

# miR-16 is highly expressed in Paget's associated osteosarcoma

## Dear Editor,

Bone is an endocrine organ that produces key hormones and cytokines (Green *et al.* 2015b). Paget's disease of bone (PDB) is a polygenic disorder of bone turnover first described by Sir James Paget in 1876. PDB is characterised by hyper differentiation and hyper activity of osteoclast cells which induces increased bone remodelling by osteoblasts. The resultant mosaic of bone is structurally weaker, larger, more vascular and porous with an increased susceptibility to fracture. On histology, the osteoclasts are increased in size, population and number of nuclei, expressing a 'pagetic phenotype' that distinguishes them from normal osteoclasts. Malignant transformation is a rare complication of PDB reported to arise in <1% of PDB patients. Paget's associated osteosarcoma (PDB-OS) consistently arises in sites of pagetic bone and may present with multifocal lesions (Hansen *et al.* 2006). On histology the lesions are osteoblastic and characterised as an exaggerated form of the accelerated bone remodelling that manifests in PDB. Median survival at diagnosis is 21 months for those treated with curative intent and 7 months for those treated palliatively (Shaylor *et al.* 1999).

PDB has a strong genetic component. A number of loci have been linked to the disorder with sequestosome 1 (*SQSTM1*) variants associated with more severe symptoms, polyostotic foci and heritable transmission (Hansen *et al.* 2006). *SQSTM1* is multifunctional protein which serves as a signalling hub for diverse cellular events including activation of nuclear factor kappa B (NFkB) and tumour necrosis factor superfamily member 11 (TNFSF11). *SQSTM1* also serves as an autophagy receptor for the degradation of ubiquitinated molecules via its ubiquitin binding domain (Katsuragi *et al.* 2015). *SQSTM1* variants associated with PDB are typically located within the coding region of the ubiquitin binding domain. Impairment of autophagy is accompanied by the massive accumulation of *SQSTM1* and formation of *SQSTM1*-positive aggregate structures (Katsuragi *et al.* 2015). As the role of *SQSTM1*

has not been fully elucidated in transformation and there is no transcriptomic analysis of this cancer, we took a next generation sequencing approach to evaluate the expression of small RNAs such as microRNAs (miRNAs) in PDB and PDB-OS.

MiRNAs are key regulators of gene expression through gene silencing. MiRNAs can also be used as biomarkers to classify poorly differentiated cancers and cancer tissue origin (Green *et al.* 2015a). We extracted RNA using the miRCURY RNA isolation kit (Exiqon) from two tissue specimens of PDB-OS with a proven histological diagnosis of osteosarcoma in Paget's affected bone (ages 73 and 81, 2 men). We extracted RNA from four *SQSTM1*-positive PDB tissue specimens obtained from affected trabecular bone (ages 79–87, 2 women and 2 men). We extracted RNA from five control bone tissue specimens obtained from the femoral heads of trauma patients (ages 68–86, 3 women and 2 men). Tissue samples were collected and preserved at –20°C. RNA was stored at –80°C. We generated small RNA libraries using high definition (HD) adapters as previously described (Xu *et al.* 2015). HD adapters increase the annealing efficiency between small RNAs and adapters. An increased annealing efficiency significantly reduces the RNA ligase-dependent ligation bias in next generation sequencing studies (Xu *et al.* 2015). We performed sequencing on the HiSeq 2500 Ultra-High-Throughput Sequencing System (Illumina) at the Earlham Institute, Norwich Research Park.

Raw fastq files were converted to fasta format. The HD signatures of the sequencing reads were trimmed. Quality checking was performed using The UEA Small RNA Workbench ([www.srna-workbench.cmp.uea.ac.uk](http://www.srna-workbench.cmp.uea.ac.uk)). Reads were mapped with no gaps allowed to the human genome v38 using PatMaN. Small RNA expression levels were normalised using a scaling approach, reads per total, to a fixed total of 10 million reads (Mohorianu *et al.* 2011). Comparison of the samples was conducted using scatter plots, size-split boxplot of the replicate-to-replicate

differential expression, intersection and Jaccard similarity analyses (Mohorianu *et al.* 2011). Differentially expressed reads between the control, PDB and PDB-OS samples were identified using both an expression interval approach and pairwise comparison using offset fold change (Mohorianu *et al.* 2011, 2013). Analysis was conducted using custom made Perl (5.24.0.1) and R (3.2.2) scripts. We observed a high proportion of reads matching to the reference genome (Table 1). The size class distributions were bimodal with peaks at 22 and 32 nt (Fig. 1). These peaks correspond to miRNAs and tRNA fragments which was confirmed using annotations. The small number of unique sequences with high abundance is also visible in the complexity distributions (Fig. 1). The lower complexities correspond to 22 and 32 nt which indicate a small number of highly abundant sequences (Fig. 1). As with all human studies there was variability in the number of reads assigned to either miRNAs or tRNA fragments in the control samples (when compared to inbred genetic models such as mice and fruit flies where variability is minimal). We identified a low expression of miR-16 in PDB compared to controls and a high expression of miR-16 in PDB-OS compared to controls and PDB. We also identified a downregulation of miR-144 and upregulation of miR-21 in PDB-OS compared to controls. Downregulation of miR-144 is in line with previous data in osteosarcoma cells. Upregulation of miR-21 is associated with many types of cancer. In this study we explored miR-16 further.

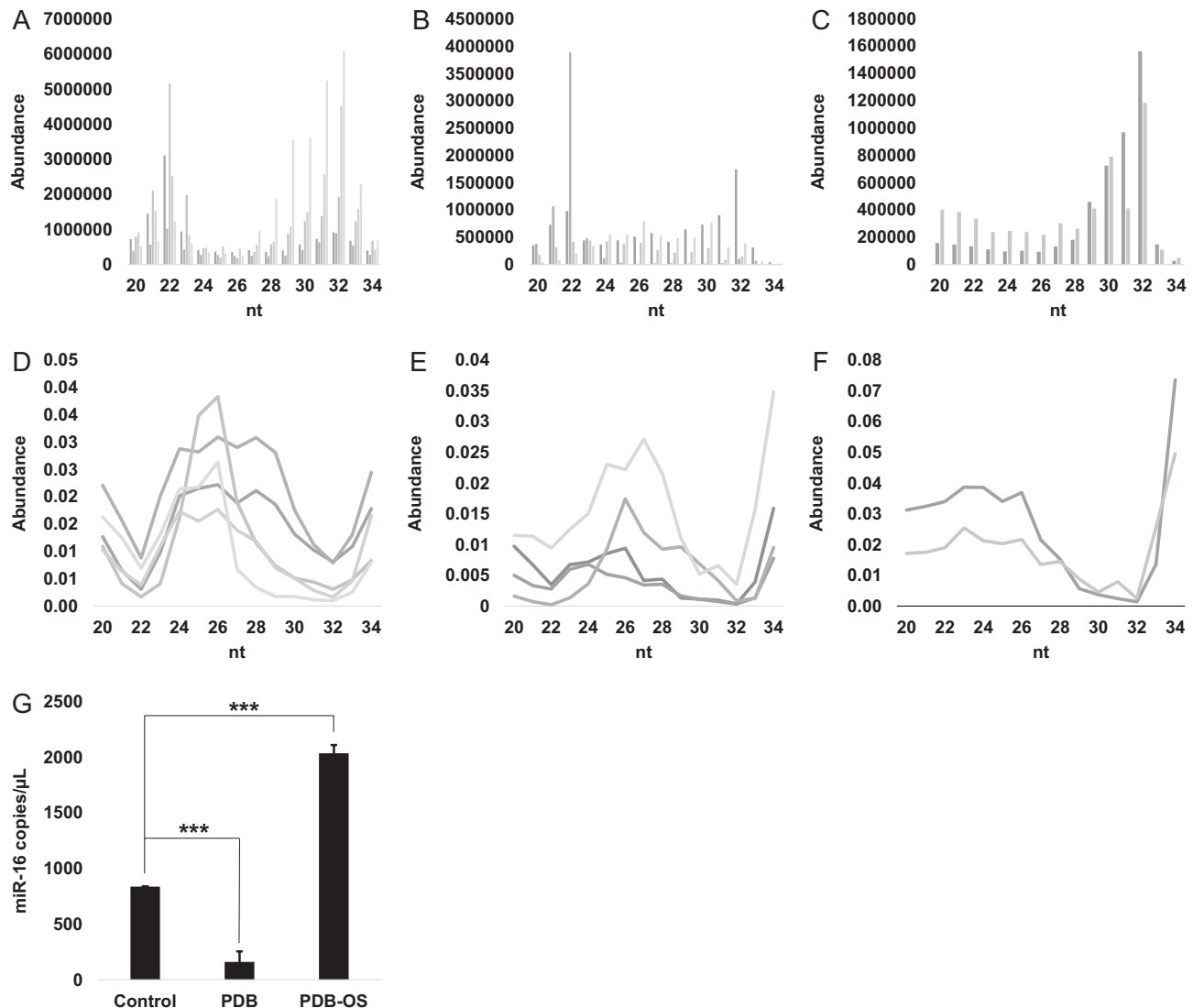
Total RNA used for next generation sequencing was quantified by density measurement after separation by agarose gel electrophoresis with ethidium bromide staining. Equal amounts of RNA across the sample pools were reverse transcribed using the TaqMan advanced

miRNA cDNA synthesis kit (Thermo Fisher Scientific). Differential expression of miR-16 was validated three times in triplicate using TaqMan miRNA advanced assays (Thermo Fisher Scientific). Digital PCR was performed on the QuantStudio 3D Digital PCR System using the GeneAmp PCR System 9700 (Thermo Fisher Scientific). After PCR the chips were imaged on the QuantStudio 3D Instrument which assesses the raw data and calculates the concentration of the cDNA sequence targeted by FAM and VIC labelled probes by Poisson distribution (Fazekas de St 1982). For more in depth analysis the QuantStudio 3D AnalysisSuite was used to report the data as copies/ $\mu$ L. The probe sequence used was miR-16-5p (5'-UAGCAGCACGUAAAUAUUGGCG-3'). We confirmed the downregulation of miR-16 in PDB compared to controls and upregulation of miR-16 in PDB-OS compared to controls and PDB (Fig. 1).

To elucidate a mechanistic role of miR-16 in PDB-OS we used TargetScan v7.1 to search for its messenger RNA targets. *SQSTM1* was identified as a candidate, which was supported by performing a second search and identification on miRSearch v3. Target identification of miRNAs is computationally difficult due to the relatively low homology between miRNAs and their targets. We searched the literature and identified *SQSTM1* as an experimentally confirmed target of miR-16, i.e. levels of *SQSTM1* are reduced by miR-16 overexpression (Selbach *et al.* 2008). We find that miR-16 is downregulated in PDB which means there is a loss of *SQSTM1* negative regulation. In line with previous data *SQSTM1* variants in PDB are activating mutations. *SQSTM1* is a signalling hub for the activation of TNFSF11. TNFSF11 is the primary ligand for stimulating the differentiation of monocyte progenitors

**Table 1** Overview of the sequencing output for control (Ctrl), PDB and PDB-OS including the total number of reads obtained per sample, total number of accepted reads which did not contain unassigned nucleotides, total number of non-redundant reads and overall sample complexities (C) which is defined as the ratio of non-redundant to redundant reads. We also show the percentage of genome matching reads assigned to miRNAs and tRNAs.

	Total reads	Accepted reads	Adapter reads	Non-redundant reads	C	Genome match (%)	miRNA match (%)	tRNA match (%)
Ctrl	26,735,842	26,711,859	17,171,584	911,323	0.057	73	28	9
Ctrl	14,090,445	14,077,678	10,211,115	741,722	0.079	71	13	14
Ctrl	28,948,651	28,922,976	25,836,859	800,979	0.034	80	34	31
Ctrl	26,936,015	26,911,371	24,367,300	811,143	0.034	84	15	39
Ctrl	35,001,366	34,969,913	33,113,846	653,789	0.020	87	5	74
PDB	8,966,345	8,954,278	8,002,650	514,395	0.067	63	4	38
PDB	13,553,618	13,535,611	12,801,465	675,943	0.053	73	12	39
PDB	18,652,438	18,627,791	14,967,804	457,315	0.031	43	37	1
PDB	11,683,775	11,668,436	9,452,776	896,236	0.095	60	1	20
PDB-OS	14,861,235	14,848,062	12,927,808	484,501	0.057	58	0.4	46
PDB-OS	21,954,888	21,935,518	17,080,048	988,000	0.094	52	1	26

**Figure 1**

Bioinformatics analysis and digital PCR results. (A and D) Redundant size class distributions and complexity in control samples (B and E) Redundant size class distributions and complexity in PDB samples (C and F) Redundant size class distributions and complexity in PDB-OS samples (G) Digital PCR for miR-16 expression performed with pooled RNA obtained from control bone, PDB and PDB-OS tissue. Data is reported as copies/μL as calculated by Poisson distribution. Error bars represent the standard deviation. Statistical significance (\*\*\* $P < 0.01$ ) was calculated using an unpaired t-test with Prism 6 (GraphPad) software.

to osteoclasts through interaction with tumour necrosis factor receptor superfamily member 11a (TNFRSF11A). TNFRSF11 is produced by osteoblasts and downstream repercussions of the TNFRSF11–TNFRSF11 interaction is the increase of bone resorption. Bone resorption achieved by various osteoclast-produced cytokines is set within a positive feedback loop which encourages osteoblast proliferation and activity. Increased proliferation of osteoblast cells containing *SQSTM1* activating variants repeats the bone remodelling cycle each time increasing the number of *SQSTM1* positive osteoblasts. PDB does

not typically present until patients are >55 years old. In patients harbouring *SQSTM1* variants, presentation can appear earlier. Presentation may be due to the speed of the bone remodelling cycle or the gradual loss of miR-16 mediated silencing of *SQSTM1*.

We find a high expression of miR-16 in PDB-OS. The effect of upregulated miR-16 means there is an increased negative regulation of genes including *SQSTM1*. The loss of *SQSTM1* has little effect on autophagy (Katsuragi *et al.* 2015). The lack of phenotypic impact might be explained by the presence of other autophagy

regulators including optineurin (OPTN) (Katsuragi *et al.* 2015). As a signalling hub SQSTM1 is a partner of the Kelch-like ECH-associated protein 1 (KEAP1)-nuclear factor erythroid 2-like 2 (NFE2L2) pathway, a major cellular defence mechanism against oxidative stress (Katsuragi *et al.* 2015). In normal conditions NFE2L2 is constitutively degraded by the ubiquitin-proteasome system because its binding partner KEAP1 is an adapter of ubiquitin ligases. Upon exposure to oxidative stress KEAP1 is structurally modified which releases NFE2L2 to translocate to the nucleus. NFE2L2 is a transcription factor which then induces a multitude of anti-oxidant and anti-inflammatory genes. SQSTM1 is able to abrogate the interaction between KEAP1 and NFE2L2 leading to protection against oxidative stress (Katsuragi *et al.* 2015). We propose that miR-16-mediated silencing of *SQSTM1* incapacitates the cell's ability to protect itself against oxidative stress-induced oncogenesis. This proposal warrants further molecular investigation to show an inverse correlation between the expression of miR-16 and *SQSTM1*.

Molecular testing may help provide a robust diagnosis and is particularly useful in rare cancers. This is especially true in PDB where transformation to osteosarcoma is often missed until late stage. Radiograph morphology is subtly different between PDB and PDB-OS. Symptoms of PDB-OS can be similar to the day-to-day symptoms experienced by a PDB patient. Biomarkers such as serum total alkaline phosphatase used in the diagnosis of PDB are the same biomarkers used to aid the diagnosis of PDB-OS. These features combined make it difficult to distinguish between a chronic non-life threatening disorder and a fatal disease which require very different treatment strategies. Quantitative PCR or next generation sequencing performed on RNA extracted from a biopsy of a presenting lesion is precise for detecting a reduced expression of miR-16 in 'normal PDB' and high expression of miR-16 in PDB-OS. The change in miR-16 expression could alert physicians to the change to an osteosarcoma phenotype.

A limitation of this study is the size of the cohorts studied. PDB-OS arises in <1% of PDB patients and donation to tissue banks is scarce. We are in the process of collecting a larger cohort of tissue specimens to validate the findings from this study. Our data highlights the value of being able to provide a robust tissue diagnosis in addition to identifying regulatory transcriptomic molecules that could be exploited for targeted therapy. Expression of miR-16 could also be investigated in

paediatric osteosarcoma to evaluate similarities and/or differences between the two age-related incidence peaks of osteosarcoma.

## Data availability

The data presented in this study is publicly available on Gene Expression Omnibus under the accession numbers GSE85809 (GSM2284729 to GSM228473 are control samples; GSM2284736 and GSM2284736 are PDB-OS samples) and GSE87018 (GSM2318966 to GSM2318970 are PDB samples).

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## Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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