In situ androgen and estrogen biosynthesis in endometrial cancer: focus on androgen actions and intratumoral production

Kiyoshi Ito1,2, Yasuhiro Miki1, Takashi Suzuki3, Keely May McNamara4 and Hironobu Sasano4

1Department of Disaster Obstetrics and Gynecology, International Research Institute of Disaster Science (IRiDeS), Tohoku University, Sendai, Japan
2Department of Disaster Obstetrics and Gynecology, Tohoku University Graduate School of Medicine, Sendai, Japan
3Department of Pathology and Histotechnology, Tohoku University Graduate School of Medicine, Sendai, Japan
4Department of Pathology, Tohoku University Graduate School of Medicine, Sendai, Japan

Abstract

In situ estrogen biosynthesis is considered to play pivotal roles in the development and progression of human endometrial carcinoma. However, the biological roles of androgen have remained virtually unknown. Various epidemiological studies have revealed that elevated serum androgen levels are generally associated with an increased risk of developing endometrial carcinoma; however, studies directly examining androgens in carcinoma tissues are relatively rare and reviews summarizing this information are scarce. Therefore, we summarized recent studies on androgens in endometrial carcinoma, especially focusing androgen actions and in situ androgen biosynthesis. Among the enzymes required for local biosynthesis of androgen, 17β-hydroxysteroid dehydrogenase type 5 (conversion from androstenedione to testosterone) and 5α-reductase (reduction of testosterone to dihydrotestosterone (DHT)) are the principal enzymes involved in the formation of biologically most potent androgen, DHT. Both enzymes and androgen receptor were expressed in endometrial carcinoma tissues, and in situ production of DHT has been reported to exist in endometrial carcinoma tissues. However, testosterone is not only a precursor of DHT production, but also a precursor of estradiol synthesis, as a substrate of the aromatase enzyme. Therefore, aromatase could be another key enzyme serving as a negative regulator for in situ production of DHT by reducing amounts of the precursor. In an in vitro study, DHT was reported to exert antiproliferative effects on endometrial carcinoma cells. Intracrine mechanisms of androgens, the downstream signals of AR, which are directly related to anticancer progression, and the clinical significance of DHT-AR pathway in the patients with endometrial carcinoma have, however, not been fully elucidated.

Introduction

Endometrial carcinoma is one of the most common female pelvic malignancies in the developed countries, and its incidence has increased. In the United States, 60,050 new cases and 10,470 deaths due to endometrial carcinoma were reported in 2016 (Siegel et al. 2016). It is well known that sex-steroid hormones play pivotal roles
in the development of hormone-dependent carcinoma, including prostate, breast and endometrium, and in the case of the latter, estrogen actions are well known to play important role in the development and progression of endometrial carcinoma. Results of many previously reported clinical studies demonstrated that excessive and/or prolonged exposure to unopposed estrogens increased the risk of endometrial carcinoma especially that of the endometrioid type, also known as type I endometrial carcinoma (Bokhman 1983). However, in contrast to studies regarding estrogens, the possible impact of androgen actions on endometrial carcinoma has largely remained unknown. Therefore, we summarize the recent studies regarding the effects and/or actions of androgens in endometrial carcinoma in this review.

Serum androgen and increased risk of developing endometrial cancer

The great majority of endometrial cancers arise during the postmenopausal period. Circulating estrogen levels decline steeply after menopause but testosterone levels remained unchanged during the menopausal years (Burger et al. 2000). Over the age of 65 years, total testosterone levels decline with age until 80, alongside free testosterone levels did not vary by age (Cappola et al. 2007). All androgens in postmenopausal women are suggested to be made locally in peripheral target tissues according to the mechanism of local intracrine formation of steroids. The contribution of ovarian production of circulating androgenic precursors and thus potentially supporting this local intracrine metabolism of steroids is underlined by studies showing serum dehydroepiandrosterone (DHEA); the inactive precursor hormone of androgens is higher in intact compared with oophorectomized women (Labrie et al. 2011, Labrie 2015), and several studies reported that the serum levels of androgens between ovarian and peripheral veins were significantly different between early and late menopausal women, suggesting that the production of androgens by the ovaries could persist even 10 years after the onset of menopause, but may subsequently decline (Fogle et al. 2007, Maruoka et al. 2014). Therefore, the degree and duration to which postmenopausal ovaries remain a viable androgen-producing organ is still controversial and not fully understood.

Several investigators reported that elevated serum levels of androgens, including DHEA, DHEA sulfate (DHEAS), androstenedione (A4), testosterone and 5α-dihydrotestosterone (DHT), were significantly associated with an increased risk of developing endometrial carcinoma (Potischman et al. 1996, Kaaks et al. 2002, Lukanova et al. 2004, Allen et al. 2008, Audet-Walsh et al. 2011). Potischman and coworkers also reported that high circulating levels of A4 were associated with 2.8-fold increased risks of developing endometrial cancer among postmenopausal women, following the adjustment for other factors (Potischman et al. 1996). These authors subsequently concluded that circulating levels of A4 yielded most significant and consistent correlation with the risks of developing endometrial carcinoma compared with those of estrone (E1), estradiol (E2) and E1-sulfate (E1S) (Potischman et al. 1996). Kaaks and coworkers also reviewed numerous epidemiological studies and concluded that endometrial carcinoma risks were increased among pre- and postmenopausal women associated with elevated plasma A4 and testosterone (Kaaks et al. 2002). Thereafter, two prospective studies have reported discrepant results for the association of cancer risk with serum androgen levels. A multicenter prospective study also demonstrated that elevated circulating levels of A4 and DHEAS were positively associated with endometrial cancer risk, although free testosterone did not correlate with an increased risk among postmenopausal women (Lukanova et al. 2004). In contrast, elevated circulating levels of total and free testosterone were positively associated with endometrial cancer risk, although A4 and DHEAS were not necessarily correlated with increased risks among postmenopausal women in the European Prospective Investigation into Cancer and Nutrition Study (EPIC) (Allen et al. 2008). Recently, using highly specific and sensitive validated methods based on liquid chromatography/electrospray tandem mass spectrometry (LC-MS/MS), Audet-Walsh and coworkers compared the levels of 18 circulating steroid hormones in 126 postmenopausal cases of endometrial carcinoma with 110 cases of healthy postmenopausal women and to investigate how these hormonal levels could be possibly related to clinical characteristics (Audet-Walsh et al. 2011). Most hormones, including DHEA, DHEAS, A4, testosterone and DHT, were significantly elevated in endometrial carcinoma cases compared with healthy controls. These particular epidemiological findings clearly indicate that elevated serum androgen levels increase the risks of developing endometrial carcinoma at least in postmenopausal women.

The impact of polycystic ovary syndrome (PCOS), a well-known common endocrine disorder affecting 5–8% of women with reproductive age and associated with menstrual irregularity, elevated androgen and polycystic ovaries (Azziz et al. 2004), has also been examined in relation to endometrial cancer risk,
and given its association with increased androgenicity may give important clues to the link between androgens 
and this malignancy. A recent systemic review and 
meta-analysis revealed that women with PCOS have 
significant increased risks of endometrial carcinoma 
(odds ratio 2.79) compared with those without PCOS. In 
addition, the risks of women with PCOS increased further 
for endometrial carcinoma (odds ratio 4.05) less than 
54 years of age (Barry et al. 2014). PCOS and endometrial 
carcinoma shared many of the similar risk factors (obesity, 
diabetes, anovulation, inflammation etc.), but elevated 
androgens themselves may be considered the pivotal factor 
for endometrial carcinoma risk. However, data on serum 
levels of androgen and risks of developing endometrial 
carcinoma among premenopausal women are yet to 
be completely established. To the best of our knowledge, 
only two prospective studies have been reported to date 
and coworkers recently reported that no significant association 
was detected between circulating androgens and risks of 
developing endometrial carcinoma before menopause, 
which was consistent with the results of other prospective 
study (Clendenen et al. 2016). However, it is entirely true 
that androgen concentrations obtained in these studies 
were measured by classical radioimmunoassay (Allen et al. 
2008, Clendenen et al. 2016) with the inherent limitations 
involved in this methodology. Thus while these studies 
provide important preliminary evidence, future studies 
employing gold standard techniques of steroid assays 
would be ideal in resolving the question of circulating 
androgens and risks of developing endometrial carcinoma 
in premenopausal women.

Despite the information above suggesting a 
relationship between androgens and endometrial cancer, 
risk studies directly examining the effects of androgens 
on endometrial cancer cells have had mixed results in 
contrast to the consistent stimulation of proliferation 
observed with estrogens in the same models (Tuckerman et al. 2000, Braunstein 2007). In addition, in a more 
physiological setting, short-term exogenous testosterone 
treatment of postmenopausal women did not stimulate 
endometrial proliferation and counteract the proliferative 
effects induced by E$_2$ (Zang et al. 2007, Shufelt & 
Braunstein 2009). Therefore, the biological link between 
the increased endometrial carcinoma risks associated with 
elevated levels of androgen in circulation and androgen 
actions on endometrial cells is not well characterized or 
understood. Several investigators reported that elevated 
plasma levels of androgens resulted in increased tissue 
estrogen levels through their aromatization in peripheral 
or local tissues, including adipose, breast and endometrial 
carcinoma tissues (Sasano & Harada 1998, Bélanger et al. 
2002, Somboonporn et al. 2004). Therefore, androgens 
may be locally converted into estrogens by adipose tissue 
and/or hiding small neoplastic cells and neoplastic cells 
could possibly grow rapidly in the presence of locally 
produced estrogens especially in postmenopausal women. 
Estrogen-dependent neoplasms such as breast and 
endometrioid endometrial carcinoma, in which in situ 
conversions from serum androgens to biologically active 
estrogens occur and are biologically important, are therefore 
considered as ‘intracrine’-dependent malignancies.

**In situ androgen metabolism and synthesis**

**Intracrinology**

Recently, a great deal of focus has been given to the 
importance of intratumoral metabolism and synthesis 
of biologically active steroids in various systems. This 
is attributable to the documented importance of the 
interactions of various enzymes in the pathogenesis 
and progression of various human hormone-dependent 
neoplasms, including breast and endometrial carcinoma. 
This complex system, shown in a simplified version in 
figure one, means that enormous local control over the 
types and amounts of steroids produces and specific to 
one tissue is possible. Results of numerous studies have 
demonstrated increased tissue estrogen content in human 
breasts, compared with serum and/or normal non-
neoplastic tissues of the same patients (van Landeghem 
et al. 1985, Pasqualini & Chetrite 1999, Chetrite et al. 
2000). In these studies, the tissue concentrations of E$_1$ 
and E$_2$ were generally several times higher than those 
detected in the plasma or in the area of the normal breast 
tissues of the same postmenopausal patients, despite 
markedly low levels of circulating estrogens. In contrast 
to breast carcinoma cases, there is limited data regarding 
tissue estrogen concentrations in endometrial carcinoma 
tissues (Bonney et al. 1986, Vermeulen-Meiners et al. 
and coworkers examined 78 endometrial carcinomas 
and detected higher concentrations of E$_1$ in carcinoma 
tissue specimens compared with macroscopically normal 
endometrium (Berstein et al. 2003). Our findings (Ito 
2005, Ito et al 2006) were generally consistent with those 
of other studies demonstrating higher E$_1$ levels associated 
with carcinoma. These results suggest that intratumoral 
estrogen metabolism and synthesis are pivotal in the 
progression of endometrial carcinoma, although the
available data of sex-steroid hormone concentrations were all obtained by classical radioimmunoassay (RIA) and enzyme-linked immunosorbent assay (ELISA) and not by the modern gold standard methods of mass spectrometry. This distinction may be important as the classical methods of RIA and ELISA have been the standard methodology to analyze steroid hormones; however, these methods harbor several problems, including sensitivity and specificity/cross-reactions (Rauh 2012). On the other hand, LC-MS/MS is highly specific and sensitive to validated methods, which could measure highly specific hormones from very small amounts of the sample (Rauh 2012).

The status of intratumoral androgen concentration has been less explored than that of estrogen in breast and endometrial carcinoma. Among the androgens, DHT binds with the highest affinity to androgen receptor (AR), and together with testosterone promotes AR transcriptional activity, while A4 and DHEA have extremely low binding affinity for AR (Avancés et al. 2001). Potent androgen DHT concentrations were significantly higher in breast carcinoma tissues than in plasma (Recchione et al. 1995), and the tissue concentration of DHT was three-fold higher in ductal carcinoma in situ of the breast compared with the non-neoplastic breast (Shibuya et al. 2008). The analysis of serum DHT concentration in the patients of endometrial carcinoma was reported (Audet-Walsh et al. 2011), but not in the cases of DHT concentration in endometrial tumor tissues. We recently measured the tissue and serum concentrations of testosterone and DHT in 31 endometrioid endometrial adenocarcinoma tissue and 5 normal endometrium samples using LC-MS/MS methods (Tanaka et al. 2015) with the advantages mentioned above. This study revealed that a markedly elevated DHT tissues/serum concentration ratio was detected in endometrioid endometrial adenocarcinoma tissues compared with that in normal endometrium tissues (8.0-fold). This finding also indicated that local production of DHT in endometrioid endometrial adenocarcinoma could be similar to that of breast carcinoma. Among the enzyme system required for local biosynthesis of androgen, 17β-hydroxysteroid dehydrogenase type 5 (17β-HSD5: conversion from A4 to testosterone) and 5α-reductase (5α-Red: reduction of testosterone to DHT) are the principal enzymes involved in the formation of biologically active androgen, DHT. The examination of these responsible enzymes in endometrial carcinoma tissues is very important to obtain the better understanding of the significance of androgen in endometrial carcinomas.

17β-HSD5

17β-HSDs are key enzymes involved in the formation of biological active sex steroids, including testosterone, E1 and E2. The reduction of A4 is known to result in the production of testosterone mainly by 17β-HSD5 (Dufort et al. 1999). 17β-HSD5, a reductive 17β-HSD, is a member of aldo-keto reductase superfamily and formally termed AKR1C3, whereas the others are members of the short-chain alcohol dehydrogenase (Deyashiki et al. 1995). 17β-HSD5 is expressed in various peripheral tissue, liver, prostate, ovary and has been also reported in prostate and breast carcinoma tissues (Luu-The et al. 2001, Vihko et al. 2004, Suzuki et al. 2010).

Increased 17β-HSD5 mRNA and protein expression were reported in endometrial carcinoma tissues compared with normal endometrium (Rizner et al. 2006, Smuc & Rizner 2009). However, one group reported weaker protein expression of 17β-HSD5 in hyperplastic and cancerous endometrium compared with normal proliferative endometrium (Zakharov et al. 2010) and no significant difference of mRNA expression between cancerous and normal endometrial tissues was detected in other groups (Cornel et al. 2012, Sinreih et al. 2013). In our previous study, 17β-HSD5 immunoreactivity was detected in 18/36 (50%) and 71/103 (69%) cases of endometrial hyperplasia and endometrioid endometrial carcinoma, respectively. However, 17β-HSD5 immunoreactivity was detected only in 5/26 (19%) and 5/20 (25%) cases of proliferative and secretory phase endometria, respectively (Ito 2005, Ito et al. 2006). 17β-HSD5 immunoreactivity in endometrioid endometrial carcinoma was significantly higher than that of proliferative and secretory phase and endometrial hyperplasia. Due to the inconsistent and limited data on the available studies of 17β-HSD5 expression, further investigation is warranted.

5α-reductase

5α-reductase (5α-Red) is an enzyme responsible for the conversion of testosterone to a most potent androgen, DHT. This enzyme is considered to be an important regulator of local actions of androgens. Two 5α-Red isoenzymes, Types 1 and 2 (e.g. 5α-Red1 and 5α-Red2), have been cloned and characterized in humans (Russell & Wilson 1994). In 18 normal cycling human endometria, 5α-Red1 and 5α-Red2 were both detected in the cytoplasm of epithelial cells, with weak or focal perinuclear immunolocalization patterns through all phases of the menstrual cycle (Ito et al. 2002). 5α-Reds mRNA and protein were also
detected in biopsies of pelvic endometriosis, as well as in the eutopic endometrium (Carneiro et al. 2008). In our previous study, immunoreactivity of 5α-Red1 and 5α-Red2 was detected in 37/44 (84%) and 34/44 (77%) cases of endometrioid endometrial carcinoma, respectively (Ito et al. 2002). 5α-Reds protein expression was demonstrated in the cytoplasm of carcinoma cells. Endometrial carcinoma tissue homogenates were associated with mRNA expression in the great majority of the cases examined. However, mRNA expression of 5α-Red2 was significantly down-regulated and that of 5α-Red1 was unchanged in endometrial carcinoma versus adjacent control endometrium, including proliferative, secretory and menopausal status (Sinreih et al. 2013). Recently, we measured the tissue concentration of androgen and performed immunohistochemical analyses of AR and 5α-Reds in 36 endometrioid endometrial carcinoma cases (Tanaka et al. 2015). Results of our study did reveal that 5α-Red1 immunoreactivity was detected in approximately 64% of endometrioid endometrial carcinoma and was positively correlated to intratumoral DHT concentration with statistical significance. No significant correlation was observed between 5α-Red2 immunoreactivity and the intratumoral concentration of DHT, although the immunoreactivity of 5α-Red2 was similar to that of 5α-Red1. In breast carcinoma, which has estrogen-dependent growth similar to endometrioid endometrial carcinoma, intratumoral concentration of DHT was significantly correlated with that of testosterone (Recchione et al. 1995), and aromatase expression was inversely correlated with intratumoral DHT concentration (Suzuki et al. 2007) suggesting the functional possibilities of such a hypothesis and its potential relevance in endometrial carcinoma.

We previously demonstrated marked aromatase immunoreactivity and mRNA, mainly in the stromal cells of endometrioid endometrial carcinoma, but not in normal or hyperplastic endometrium (Watanabe et al. 1995). This result suggested an induction of aromatase

Therefore, 5α-Red1 is reasonably postulated to act predominantly for the local production of DHT in endometrial carcinoma, as reported in breast carcinoma.

**Negative regulator for in situ DHT production in endometrial cancer**

A4 and testosterone are precursors of not only DHT production, but also those of estradiol synthesis, as the substrate of aromatase (Fig. 1). Aromatase is an enzyme which catalyzes the conversion for androgens, mainly A4 and testosterone, to E1 and E2, respectively (Bulun et al. 2005). DHT itself is nonaromatizable. Therefore, aromatase could be a negative regulator for in situ production of DHT in endometrial carcinoma tissues by possibly reducing concentration or availability of the precursor testosterone and/or A4. Of particular interest, in breast carcinoma, which has estrogen-dependent growth similar to endometrioid endometrial carcinoma, intratumoral concentration of DHT was significantly correlated with that of testosterone (Recchione et al. 1995), and aromatase expression was inversely correlated with intratumoral DHT concentration (Suzuki et al. 2007) suggesting the functional possibilities of such a hypothesis and its potential relevance in endometrial carcinoma.

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![Figure 1](http://erc.endocrinology-journals.org/doi/figure/10.1530/ERC-15-0470)
expression by tumor-stromal interactions. In addition, a significant correlation was detected between aromatase expression in stromal cells and poor prognosis in the patients with endometrial carcinoma (Segawa et al. 2005), although several studies reported that aromatase expression was demonstrated in carcinoma and stromal cells together (Segawa et al. 2005, Jongen et al. 2009, Che et al. 2014). It is very important to consider in vitro stromal–carcinoma cell interactions when studying aromatase expression in vitro. We previously examined whether estrogen biosynthesis in the tumor microenvironment promoted endometrial carcinoma or not. In order to study the contribution of stromal cells to estrogen signaling pathways in endometrial carcinoma, we examined reporter cells stably transjected with the estrogen response element (ERE) fused to the destabilized green fluorescent protein (GFP) gene (Matsumoto et al. 2008). In this system, the ERE of carcinoma cells was activated to a variable extent by intratumoral stromal cells isolated from endometrial carcinoma. When testosterone was added as a substrate for aromatase, the GFP expression levels increased. Thereafter, these effects were variably inhibited by aromatase inhibition. These results indicated that GFP expression is driven by estrogen synthesized by aromatase in the endometrial cancer stromal cells. In addition, in order to further confirm the local biosynthesis of estrogens and tumor–stromal interactions on aromatase activity, endometrial carcinoma cell lines were cocultured with stromal cells isolated from endometrial carcinomas, and aromatization activity was measured using LC-MS/MS (Takahashi-Shiga et al. 2009). We also examined the effects of aromatase inhibitors on cell proliferation. Aromatase activity was significantly higher in the coculture with carcinoma cell lines and stromal cells than in each monoculture, respectively. Cell proliferation was significantly inhibited in carcinoma cells treated with aromatase inhibitors compared with control cultures. Recently, Che and coworkers also reported that aromatase expression in stromal, but not in carcinoma epithelium cells, was associated with interleukin 6 expression in epithelial cells of carcinomas by immunohistochemistry, which were subsequently confirmed also using laser capture microdissection/real-time RT-PCR in endometrial carcinoma tissues (Che et al. 2014). Their results did demonstrate the presence of the activation of a positive feedback loop in which interleukin 6 stimulated by E2 in endometrial carcinoma epithelial cells induced aromatase expression in stromal cells, thereby promoting intratumoral E2 synthesis. These findings above did confirm the importance of carcinoma–stromal cells interaction in the process of induction of aromatase in endometrial carcinoma tissues, and demonstrated that aromatase is a key enzyme in the local biosynthesis of estrogen in endometrial carcinoma as well as breast carcinoma. However, further investigation should be required to clarify the association between the expression of aromatase and intratumoral DHT concentration in endometrial carcinoma.

DHT is catalyzed to 3α and 3β androstanediol, which are relatively inactive, by 3α-keto-reductase (AKR1C) and 3β-keto-reductase (AKR1C2), respectively. The enzyme of AKR1C2 has the predominant catalytic efficiency. Rižner and Penning (2014) reviewed the possible roles of AKR1 enzymes in human steroid metabolism and alluded on the possibility of AKR1C gene expression in endometrial carcinoma. However, AKR1C1 and AKR1C2 mRNA expression in cancerous endometrium did not differ significantly to that in adjacent control normal tissue (Smuc & Rižner 2009, Sinreih et al. 2013). Therefore, it is unlikely that AKR1C1 and AKR1C2 serve as negative regulators of in situ production of DHT in endometrial carcinoma.

**Androgen receptor and androgen action in endometrial cancer**

**Structure of androgen receptor**

Androgens mediate their effects through the androgen receptor (AR). AR belongs to the nuclear receptor subfamily 3 group C gene 4 (NR3C4) and also to the nuclear receptor superfamily for steroid hormones. DHT binds with the highest affinity to AR, and together with testosterone, promotes AR transcriptional activities (Avancès et al. 2001). A polymorphic CAG repeat in exon 1 of the AR gene is well known to encode a polyglutamine tract and to increase the length of CAG repeat, which is also well known to be associated with decreased transactivation activities (Chamberlain et al. 1994). Of particular interest, a statistically significant inverse association has been reported between increased CAG repeat length and endometrial carcinoma risk in case-control studies (McGrath et al. 2006, Ashton et al. 2010), although controversies exist (Yang et al. 2009). On the other hand, Yang and coworkers evaluated 36 sex hormone-related genes using a tagging approach in a population-based case-control study of 417 endometrial carcinoma cases and reported that the possible evidence of the correlation with endometrial cancer risk could be the common genetic variation in...
AR (Yang et al. 2010). All of these results above did suggest that variant structures of AR played pivotal roles in the risks of developing endometrial carcinoma.

Androgen receptor expression

In the nonmalignant endometrium, AR was reported to be expressed in the endometrium throughout the menstrual cycle (Mertens et al. 2001, Apparao et al. 2002, Ito et al. 2002, Marshall et al. 2011, Gibson et al. 2014). The status of AR immunoreactivity in stromal cells is higher than that of epithelial AR throughout the menstrual cycle (Apparao et al. 2002). In our previous study, AR immunoreactivity was detected in the nuclei of both epithelial and stromal cells (Ito et al. 2002). AR immunoreactivity was also detected in the nuclei of 60–70% of stromal cells in the proliferative phase, while negative at functional is of endometrium in the secretory phase. AR immunoreactivity was detected in the nuclei of around 5–15% of epithelial cells in the secretory phase, while negative or very weak in the proliferative phase. Endogenous and exogenous androgens should have the ability to alter endometrial function through AR expression. For example, in women with PCOS, who generally exhibit chronic hyperandrogenism, exhibited elevated endometrial AR immunoreexpression has been documented compared with normal fertile control (Apparao et al. 2002) suggesting the effects on androgens in changing the intracrine environment of these tissues. In a similar vein, albeit circumstantial, correlation between endometrial AR expression and serum androgens levels has also been demonstrated in the context of obesity and weight loss. Serum androgens are reported to be positively correlated with BMI (Allen et al. 2008) and abdominal fat accumulation (Cao et al. 2013), but in asymptomatic morbidly obese women in response to interventions that reduce abdominal fat and/or BMI such as bariatric surgery and weight loss, AR immunoreexpression dropped significantly in response to these interventions (Argenta et al. 2014) perhaps hinting at the correlation between serum androgens and endometrial AR expression.

The expression of AR in endometrial carcinoma has been detected in several studies (Horie et al. 1992, Sasaki et al. 2000, Ito et al. 2002, Qiu et al. 2014, Tanaka et al. 2015). The number of cases was small (n=4), but Horie and coworkers reported that all specimens of endometrial carcinoma tissues examined expressed AR (Horie et al. 1992). On the other hand, Sasaki and coworkers reported that AR was not detected in 79% of endometrial carcinomas, although their cases included several unknown histological types (Sasaki et al. 2000). Qiu and coworkers recently reported AR-highly expressing cells in 50 of 76 cases (66%) of endometrial carcinoma (Qiu et al. 2014). In our previous study, AR immunoreactivity was detected in the nuclei of carcinoma cells and the number of positive cases was 39/44 (89%) (Ito et al. 2002). In the great majority of cases, carcinoma tissue homogenates did also have mRNA expression. Thereafter, AR expression was recently reported in 92 of 122 cases (82%) of endometrioid endometrial carcinoma (Tanaka et al. 2015). These studies all demonstrated that AR was expressed in the great majority of endometrial carcinoma, especially in endometrioid type.

Androgen action

The possible impact of androgen actions and the pivotal roles of androgen signaling through its receptor in endometrial carcinoma have not yet been clarified. DHT is nonaromatizable androgen and not converted to estrogen. DHT acts most markedly as an androgen through the AR pathway. Hackenberg and coworkers and Hackenberg & Schulz first demonstrated the inhibition of cell proliferation in human endometrial carcinoma cell line, MFE-296 cells, by DHT (Hackenberg et al. 1994, Hackenberg & Schulz 1996). Lovely and coworkers subsequently reported that DHT can act as antiestrogens through inhibiting the alkaline-phosphatase enzymatic activity in a dose-dependent manner, using hormonally responsive cell lines (a well-differentiated endometrial adenocarcinoma cell lines, Ishikawa) (Lovely et al. 2000). AR protein expression in Ishikawa cell lines was subsequently reported to be up-regulated by DHT or E2 and to be inhibited by the antiandrogen, hydroxyflutamide or bicalutamide, or medroxyprogesterone acetate (MPA) based on Western blot analysis (Apparao et al. 2002). These studies consistently demonstrated that androgens had the important ability to inhibit cell proliferation through DHT-AR pathway in endometrial carcinomas in vitro. In contrast, in vivo animal model study demonstrated that DHT excess neither significantly suppressed nor increased the growth rate of well-differentiated human endometrial carcinoma implants in the nude mice, although AR expression in tumor tissues were not examined (Legro et al. 2001).

In breast carcinoma, DHT is considered to be locally produced mainly via 5α-Red1. In addition, immunohistochemical analysis showed that patients with positive for both AR/5α-Red1 demonstrated significant correlations with decreased risk of recurrence.
and improved prognosis for overall survival, and AR/5α-Red1 status was an independent prognostic factor (Suzuki et al. 2007). AR/5α-Red1 double-negative status was significantly correlated with worse prognosis even in triple-negative invasive breast carcinoma, which is defined as the absence of ER, PR and human epidermal growth factor receptor 2 (HER2) expression in tumor cells (McNamara et al. 2013). In addition, the absence of intratumoral androgenic enzymes, both 5α-Red1 and 17β-HSD5, was significantly associated with adverse clinical outcome in patients with invasive lobular carcinoma of the breast (Yoda et al. 2014). All of these findings above did indicate that DHT could inhibit the cell proliferation through DHT-AR pathway and influence the eventual clinical prognosis of the patients with breast cancer.

Using the logic and evidence of studies in the breast as guidelines, it then becomes clear that it is very important to analyze the correlation between the status of AR and 5α-Reds and clinicopathological findings to further clarify the clinical significance of DHT-AR pathway in endometrial carcinoma. AR expression was significantly inactivated by hypermethylation of the AR gene CpG islands in human endometrial carcinoma compared with normal endometria, using pairs of carcinomatous and normal samples from 28 patients (Sasaki et al. 2000). All carcinoma samples from advanced stage did demonstrate only methylated AR alleles and immunoexpression of AR negative, although about half of the cases from early stage harbored these alleles. In addition, Rodríguez and coworkers had genotyped both CAG and GGN repeats from 204 cases of endometrial carcinoma tissues and revealed that the presence of short CAG or GGN repeats of the AR genes in a tumor specimens was correlated with a favorable conditions of prognostic values in endometrial carcinoma (Rodríguez et al. 2006). Shortening each repeat has been considered to increase either the amount of AR protein (GGN) or its activity (CAG) (Chamberlain et al. 1994, Ding et al. 2005). In our recent study using 86 endometrioid endometrial adenocarcinoma samples with immunohistochemical analysis, the AR-positive status was significantly correlated with progression-free survival (PFS), but not with endometrial carcinoma-specific survival (ECSS) in endometrial carcinoma patients (Tanaka et al. 2015). 5α-Red1-positive status was significantly associated with both PFS and ECSS in those patients. In addition, AR/5α-Red1 double-negative status was significantly correlated with worse PFS and ECSS, when these patients were divided into four groups, according to the status of AR and 5α-Red1, such as AR/5α-Red1 double positive, positive/negative, negative/positive and double negative status, respectively (Tanaka et al. 2015). In addition, Kamal and coworkers recently reported that AR was expressed in postmenopausal endometrial epithelium and their subsequent loss in endometrial carcinoma was significantly associated with disease-free survival (Kamal et al. 2016). Those results consistently demonstrated that a lack of androgen action in intratumoral levels is correlated with a poor prognosis in the patients, and androgen signaling exerts anticancer effects through DHT-AR pathway in endometrial carcinoma. Results of our study also indicated that it was important to evaluate both AR status and 5α-Red1, the latter serving as a surrogate marker of intratumoral DHT production, for understanding the effect of androgenic signaling on carcinoma progression (Miki et al. 2015, Tanaka et al. 2015).

### Androgen action from the viewpoint of therapeutic target

In order to understand the anticancer effects through DHT-AR pathway, it is necessary to analyze the downstream signals of AR. However, such DHT-induced genes have remained largely unknown in endometrial carcinoma. Qiu and coworkers recently demonstrated that the mRNA expression of AR target gene (XBP1, MYC, ZBTB16, and UHRF1) increased after treatment with DHT in the human endometrial carcinoma cell line MFE-296 cells, although the roles of those target genes has still remained virtually unknown (Qiu et al. 2014). In addition, the possible roles of androgen-induced genes in endometrial carcinoma have been recently elucidated. In breast carcinoma cells, DHT predominantly exerted antiproliferative effects on mitogenic effects of estrogens (Lapointe & Labrie 2001). Those inhibitory effects were correlated with increased levels of cyclin-dependent kinase inhibitor p21 and/or p27 (Lapointe & Labrie 2001, Grieve et al. 2004). Takagi and coworkers also reported that E2-mediated proliferation was significantly inhibited by DHT in T-47D breast carcinoma cells, which express AR and ER (Takagi et al. 2010). This proliferation was also associated with an increase in 17β-HSD2 expression level and 17β-HSD2 was induced by DHT in a dose-dependent manner. In addition, local E2 concentration was inversely associated with 17β-HSD2 status in breast carcinoma tissues. It is well known that 17β-HSD2 preferentially catalyzes testosterone and E2, to A4 and E1, respectively (Wu et al. 1993). However, 17β-HSD2 expression was reported to be significantly associated with decreased risk to develop late relapse of breast carcinoma (Gunnarsson et al. 2001, 2005).
In addition, significant positive association was reported between the status of androgenic enzymes, 17β-HSD5 and 5αR1, and 17β-HSD2, and in particular, 17β-HSD2 expression was inversely correlated with tumor size in invasive lobular carcinoma of the breast (Yoda et al. 2014). These findings above all indicated that DHT acted, at least partly, through the up-regulation of 17β-HSD2 and decreasing local $E_2$ concentration, and finally played the pivotal roles involved in antiproliferative effects in breast cancer. We have recently reported that 17β-HSD2 immunoreactivity was detected in 37% of endometrial carcinoma cases and correlated with 17β-HSD2 enzymatic activity and expression of 17β-HSD2 mRNA (Utsunomiya et al. 2001). In normal endometrium, 17β-HSD2 immunoreactivity was present at all the cases of secretory phase, but not at any endometrial mucosa of proliferative phase. 17β-HSD2 mRNA expression was consistently detected in endometrial carcinoma compared with adjacent normal endometrium at menopausal status in several studies (Lépine et al. 2010, Cornel et al. 2012, Sinreih et al. 2013). Therefore, the enzyme of 17β-HSD2 should play an important role to modify the balance of estrogen production in not only breast but also endometrial carcinoma. If DHT is involved in the modulation of in situ estrogen metabolism by stimulating the expression of 17β-HSD2, DHT could be one of the important candidates as a new endocrine-related agent in endometrial carcinoma. Further investigation should be required to clarify the therapeutic roles of DHT in endometrial carcinoma.

As we mentioned above, aromatase is a negative regulator for in situ production of DHT. Aromatase-inhibitory therapeutic agents are considered the most effective endocrine therapies in postmenopausal patients with estrogen-dependent breast carcinoma (Bulun et al. 2005). Of particular interest, intratumoral concentration of DHT in breast carcinoma tissue was significantly higher in the patients treated with exemestane, the aromatase inhibitor, than in those who did not receive this therapy (Takagi et al. 2010). This result indicated that aromatase-inhibitory agents could cause increased concentrations and actions of DHT, in conjunction with estrogen deprivation in breast carcinoma. In contrast, there remain some controversies as to whether or not aromatase-inhibitory agents could be effective in the patients with endometrial carcinoma. Previous Gynecologic Oncology Group Study (GOG) could not possibly demonstrate distinct clinical efficacy with aromatase inhibitor treatment (Rose et al. 2000). Thereafter, a prospective phase II trial for fulvestrant was conducted in S3 recurrent or persistent endometrioid endometrial carcinoma (Covens et al. 2011). Patients were stratified into subsets of ER-positive and -negative patients. In ER-positive patients, 3, 13 and 29% cases demonstrated a complete or partial response and stable disease, although no cases were associated with either a complete or a partial response, and 18% demonstrated stable disease in ER-negative patients. Nordic Society of Gynecologic Oncology Study (NSGO) reported similar results (Lindemann et al. 2014). Thangavelu and coworkers examined the biological changes in human endometrial carcinoma tissues before and after aromatase inhibitor treatment (Thangavelu et al. 2013). Patients were randomized to receive anastrozole or placebo before definitive surgery. Treatment with anastrozole caused a marked decrement in proliferation as demonstrated by decreased Ki-67 expression. Sasano previously reported similar results using [3H] thymidine uptake or Ki-67 labeling (Sasano et al. 1999). Therefore, it is important to clarify the roles, including the possibility of application and indication, of aromatase-inhibitory drugs in hormone-receptor positive or hormone-sensitive endometrial carcinoma in postmenopausal patients. In addition, aromatase inhibitor therapy might be established as one form of endocrine treatment of endometrial carcinoma, if aromatase-inhibitory drugs cause not only estrogen deprivation but also increased antiproliferative actions of DHT. Further analysis is required to clarify the clinical importance of DHT actions in association with the therapeutic efficacy of aromatase-inhibitory drugs in endometrial carcinoma.

**Summary and conclusion**

Local androgens exert both direct and indirect impacts on endometrial carcinoma. Testosterone is not only a precursor of estrogen synthesis as the substrate of aromatase, but also a precursor of DHT production as the substrate of 5α-Red enzymes. Recent studies revealed that in situ DHT synthesis occurred at the same time with estrogen synthesis, and DHT signaling could directly exert anticancer effects through DHT-AR pathway in endometrial carcinoma. Aromatase is a negative regulator for in situ production of DHT. If aromatase-inhibitory agents cause not only estrogen deprivation but also increased antiproliferative actions of DHT, combination therapy of DHT and aromatase inhibitor agents could be one of the important candidates as a new endocrine-related agent for endometrial carcinoma especially in postmenopausal patients. However, the downstream signals of AR, which are directly involved in anticancer
Endometrial carcinoma. Further investigation should be required to understand the mechanisms and impacts of anticancer signaling by AR-activation for establishing the possible androgen therapies.

Declaration of interest
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Androgen action in endometrial cancer

K Ito et al.

Androgen action in endometrial cancer


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