Growth hormone and risk for cardiac tumors in Carney complex

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Abstract

Carney complex (CNC) is a multiple neoplasia syndrome that is caused mostly by PRKAR1A mutations. Cardiac myxomas are the leading cause of mortality in CNC patients who, in addition, often develop growth hormone (GH) excess. We studied patients with CNC, who were observed for over a period of 20 years (1995–2015) for the development of both GH excess and cardiac myxomas. GH secretion was evaluated by standard testing; dedicated cardiovascular imaging was used to detect cardiac abnormalities. Four excised cardiac myxomas were tested for the expression of insulin-like growth factor-1 (IGF-1). A total of 99 CNC patients (97 with a PRKAR1A mutation) were included in the study with a mean age of 25.8 ± 16.6 years at presentation. Over an observed mean follow-up of 25.8 years, 60% of patients with GH excess (n = 46) developed a cardiac myxoma compared with only 36% of those without GH excess (n = 54) (P = 0.016). Overall, patients with GH excess were also more likely to have a tumor vs those with normal GH secretion (OR: 2.78, 95% CI: 1.23–6.29; P = 0.014). IGF-1 mRNA and protein were higher in CNC myxomas than in normal heart tissue. We conclude that the development of cardiac myxomas in CNC may be associated with increased GH secretion, in a manner analogous to the association between fibrous dysplasia and GH excess in McCune–Albright syndrome, a condition similar to CNC. We speculate that treatment of GH excess in patients with CNC may reduce the likelihood of cardiac myxoma formation and/or recurrence of this tumor.

Introduction

Carney complex (CNC) is a multiple neoplasia syndrome that affects several organs, including the endocrine glands, heart and skin, and is caused mostly by mutations in the PRKAR1A gene, which codes for the regulatory subunit type 1α of the cyclic AMP-dependent protein kinase A (Stratakis et al. 2001). The most common endocrine tumors in CNC are primary pigmented nodular adrenocortical disease (PPNAD) and growth hormone (GH) and prolactin-producing pituitary adenomas. Antecedent somatotroph hyperplasia without overt adenoma is a common finding, whereas frank acromegaly is seen in only about 15% of the patients

Key Words

- tumor
- acromegaly
- growth hormone
- insulin-like growth factor-1 (IGF-I)
- myxoma
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Materials and methods

Clinical studies

The research protocol was approved by the National Institute of Child Health and Human Development (NICHD) Institutional Review Board. All participants gave written informed consent for clinical trial NCT00001452. Patients were enrolled and screened in this natural history study from 1995 to the current time. Detailed family and medical histories were obtained, and the patients underwent extensive laboratory and imaging evaluations. Data from 269 patients evaluated at the NIH Clinical center for potential CNC were retrospectively reviewed. Only patients who fulfilled the criteria for the diagnosis of CNC\(^1\) and had investigations of their GH secretion and heart imaging at the NIH CRC were included in this study.

Echocardiography was performed in all patients, and cardiovascular magnetic resonance imaging (MRI) in a subset of patients, as part of the patients' initial assessment and routine follow-up. In all patients, GH and IGF-1 levels were measured. All patients underwent screening in order to detect possible GH excess. This was achieved by measuring insulin-like growth factor-1 (IGF-1), GH levels after oral glucose tolerance test (OGTT) (\(n=74\)) and 24-h GH levels (\(n=22\)). In a subset of patients, GH response to thyrotropin-releasing hormone (TRH) test was measured (\(n=13\)). For the evaluation of IGF-1, assay-specific age- and gender-matched normal range levels were considered. The assay for GH changed over the duration of this study as follows: From 1995 to January 1999, double antibody radioimmunoassay technique; from January 1999 to July 2012, chemiluminescence immunoassay on Immulite 2000 analyzer; from July 2012
to present, chemiluminescence immunoassay on Siemens Immulite 2000 XPI. A mean GH level of 2.5 μg/L and above (Ho & Weissberger 1994, Tzanela 2006) indicated normal GH secretion. Regarding GH levels after OGTT and GH response to TRH test, nadir GH level above 1 μg/L and a two-fold increase in GH levels was used as an indication for GH excess, respectively (Kageyama et al. 2005, Tzanela 2006, Chin et al. 2013).

**Table 1** Summary of clinical findings in the 99 study patients with Carney complex (CNC) and cardiac myxomas.

<table>
<thead>
<tr>
<th>Manifestations of CNC</th>
<th>Number of patients (%)</th>
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<tbody>
<tr>
<td>Spotty skin pigmentation (lentiginosis)</td>
<td>84 (84.8)</td>
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<tr>
<td>Cardiac myxoma</td>
<td>47 (47.5)</td>
</tr>
<tr>
<td>Cutaneous or mucosal myxoma</td>
<td>25 (25.3)</td>
</tr>
<tr>
<td>Breast myxomatosis</td>
<td>15 (15.2)</td>
</tr>
<tr>
<td>PPNaD*</td>
<td>61 (61.6)</td>
</tr>
<tr>
<td>Acromegaly</td>
<td>18 (18.2)</td>
</tr>
<tr>
<td>LCCST** or calcification on testicular U/S***</td>
<td>27 (27.3)</td>
</tr>
<tr>
<td>Thyroid carcinoma or multiple hypoechoic nodules</td>
<td>36 (36.4)</td>
</tr>
<tr>
<td>Psammomatous melanotic schwannoma</td>
<td>11 (11.1)</td>
</tr>
<tr>
<td>Blue nevus</td>
<td>42 (42.4)</td>
</tr>
<tr>
<td>Breast ductal adenoma</td>
<td>2 (2.0)</td>
</tr>
<tr>
<td>Osteochondromyxoma</td>
<td>2 (2.0)</td>
</tr>
</tbody>
</table>

*Primary pigmented nodular adrenocortical disease; **Large cell calcifying Sertoli cell tumor; ***U/S, ultrasound.

**Tumor studies**

IGF-1 mRNA and protein expression were tested in samples of cardiac myxomas. Four intraoperative specimens of myxomas from patients with CNC were frozen in liquid nitrogen after excision. The tumors were defrosted and lysed using a Bullet Blender homogenizer (Next Advance, Averill Park, NY, USA) in the appropriate buffer (below).

RNA was isolated with the RNeasy kit (Qiagen) including on-column DNase I treatment. Five hundred nanograms of RNA were reverse-transcribed using Superscript III (Life Technologies), and cDNA levels were quantified in triplicate by real-time quantitative PCR (RT-qPCR) using TaqMan gene expression assays for IGF-1 along with GAPDH on a Viaa Real-Time OCR system (all from Life Technologies) according to manufacturer’s instructions. Normal human heart RNA pooled from three donors was obtained from Clontech.

To study protein expression, myxomas were lysed in T-PER buffer (Thermo Scientific) containing protease inhibitors (Complete, Roche) and protease inhibitors (Complete, Roche). Protein concentrations were determined by BCA protein assay (Thermo Scientific). Normal human heart protein medley was obtained from Clontech. Ten micrograms of lysates were subjected to SDS-PAGE and Western blotting. Blots were probed using α-IGF1 (insulin-like growth factor 1) antibody (Novus, Littleton, CO, USA) followed by α-GAPDH (glyceraldehyde 3-phosphate dehydrogenase) antibody (Santa Cruz Biotechnology) and HRP-conjugated secondary antibodies (Jackson ImmunoResearch). Visualization was achieved by enhanced chemiluminescence on a gel imager (Bio-Rad).

**Statistical methods**

Data were described by frequency distributions and percents or by mean ± standard deviation. Categorical data were compared by Fisher’s exact tests or by the Jonckheere–Terpstra test for doubly ordered data. Logistic regression modeling tested the relation between GH excess and cardiac myxomas or their recurrence, adjusting for covariates as appropriate. Time to penetrance (development of event (first cardiac myxoma)) was estimated using the Kaplan–Meier survival function and is described as median (95% confidence intervals (CIs)); differences in survival distributions between GH excess groups were compared using the log-rank test. P values less than 0.05 and odds ratios (OR) with 95% CI exclusive of 1.0 were considered statistically significant. Data analysis was carried out by SAS v. 9.2 (SAS Institute, Inc, Cary, NC, USA) or Stata v. 11.2 (StataCorp, College Station, TX, USA).

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**Table 2** Growth hormone (GH) evaluation in patients with CNC.

<table>
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<tr>
<th>Evaluation test for GH excess</th>
<th>Patients with GH excess (n=46)*</th>
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<tr>
<td>Elevated GH levels after OGTT test</td>
<td>30 (65.2%)</td>
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<tr>
<td>Elevated IGF-1 levels</td>
<td>27 (58.7%)</td>
</tr>
<tr>
<td>Elevated GH levels after TRH test</td>
<td>10 (21.7%)</td>
</tr>
<tr>
<td>Elevated 24-h GH levels</td>
<td>8 (17.4%)</td>
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*21 patients had GH excess in two or more evaluation tests.
Results

Clinical and genetic epidemiology

Ninety-nine patients fulfilled criteria for entry into study: 42 (42.4%) were male. The group was predominantly white (82.8%), with a mean age of 25.8 ± 16.6 years at initial evaluation at the NIH. Almost all (n = 97) patients had a PRKAR1A mutation. Table 1 provides a summary of findings in these patients.

GH excess and cardiac myxomas

Forty-six (46.5%) of the 99 patients had GH excess by laboratory assessment. Thirty patients had elevated nadir GH levels after OGTT, and 27 presented with increased IGF-1 levels, matched for age and gender. In patients that were not evaluated recently for GH excess, 10 presented with elevated GH levels after TRH stimulation and 8 had increased 24-h GH levels. Twenty-one (45.7%) had GH excess in two or more tests (Table 2).

Figure 1 illustrates the distribution of patients with and without GH excess and with and without cardiac myxoma. The majority (n = 28, 60.9%) of those patients with a history of GH excess had a cardiac myxoma during their lifetime; whereas only one-third (n = 19, 35.9%) of those without GH excess had cardiac myxomas (P = 0.0160). Thus, GH excess was a statistically significant predictor of cardiac myxoma (OR: 2.78, 95% CI: 1.23–6.29; P = 0.0139). Controlling for age at the time of the GH test did not affect the increased odds of occurrence of myxoma (OR: 2.72, 95% CI: 1.17–6.33, P = 0.0202). Although patients with GH excess also tended to have more recurrent myxomas (30.4% vs 18.9%) the difference was not statistically significant (P = 0.2405). However, when we evaluated the absolute number of myxomas occurring within the period of observation, 23 of 47 patients (48.9%) had a single myxoma, 12 (25.5%) had two myxomas, 9 (19.2%) had three myxomas, 1 (2.1%) had four myxomas, and 2 (4.3%) patients had greater than five myxomas. Indeed, patients with the highest number of cardiac myxomas were those with GH excess (P = 0.0154).

Age, cardiac myxomas and GH excess

The median time to detection of a patient’s initial cardiac myxoma was 34.5 years (95% CI: 31.5–45.2 years). Over the observed duration of 65.3 years of follow-up (mean 25.8 years), the Kaplan–Meier survivor function estimated up to 25% chance of having a cardiac mass by age 25.3 years, a 50% chance by 34.5 years and a 75% chance by 50.7 years.
by 46.6 years. In those patients with GH excess, the median time to penetrance was 32.6 years (95% CI: 26.5–44.5). The median time to penetrance in those without GH excess was 39.4 years (95% CI: 33.1–46.6). While there may be a modest earlier manifestation in those with GH excess, the finding was not statistically significant ($P=0.1061$). The median time to presentation with a myxoma was also compared between patients with CNC and GH excess before and after the age of 21 years: those with GH excess had a median time to first myxoma of 34.5 years (95% CI: 28.7–46.2) compared with 45.2 years (95% CI: 33.4–(undefined upper bound)) in those without GH excess ($P=0.0709$) (Fig. 2).

**GH signaling in CNC**

*In vitro* analysis of IGF-1/GAPDH-relative mRNA expression in four cardiac myxomas from separate patients was performed. IGF-1 expression was found to be upregulated in myxomas at the mRNA and protein levels, when compared with pooled normal heart tissue (3 donors) ($P=0.002$) (Fig. 3).

**Figure 3**

IGF-1 mRNA and protein expression in cardiac myxomas. Samples were four different CNC-associated cardiac myxomas (M1–M4) and normal control heart (C). (A) IGF-1 mRNA expression levels were determined by RT-qPCR performed in triplicate. Error bars represent the S.E.M. and show that IGF-1 expression is clearly upregulated in myxomas on the mRNA ($P=0.002$). (B) Western blotting showing the protein expression of IGF-1.

**Treatment for acromegaly and myxomas**

Of the 46 patients with GH excess, 9 (19.6%) underwent transsphenoidal surgery for removal of pituitary adenoma, 1 (2.2%) received radiation therapy and 5 (10.9%) had medical therapy (somatostatin analogs or dopamine agonists). There were 10 patients (21.7%) that had to have a combination of these treatments because of aggressive acromegaly that did not respond to previous therapeutic attempts; of these, 8 had a cardiac myxoma, a difference that was also significant ($P=0.0438$); 2 had recurrent myxoma.

**Discussion**

This study represents the first report of an association between excess GH and increased risk of cardiac myxomas in patients with CNC. The differences between CNC patients with GH excess and those without GH excess were striking: 60.9% of patients with GH excess had a cardiac myxoma versus 35.9% of the patients without GH excess. Certainly, not all cardiac myxomas in CNC patients can be attributed to GH excess, as evidenced by the number of patients with cardiac myxoma in the absence of elevated GH levels. However, GH excess appears to be the only significant risk factor, in addition to the underlying disease, for the development of the cardiac myxoma in patients with CNC.

This finding has significant clinical implications for a potential therapeutic target to decrease myxoma occurrence, i.e. those patients with CNC and excess GH may need earlier and/or more aggressive treatment of hypogonadism. It should be noted, however, that although our data confirm that CNC patients with frank acromegaly who required treatment for their condition had a higher number of cardiac tumor recurrences, only one patient received treatment for acromegaly preceding myxoma development. Thus, we do not know if treatment will alter the likelihood of developing myxomas. Interestingly, the recent paper by Chesnokova and coworkers found that using pegvisomant to block colon cell GH receptors *in vitro* and in patients with acromegaly increased p53 expression, thus suggesting that blocking GH signaling may yield tumor-protective effects (Chesnokova *et al.* 2016).

This study is also the first to report the lifetime prevalence of cardiac myxoma in CNC patients with a 25% risk of developing the tumor by the age of 25 and an increasing risk of developing cardiac myxoma over time. What causes the myxomas remains unclear. Almost all
(97 out of 99) of our patients had a PRKAR1A mutation, which is a slightly higher percentage than previously reported (Bertherat et al. 2009, Horvath et al. 2010). PRKAR1A is a tumor-suppressor gene whose inactivation causes tumors in mice (Kirschner et al. 2005) and abnormal growth and proliferation of human cells in vitro (Nadella et al. 2008, Nesterova et al. 2008). In addition, inactivation of the gene in murine cardiac tissue leads to myxomatous lesions (Yin et al. 2008).

Gender does not appear to play a role in CNC-related myxomas (Carney 1985). Although more than 75% of sporadic cardiac myxomas occur in females (Bjessmo & Ivert 1997, Zheng et al. 2008), this study, in accordance with older data in CNC (Carney 1985), did not show a higher prevalence of myxomas among female patients with CNC. Likewise, GH elevation did not have a different effect in male versus female patients with CNC (data not shown).

Our study does not suggest that GH is a factor in the first molecular events that are needed for the formation of cardiac myxoma, but it does implicate GH and/or IGF-1 in the facilitation of new tumor formation or growth. In the cardiac myxomas we studied after surgical removal, IGF-1 was upregulated compared with normal heart tissue (Fig. 3). GH and/or IGF-1 can promote neoplastic growth of various cell types. However, there is controversy regarding the effect of exogenous GH treatment (Renehan et al. 2004, Raman et al. 2015). Perhaps the best example of increased endogenous GH and/or IGF1 associated with growth of a neoplasm in the context of a multiple neoplasia disorder is that of fibrous dysplasia in patients with McCune–Albright syndrome (MAS) and hypersomatotropinemia (Boyce et al. 2013). Also, bone sarcoma in MAS may be associated with excess GH and/or IGF-1 (Salenave et al. 2014).

It is unclear whether our findings have implications for the sporadic cardiac myxoma. In autopsy studies, myxoma is the most common primary cardiac tumor found, accounting for one-half of all postmortem benign cardiac tumors (McAllister & Fenoglio 1978). It would be interesting to study the GH and/or IGF-1 levels of patients with sporadic myxomas, as well as possible co-occurrence with other tumors, especially those of organs that have been linked to GH and/or IGF-1.

In conclusion, excess GH has a significantly increased risk of cardiac myxoma in patients with CNC. GH excess may be playing an augmentative role in the growth of cardiac myxoma; however, other additional factors likely contribute as 36% of patients without GH excess were found to have cardiac myxoma. This association poses the important clinical question of whether or not early identification and treatment of GH excess in patients with CNC will reduce morbidity and mortality from cardiac myxomas. The time course of the development of cardiac myxoma in CNC underscores the need for affected patients to undergo cardiac surveillance imaging, starting at a young age and continuing indefinitely.

Declaration of interest
The authors have no conflicts of interest related to the manuscript to report. Dr Scherthaner-Reiter reports personal fees from Boehringer Ingelheim, outside the submitted work. Unrelated to the submitted work, Dr Arai has a U.S. Government Cooperative Research and Development Agreement (CRADA) with Siemens. Dr Stratakis also holds patents not directly related to the submission.

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Author contribution statement
Drs Stratakis and Lodish had full access to all of the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis. Arai, Lodish, Stratakis and Carney performed study concept and design. Bandettini, Karageorgiadis, Lodish, Sinaii, Scherthaner-Reiter, Gourgari, Papadakis and Lyssikatos performed acquisition, analysis or interpretation of data. Bandettini, Karageorgiadis, Sinaii, Lodish, Stratakis and Scherthaner-Reiter performed drafting of the manuscript. Bandettini, Karageorgiadis, Sinaii, Rosing, Sachdev, Scherthaner-Reiter, Gourgari, Papadakis, Keil, Lyssikatos, Carney, Arai, Lodish and Stratakis performed critical revision of the manuscript for important intellectual content. Sinaii performed statistical analysis. Sachdev, Scherthaner-Reiter, Gourgari, Papadakis, Keil and Lyssikatos administrative, technical or material support. Arai, Stratakis and Rosing performed study supervision.

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References


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