevant paper was published by Algeciras-Schimnich et al. (2009); it collected data on a series of 133 MTC cases (42 cured and 91 active), and 83 out of 91 (91.2%) of the recurrent MTCs had ProCT of over 0.15 ng/ml, whereas none of the cured MTCs had positive ProCT (specificity 100%). Among the active MTCs, those with stable disease had the lowest mean ProCT levels (3.6 ng/ml), patients with recurrent/metastatic disease had the highest mean value (241.7 ng/ml), and patients with newly diagnosed cancer had intermediate levels (13.8 ng/ml). The ProCT concentration that gave the best diagnostic accuracy was 0.16 ng/ml, which corresponds to the upper reference limit for normal subjects (≤0.15 ng/ml). The authors concluded that ProCT is a promising complementary MTC tumor marker because of its great analytical stability. An important contribution was also made by the Kratzsch et al. (2011) study. Those authors compared ProCT to several CT assays (Immulite, Liaison, and IRMA Medipan) in different groups of patients with hypercalcitoninemia, including: subjects affected by recurrent/persistent MTC, those with chronic kidney disease (CKD), and those with Hashimoto’s thyroiditis. The ten MTCs in that study had CT levels that ranged from 18 to 1511 pg/ml, and ProCT levels were between 0.226 and 11.6 ng/ml. The authors found the highest CT and ProCT values in CKD patients and showed a diagnostic sensitivity for MTC of 100% with ProCT and IRMA-CT but only 90 and 80% for Immulite_CT and Liaison-CT respectively. A ProCT value of less than 0.25 ng/ml could exclude MTC. Of relevance, ProCT concentrations appeared to show better stability than CT levels did. In addition, Kaczka et al. (2010, 2012) reported two series, the first of which (Kaczka et al. 2010) recorded undetectable (<0.1 ng/ml) ProCT in 20 out of 23 MTC patients in remission (87% specificity) and detectable (range 0.63–5.52 ng/ml) ProCT in persistent MTC (100% sensitivity). Later, these Polish authors evaluated ProCT in four active MTC cases after surgery, in two MTC patients before and after surgery, and in 23 other MTC patients in remission (Kaczka et al. 2012); their most relevant findings were that 20 out of 23 (87%) disease-free patients had undetectable (<0.1 ng/ml) ProCT levels, and the remaining three had 0.15, 0.16, and 0.27 ng/ml. On the other hand, CT measurement in these patients showed undetectable (<5 pg/ml) values in 15 cases, values between the lower reference limit and 10 pg/ml in five cases, and levels above 10 pg/ml in three patients. Also, the mean ProCT concentration was 3.5 in the active MTC group and 0.06 ng/ml in the MTC patients in remission. A study by Walter et al. (2010) evaluated 69 MTC patients during follow-up, and ProCT was detectable (i.e., >0.06 ng/ml) in 67 cases; the best cutoff for discriminating MTC from the control was 0.5 ng/ml (84% sensitivity and specificity), whereas the threshold for distinguishing MTC from C-cell hyperplasia was 0.16 ng/ml (59% sensitivity and 100% specificity). Also, the ProCT:CT ratio was as an accurate predictor of progression-free survival and was correlated with the clinical outcome. Finally, two older papers should also be cited. Very high sensitivity (100%) of ProCT in detecting recurrent MTC was also recorded by Bolko et al. (2003). They used a sandwich assay with anti-catacalcin antibody as the marker and anti-CT antibody as the solid phase; therefore, the results were not perfectly comparable to those in more recent studies. A study by Bihan et al. (2003) used a chemiluminescent assay to detect both intact ProCT molecule and CT-conjoined C-terminal cleavage peptide1 (CCP1), but the assay did not distinguish between

### Table 1 Main characteristics of the studies included in the systematic review

<table>
<thead>
<tr>
<th>First author</th>
<th>Year</th>
<th>Country</th>
<th>Study aim</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bolko</td>
<td>2003</td>
<td>Poland</td>
<td>To evaluate PCT in patients followed-up for MTC</td>
</tr>
<tr>
<td>Bihan</td>
<td>2003</td>
<td>France</td>
<td>To evaluate the clinical utility of CT precursor serum levels</td>
</tr>
<tr>
<td>Algeciras-Schimnich</td>
<td>2009</td>
<td>USA</td>
<td>To evaluate PCT in patients followed-up for MTC and in patients with other conditions</td>
</tr>
<tr>
<td>Kaczka</td>
<td>2010</td>
<td>Poland</td>
<td>To evaluate PCT in patients followed-up for MTC and in patients with goiters at histology</td>
</tr>
<tr>
<td>Walter</td>
<td>2010</td>
<td>Switzerland–The Netherlands</td>
<td>To evaluate PCT in patients followed-up for MTC</td>
</tr>
<tr>
<td>Kratzsch</td>
<td>2011</td>
<td>Germany</td>
<td>To test if PCT could be a marker of MTC after surgery</td>
</tr>
<tr>
<td>Kaczka</td>
<td>2012</td>
<td>Poland</td>
<td>To evaluate PCT in patients followed-up for MTC</td>
</tr>
<tr>
<td>Giovannella</td>
<td>2013</td>
<td>Switzerland</td>
<td>To evaluate PCT in diagnosing and excluding MTC in thyroid nodule patients</td>
</tr>
<tr>
<td>Machens</td>
<td>2014</td>
<td>Germany</td>
<td>To evaluate PCT in diagnosing and excluding MTC in thyroid nodule patients</td>
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</tbody>
</table>

PCT, procalcitonin; CT, calcitonin; MTC, medullary thyroid carcinoma.
the two; therefore, unfortunately, the ProCT results were not specified. All of the results for ProCT in recurrent and cured MTC cases are shown in Table 2.

Use of ProCT for the preoperative diagnosis of MTC in patients with thyroid nodules

Two studies (Giovanella et al. 2013, Machens et al. 2014) aimed to evaluate ProCT as a marker for the initial diagnosis of MTC in a series of thyroid nodule patients. All of the patients underwent surgery and had a final histological diagnosis of MTC. Relevant data about the prognoses of the patients was described in one of the papers (Machens et al. 2014). In that study, Machens and colleagues evaluated 457 consecutive patients with previously untreated MTC, 112 of whom had ProCT and CT serum levels that were determined before the initial surgery. Receiver operating characteristic analyses revealed similar diagnostic accuracy for ProCT and CT and yielded similar areas under the curve for primary tumors, extrathyroidal extension, number of lymph nodes involved, and distant metastasis. However, ProCT levels were only correlated with the number of lymph nodes involved and the distant extent of disease. Also, 107/112 cases had ProCT levels of more than 0.1 ng/ml, with a sensitivity of 95.5%. As the ProCT levels increased, the biochemical cure rates declined (71% for 1 ng/ml; 36% for 5 ng/ml; 23% for 10 ng/ml; 10% for 50 ng/ml). In another prospective study (Giovanella et al. 2013), both CT and ProCT were measured in a consecutive series of 1236 patients’ thyroid nodules, and 14 cases displayed increased (i.e., above 10 pg/ml) calcitonin levels and were selected for pentagastrin stimulation; upon histology, two MTCs were recorded, and they had basal CT of over 100 pg/ml and ProCT of >0.1 ng/ml. The main results of that study were that basal and pentagastrin-stimulated CT had some false-positive results, whereas all of the patients without MTC had undetectable levels of both basal and stimulated ProCT (100% positive predictive value (PPV) and 100% negative predictive value (NPV)). The authors recommended measuring ProCT and avoiding CT stimulation in patients with moderately elevated (i.e., between 10 and 100 pg/ml) basal CT.

Discussion

Since its first description (Horn & Ravdin. 1951), diagnosing MTC has been a challenge (Pusztaszeri et al. 2014, Trimboli et al. 2014, 2015). Calcitonin, a 32-amino-acid monomeric peptide that results from the processing of ProCT, has traditionally been reported as the most accurate serum tool for identifying MTC both before and after surgical treatment. However, some relevant limits of CT measurement in clinical practice have affected its accuracy (Costante et al. 2007, Cheung et al. 2008). CT belongs to the calcitonin gene-related peptide (CGRP) superfamily and is generated by the alternative splicing of the calcitonin gene (CALCI). Calcitonin is the main peptide product of thyroid C cells, but alternative expression of the gene results in the formation of CGRP in neural tissue. The biosynthetic secretory pathway for CT involves a complex series of progressive modifications: after the biosynthesis and folding of precursors, subsequent proteolytic processing occurs both within the Golgi apparatus and later within the secretory granules because of the actions of prohormone convertase (PC) enzymes. The precursor of CT is pre-procalcitonin (Pre-ProCT), and it contains 141 amino acids with a signal peptide of 25 residues. Pre-ProCT (1–116) is processed to mature calcitonin, amino acids 60–91, and to CCP1, amino acids 96–116. Figure 1 depicts and summarizes the process that leads to CT production. In the pre-analytical phase, the marked variation of CT that occurs during the day because of its pulsative secretion may be

<table>
<thead>
<tr>
<th>First author</th>
<th>MTC (n)</th>
<th>ProCT Positive</th>
<th>Sensitivity</th>
<th>ProCT MTC (n)</th>
<th>Negative</th>
<th>Specificity</th>
</tr>
</thead>
<tbody>
<tr>
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<td>10</td>
<td>10</td>
<td>100</td>
<td>14</td>
<td>8</td>
<td>57.1</td>
</tr>
<tr>
<td>Algeciras-Schimnich</td>
<td>91</td>
<td>83</td>
<td>91.2</td>
<td>42</td>
<td>42</td>
<td>100</td>
</tr>
<tr>
<td>Kaczka</td>
<td>9</td>
<td>9</td>
<td>100</td>
<td>23</td>
<td>20</td>
<td>87.0</td>
</tr>
<tr>
<td>Walter</td>
<td>34</td>
<td>ND</td>
<td>84.0</td>
<td>35</td>
<td>ND</td>
<td>84.0</td>
</tr>
<tr>
<td>Kratzsch</td>
<td>10</td>
<td>10</td>
<td>100</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Kaczka</td>
<td>6</td>
<td>6</td>
<td>100</td>
<td>23</td>
<td>20</td>
<td>87.0</td>
</tr>
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ND, not detailed.
influenced by food intake; also, CT is rapidly degraded at room temperature, and it decays by 23% after 12 h, by 35% after 24 h, and by 65% after 7 days (Perdrisot et al. 1990, Zink et al. 1992). Therefore, blood samples should be drawn in the morning after overnight fasting, and it is necessary to centrifuge the samples immediately after blood coagulation and then to freeze and transport them in ice to the laboratory. The analytical methods of CT assay have evolved over time, and most laboratories that perform routine CT measurement have moved away from manual IRMA methods (Perdrisot et al. 1990, Zink et al. 1992) to automated chemiluminescent immunoassay (ICLA) platforms that have similar analytical performance (Grauer et al. 1998). However, the variable levels of the different products of the CT gene result in several circulating immunoreactive isoforms and fragments (Perdrisot et al. 1990, Zink et al. 1992), and, consequently, poor interassay and inter-laboratory agreement was demonstrated between the reviewed studies (Giovanella et al. 2012).

In the post-analytical phase, establishing normal limits of CT in healthy subjects is still a challenge because of the significant variability in diagnostic accuracy and reference intervals of the different commercially available assays (Giovanella et al. 2012). Healthy subjects almost always have serum CT concentrations of less than 10 pg/ml, but only a CT value that exceeds 100 pg/ml is considered to be a reliable marker for the presence of MTC. A stimulation test with an infusion of pentagastrin or calcium gluconate is required in patients with mildly elevated basal CT levels; stimulated CT values of >100 pg/ml should prompt surgery (Trimboli et al. 2014). These diagnostic guidelines, which have been widely adopted in clinical practice, were established by using the manual IRMA provided by CisBio (Perdrisot et al. 1990, Zink et al. 1992). In addition, the upper reference limits for females and males should be different, because the number of C cells is approximately two times higher in men than in women (Trimboli & Giovanella 2015). As a practical consequence and in order

![Figure 1](http://erc.endocrinology-journals.org) The process that leads to calcitonin production.
to avoid false diagnoses, specific reference values are required before each new CT assay can be used in clinical practice. Finally, increased CT may be associated with bPreoperative ProCT value.
aPostoperative ProCT value.
ProCT values are reported in ng/ml.

First author ProCT levels
Boiko> 0.5
Algeciras-Schimnich> 0.15
Kaczka> 0.1
Walter> 0.5
Kratzsch> 0.25
Kaczka> 0.1
Giovanella> 0.1
Machens> 0.1

ProCT values are reported in ng/ml.
*Postoperative ProCT value.
*Preoperative ProCT value.

In order to overcome such limits, potential alternative MTC serum markers, such as CEA, chromogranin A, and ProCT, have been investigated. Of these molecules, ProCT has appeared to be the most promising. In the more recent literature, ProCT has been reported to be an accurate new serum marker for the diagnosis and follow-up of MTC. Some studies that compared the abilities of ProCT and CT in order to distinguish MTC from other conditions revealed results (Perdrisot et al. 1990, Zink et al. 1992, Giovanella et al. 2010, Giovanella 2012). Unlike CT, ProCT is a very stable protein that has a concentration-independent in vivo half-life of 20–24 h, and it does not need to be kept cold on ice or frozen. It is also easier to manage at the pre-analytical level. Because they use the same antibodies, all commercial ProCT assays yield similar results, which enables the standardization of biomarker-based diagnosis, therapy, and monitoring of patients with MTC. This feature of ProCT differs clearly from CT and has high relevance in clinical practice. Some limitations, however, affect ProCT determination; in fact, a marked increase in this molecule has been reported in systemic infections (mainly bacteriemia) and inflammatory diseases, and these conditions represent the main indications for ProCT measurement (Plebani & Fabbri 2015).

In the present report, we systematically reviewed the published articles on the use of ProCT in MTC patients. Different cutoffs of positivity for ProCT were adopted in the reviewed studies (Table 3). The majority of these reports described ProCT measurement during the follow-up of patients after surgical treatment. In this regard, cured patients very rarely had detectable ProCT, with a specificity of 100% in the largest study and specificity that ranged from 57 to 87% in the other three series; this was the most interesting result for clinical practice. Also, in patients with active/recurrent MTC, a 100% sensitivity was recorded in four papers, and the two others listed sensitivities of 84 and 91%. Patients with stable disease had ProCT values that were significantly lower than those in cases of MTC in progression. All of the results of ProCT in recurrent and cured MTC are shown in Table 2. Two of the studies reported the evaluation of ProCT as a marker for the initial preoperative detection of MTC. A significant finding in terms of prognosis was described as part of one study (Machens et al. 2014): 107 out of 112 MTC had ProCT values of more than 0.1 ng/ml, with a sensitivity of 95.5%, and the ProCT values correlated with the extent of the disease. Furthermore, the biochemical cure rates were inversely correlated with the ProCT levels. In the other study (Giovanella et al. 2013), pentagastrin-stimulated CT and ProCT were tested in patients with moderately elevated CT, and undetectable levels of both basal and stimulated ProCT had 100% PPV and NPV in identifying MTC cases. In light of these data, serum ProCT could have the potential to replace serum CT as a new standard of care in the management of MTC. A relevant advantage of using ProCT assays is that all commercial assays yield quite similar results. One limitation might be that measurements of ProCT are not very widely available; however, because of its increasing use as a pivotal marker in internal

<table>
<thead>
<tr>
<th>First author</th>
<th>Athyreotic patients*</th>
<th>Healthy subjects</th>
</tr>
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<tbody>
<tr>
<td>Algeciras-Schimnich Kaczka</td>
<td>&lt;0.1 (mean) 0.06 (mean) 0.00–0.28 (min–max)</td>
<td>&lt;0.1 (mean) 0.06–0.28 (min–max)</td>
</tr>
<tr>
<td>Walter</td>
<td>0.07–0.60 (min–max)</td>
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</table>

ProCT values are reported in ng/ml.
*Patients with a non-MTC histological diagnosis.
The two studies by Kaczka et al. probably reported the same control group of thyroidectomized patients.
medicine and antibiotic therapy, the availability of ProCT is expected to grow during the coming years (Plebani & Fabbri 2015). The most critical challenge is that further studies are needed to identify the optimal cutoff level of ProCT, so that active (i.e., recurrent/persistent) MTC cases can be distinguished from cured ones. To date, data recorded in healthy and athyreotic subjects could be used in an initial approach to define an accurate threshold. As shown in Table 4, ProCT levels of up to 0.28 ng/ml have been recorded in athyreotic and healthy subjects.

Interestingly, ProCT appears to be useful in those rare MTC cases where CT is undetectable both preoperatively and postoperatively and where this marker of disease activity is therefore lacking. Brutsaert et al. (2015) tested ProCT levels of one patient before and after surgery: its value converted from detectable (0.21 ng/ml) to undetectable (<0.1 ng/ml) respectively. Bugalho et al. (2014) described a man with recurrent MTC with liver metastasis of 34 mm, a low CT value (<50 ng/ml), and a high ProCT value (10.1 ng/ml). These reports should receive attention specifically because they could help with the management, before and after surgery, of those MTC cases with negative/undetectable CT (Trimboli & Giovanella 2015). Previously, Novotny et al. (2009) showed that ProCT can be used for monitoring sepsis in a patent affected by MTC. However, we still do not know whether common bacterial infections pose particular difficulties in the interpretation of ProCT levels. This issue has to be considered when CT and ProCT are used as MTC markers in patients with suspected infectious disease.

So far, CT has been considered an ‘ideal’ marker for diagnosing MTC in patients who are suspected to have MTC or are routinely screened for MTC as well as for the follow-up of those who have already been diagnosed with MTC. Results of previous studies have indicated the superiority of CT as compared with other markers, such as CGRP. However, CT assays have some intrinsic limitations that affect their reliability in the diagnosis and management of MTC patients, especially at the pre-analytical level. The ProCT data that are so far available from a sizable number of studies show very promising results, and it is tempting to speculate that the ProCT assay may in the future replace CT in the management of MTC patients.

All of the data retrieved in the present review are indicative of a potentially important role of ProCT measurement in the management of MTC patients. It would therefore be worthwhile to establish cutoff levels using larger patient series in a prospective manner. Until then, values in the normal population could be used as a reference range.


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