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GLUT12 promotes prostate cancer cell growth and is regulated by androgens and CaMKK2 signaling

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Abstract

Despite altered metabolism being an accepted hallmark of cancer, it is still not completely understood which signaling pathways regulate these processes. Given the central role of androgen receptor (AR) signaling in prostate cancer, we hypothesized that AR could promote prostate cancer cell growth in part through increasing glucose uptake via the expression of distinct glucose transporters. Here, we determined that AR directly increased the expression of SLC2A12, the gene that encodes the glucose transporter GLUT12. In support of these findings, gene signatures of AR activity correlated with SLC2A12 expression in multiple clinical cohorts. Functionally, GLUT12 was required for maximal androgen-mediated glucose uptake and cell growth in LNCaP and VCaP cells. Knockdown of GLUT12 also decreased the growth of C4-2, 22Rv1 and AR-negative PC-3 cells. This latter observation corresponded with a significant reduction in glucose uptake, indicating that additional signaling mechanisms could augment GLUT12 function in an AR-independent manner. Interestingly, GLUT12 trafficking to the plasma membrane was modulated by calcium/calmodulin-dependent protein kinase kinase 2 (CaMKK2)-5′-AMP-activated protein kinase (AMPK) signaling, a pathway we previously demonstrated to be a downstream effector of AR. Inhibition of CaMKK2-AMPK signaling decreased GLUT12 translocation to the plasma membrane by inhibiting the phosphorylation of TBC1D4, a known regulator of glucose transport. Further, AR increased TBC1D4 expression. Correspondingly, expression of TBC1D4 correlated with AR activity in prostate cancer patient samples. Taken together, these data demonstrate that prostate cancer cells can increase the functional levels of GLUT12 through multiple mechanisms to promote glucose uptake and subsequent cell growth.

Key Words

- GLUT12
- SLC2A12
- glucose metabolism
- androgen receptor (AR)
- calcium/calmodulin-dependent protein kinase kinase 2 (CaMKK2)
- 5′-AMP-activated protein kinase (AMPK)
- prostate cancer
Introduction

Altered cell metabolism is now acknowledged as one of the emerging hallmarks of cancer (Hanahan & Weinberg 2011). Common among most cancer cells is an increased ability to take up and metabolize large amounts of glucose to help meet the growing cells’ energetic and anabolic demands. The first rate-limiting step in glucose metabolism is the cellular uptake of the sugars (Hatanaka 1974). This is done primarily by a family of facilitative glucose transporters (GLUTs). To date, fourteen GLUTs have been identified in humans (Barron et al. 2016). While many of these transporters have known functions in basic physiology (ex. GLUT4 functions in muscle), growing evidence indicates roles for some of these transporters in cancer (Luiken et al. 2015, Barron et al. 2016, Labak et al. 2016). To that end, multiple members of the GLUT family are overexpressed in various cancer types (reviewed in (Barron et al. 2016)). Not surprisingly, several oncogenic and tumor suppressive signaling pathways have been shown to regulate glucose uptake (Cairns et al. 2011). The delineation of which specific transporters play a functional role in the disease and the determination of how they are regulated are needed to improve our understanding of this important aspect of pathogenic cell biology.

Prostate cancer is the most commonly diagnosed non-cutaneous cancer in US men (American Cancer Society 2015). One of the central drivers of the disease is the androgen receptor (AR). Despite AR's established role in prostate cancer (Schmidt & Tindall 2013), our understanding of the specific downstream events that are regulated by AR and that promote the disease is not complete. Recent work from both our laboratory as well as others has demonstrated that androgen signaling increases glucose metabolism (Massie et al. 2011, Moon et al. 2011, Itkonen et al. 2013, Tennakoon et al. 2014, Tsouko et al. 2014). This augmented carbohydrate metabolism was mediated in part through a signaling cascade involving the calcium/calmodulin-dependent protein kinase kinase 2 (CaMKK2) and the 5′-AMP-activated protein kinase (AMPK) (Massie et al. 2011, Tennakoon et al. 2014). Given the requirement for initial glucose uptake prior to its subsequent metabolism, we hypothesized that AR signaling promoted glucose metabolism by increasing the expression and/or activity of select GLUTs in prostate cancer. We further speculated that AR could increase glucose uptake in part through CaMKK2-AMPK signaling. Here, we sought to determine whether androgens increased the expression of any of the GLUTs that were overexpressed in prostate cancers relative to benign tissue. In addition, we wanted to determine the mechanism(s) behind this regulation to aide future efforts to treat the disease.

Materials and methods

Cell culture and reagents

LNCaP, C4-2, VCaP, 22Rv1 and PC-3 human prostate cancer cell lines and RWPE-1 immortalized prostate epithelial cells were obtained from American Type Culture Collection (ATCC). LAPC4 cells were a gift from Dr Charles Sawyers (Memorial Sloan-Kettering Cancer Center, New York, NY, USA). Cells were authenticated using short-tandem repeat analysis. Cells were also validated using a PCR-based assay to ensure the cells were free of mycoplasma. All experiments were performed using cells that were <10 passages. LNCaP, VCaP, LAPC4, C4-2 and 22Rv1 cells were maintained as previously described (Frigo et al. 2011, Tennakoon et al. 2014, Blessing et al. 2015). RWPE-1 cells were maintained in keratinocyte serum-free medium (K-SFM; contains 1.04 g/L d-glucose) supplemented with bovine pituitary extract and human recombinant epidermal growth factor per ATCC’s recommendations. PC-3 cells were maintained in RPMI-1640 (Mediatech, Manassas, VA, USA) supplemented with 8% fetal bovine serum (FBS; Sigma). Prior to all experiments, cells were plated and incubated for 72 h in media that were phenol-red-free and contained charcoal-stripped FBS (CS-FBS; Gemini Bio-Products, West Sacramento, CA, USA). RWPE-1 cells were switched to phenol-red-free RPMI-1640 medium for experiments.

Methyltrienolone (R1881) was purchased from PerkinElmer. Enzalutamide was from SelleckChem (Houston, TX, USA). STO-609 was purchased from Tocris (Bristol, UK). Anti-AMPK and anti-p-AMPK (T172) antibodies were purchased from Cell Signaling. Anti-GLUT12 antibody was purchased from Bioss (Woburn, MA, USA). Anti-TBC1D4, anti-p-TBC1D4 (T642), anti-ATPase antibodies and the Plasma Membrane Protein Extraction Kit were purchased from Abcam. Mission siRNAs targeting SLC2A12 (GLUT12) and TBC1D4, universal siRNA-negative control and anti-GAPDH antibody were purchased from Sigma-Aldrich. Stealth siRNAs targeting control or AMPK were purchased from Santa Cruz Biotechnology.
RNA isolation, cDNA synthesis and quantitative PCR (qPCR)

RNA isolation, cDNA preparation and qPCR were carried out as previously described using RPLP0/36B4 as an internal control (Frigo et al. 2011). All primers used in this study are listed in Supplementary Table 1 (see section on supplementary data given at the end of this article).

Immunoblot analysis

Immunoblot analysis was performed as previously described (Tennakoon et al. 2014). All antibodies were used at the manufacturers’ recommended concentrations. Results shown are representative blots. Densitometry was performed using ImageJ software (National Institutes of Health (NIH)), and samples were normalized to indicated controls. Results are presented as normalized mean values ± S.E.M. from experimental repeats (n ≥ 3).

Bioinformatic analyses of gene expression in clinical datasets

For the gene expression signature comparisons, transcriptomic profiles of human prostate cancer cohorts were downloaded from GEO (GSE21034, Taylor et al. (2010); GSE35988, Grasso et al. (2012)) or The Cancer Genome Atlas (TCGA). Androgen-induced signatures (Hieronymus AR and Nelson AR) were generated from previously defined data (Nelson et al. 2002, Hieronymus et al. 2006). For each of the signatures, an activity score for each sample in each cohort was generated as described previously (Taylor et al. 2010). Briefly, gene expressions of prostate cancer cohorts were converted to z scores with respect to normal samples. The activity score for each sample for a signature was evaluated by adding the z scores of upregulated genes and subtracting the z scores of downregulated genes. Correlation between pairs of gene signature activity scores were evaluated using the Pearson correlation coefficient as implemented in the Python statistical library SciPy; significance was assessed at P < 0.05.

RNAi transfection

Transient transfections of siRNAs were performed as previously described (Blessing et al. 2015) with the lone exception that PC-3 cells were transfected twice 24 h apart with siRNAs to achieve sufficient knockdown. The sequences of the siRNAs are listed in Supplementary Table 1.

Proliferation assay

Cell proliferation assays were carried out as previously described by measuring the cellular double-stranded DNA content using a fluorescent DNA stain (Shi et al. 2013).

Glucose uptake assay

LNCaP, VCaP or RWPE-1 cells were plated at a density of 2.5 × 10^4 cells per well in a 6-well plate while PC-3 cells were plated at a density of 1.5 × 10^5 cells per well in a 6-well plate. Cells were transfected and treated (LNCaP, VCaP and RWPE-1 cells only) for 24 h ± 10 nM R1881 in starvation medium (phenol-red free RPMI-1640 medium alone containing 2 g/L d-glucose with 10 mM HEPEs (Sigma) and 0.1% bovine serum albumin (BSA; Sigma)). Afterward, medium was switched to phenol-red-free RPMI-1640 medium (LNCaP, VCaP, PC-3) or K-SFM (RWPE-1) for 2 h. The medium was then collected and analyzed using an YSI 2700 Bioanalyzer (YSI Life Sciences, Yellow Springs, OH, USA). Glucose uptake levels were normalized to cellular DNA amounts.

Chromatin immunoprecipitation sequencing (ChiP-Seq)

ChiP-Seq analysis was performed as previously described (Chng et al. 2012, Tan et al. 2012).

Reporter gene assays

Cells were transfected with plasmids using Lipofectamine 2000 (ThermoFisher Scientific) according to the manufacturer’s instructions and then treated with hormones approximately 16 h before the assay. Luciferase and β-galactosidase (transfection normalization) activities were measured as previously described (Blessing et al. 2015). Each treatment was performed in triplicate, and results are expressed as mean relative light units (RLUs) normalized to β-galactosidase transfection control ± s.e. Each experiment was repeated at least 3 times with a representative experiment shown. The pGL4.26-SLC2A12 (SLC2A12-Luc) enhancer construct was created by PCR amplifying the sequence (forward primer: 5′-CAGTCGGTACCTACCCTCCCTGGATTTCAAAAT-3′; reverse primer: 5′-CGAGGCTCGAGCTTTACCTACCCTCCCTGGATTTCAAAAT-3′) that encompassed the potential AR-binding site identified using ChiP-Seq (Chng et al. 2012, Tan et al. 2012) and shown in Supplementary Fig. 6 that was located within intron 2 of SLC2A12. This fragment
was then cloned into the pGL4.26 vector (Promega) using KpnI and XhoI restriction sites. The ARE deletion plasmids were generated by overlap extension PCR. Primary DNA fragments were amplified from full-length enhancer PCRs generated above using Hot Start Phusion polymerase (NEB, Ipswich, MA, USA) and deletion primers (ARE-1 deletion, forward primer: 5′-GTGTAATAGAGGTA TGAAGAAAAAGTAACTTTGAAGCAAAGATTAA-3′, reverse primer: 5′-TAAATCTGCTTTCAATGTACCTTTTT TTCTTCTACCTCATTACAC-3′; ARE-2 deletion, forward primer: 5′-TGAGTTTTAAGAGATTTTATTATACC AGATCTGGAATGCATTAC-3′, reverse primer: 5′-GTAAA TGGCATTCAAGATCTGGTATAATAAAAA TCTTCTTAAACTCCA-3′) combined with the above primers in the appropriate pairs to generate single or double deletions. The resulting PCR products were purified and eluted into 10 µL water with a Zymo Clean and Concentrator Kit (Irvine, CA, USA) following the manufacturer's instructions. Full-length deletion fragments were generated in a second PCR by mixing 2 µL of the appropriate 2 or 3 purified fragments to produce the desired deletion mutants. The resulting products were subsequently purified and cloned using the same procedure as the full-length enhancer. All sequences were confirmed using restriction digests and sequencing. The resulting PCR products were subsequently purified and cloned using the same procedure as the full-length enhancer. All sequences were confirmed using restriction digests and sequencing. The KLK3 (PSA)-Luc reporter plasmid was previously described (Frigo et al. 2011).

**Creation of stable cell lines**

LNCaP cells that stably express CAMKK2 under the control of a doxycycline-inducible promoter were made by first creating a pLNDUCER20-CAMKK2 lentiviral construct using the Gateway recombinein cloning system. To do this, the CAMKK2-coding sequence was shuttled from pOTB7-CAMKK2 (American Type Tissue Culture) to pLNDUCER20 (gift from Thomas Westbrook). LNCaP cells were then infected with lentivirus expressing pLNDUCER20-CAMKK2, and cells were selected using G418 (Sigma).

22Rv1 cells that stably express an shRNA targeting CAMKK2 under the control of a doxycycline-inducible promoter were made by first creating a pLNDUCER10-shCAMKK2 lentiviral construct via subcloning an shRNA targeting CAMKK2 from pGIPZ-shCAMKK2 (Baylor College of Medicine C-BASS Core) into pLNDUCER10 (gift from Thomas Westbrook) using XhoI and MluI restriction sites. 22Rv1 cells were then infected with lentivirus-expressing pLNDUCER10-shCAMKK2, and cells were selected using puromycin (Sigma).

**Plasma membrane fractionation**

Cells were plated at a density of 6 × 10⁶ cells per 45 × 120 mm dish. Three dishes were used per treatment group to total 18 × 10⁶ cells. Cells were then transfected with siRNAs and/or treated as indicated. After transfection/treatment, cells were harvested for whole-cell lysates or plasma membrane subcellular fractionation. The fractionation was performed according to manufacturer's instructions (Abcam). Protein levels were determined using immunoblot analysis.

**Statistical analysis**

Unless otherwise noted, two-sample comparisons were performed using Student's t tests. Multiple comparisons were performed using one-way ANOVA and post hoc Dunnett's test with GraphPad Prism, version 5 (GraphPad Software). Statistically significant changes were determined at the P<0.05 level.

**Results**

**Identification of SLC2A12 as a direct AR-regulated gene that is overexpressed in prostate cancer**

We first sought to determine if AR signaling, a major driver of prostate cancer (Schmidt & Tindall 2013), increased the expression of members of the GLUT family. To test this, AR+ LNCaP prostate cancer cells were treated for 24 or 72 h with vehicle or the synthetic androgen R1881. Cells were then lysed and subjected to qRT-PCR analysis to quantitate the mRNA levels of the 14 known GLUT family members. Of the 14 members, only two (SLC2A2 (GLUT2) and SLC2A14 (GLUT14)) could not be detected at the mRNA level (Fig. 1A). Three family members (SLC2A3 (GLUT3), SLC2A10 (GLUT10) and SLC2A12 (GLUT12)) were found to be androgen responsive (Fig. 1A). While androgen treatment increased SLC2A3 mRNA levels, this observation did not correspond to an increase in the functional protein levels of GLUT3 as siRNA-mediated knockdown of SLC2A3 had no effect on androgen-mediated glucose uptake (D Frigo, unpublished observations). These data are consistent with previous reports of high SLC2A3 mRNA levels being detected in the absence of GLUT3 protein expression (Simpson et al. 2008). Like SLC2A3, androgens also increased the mRNA levels of SLC2A10 twofold (Fig. 1A). However, the baseline mRNA levels of SLC2A10 were low (Ct values ~31–32 cycles) and, more importantly, SLC2A10 expression did...
not correlate with prostate cancer in clinical samples (Welsh et al. 2001, Vanaja et al. 2003, Varambally et al. 2005, Taylor et al. 2010, Grasso et al. 2012). Conversely, androgens increased the expression of SLC2A12 (Fig. 1A), the expression of which was also significantly elevated in prostate cancer patient samples relative to benign controls in multiple clinical cohorts (Table 1) (Welsh et al. 2001, Vanaja et al. 2003, Varambally et al. 2005, Taylor et al. 2010, Grasso et al. 2012). Consistent with these findings, androgen treatment increased the expression of SLC2A12 in additional AR+ prostate cancer cell models (LAPC4 and VCaP; Supplementary Fig. 1). Androgens also increased the protein levels of GLUT12 in multiple cell models (Fig. 1B). Conversely, we were unable to detect GLUT12 in non-transformed RWPE-1 prostate epithelial cells (Supplementary Fig. 2). Further analysis indicated that the peak androgen-mediated expression of GLUT12 in prostate cancer cells occurred ~24 h after initial treatment (Fig. 1C), indicating that androgens promoted a pulse of GLUT12 synthesis. To assess whether AR could also regulate SLC2A12 in patients, we leveraged a previously published, curated AR gene signature of identified AR target genes (genes that were increased in response to androgens and modulated by AR antagonists) (Hieronymus et al. 2006). Using an informatics approach, we determined that this AR gene signature positively correlated with increased SLC2A12 mRNA transcript levels in a well-known clinical prostate cancer cohort (Fig. 1D), suggesting AR also regulates the expression of SLC2A12 in patients (Taylor et al. 2010). In support of this, similar results were obtained using this AR activity signature as well as an additional AR activity signature (Nelson et al. 2002).
across multiple clinical cohorts (Taylor et al. 2010, Grasso et al. 2012) including The Cancer Genome Atlas (TCGA) (Cerami et al. 2012). As a comparison, AR activity did not correlate with the expression of other genes such as GAPDH and TGFβ1 and inversely correlated with GLRA2 and CD200 (Supplementary Fig. 3), two genes previously demonstrated to be downregulated by AR (Hieronymus et al. 2006).

To determine how androgens increased SLC2A12 expression, we first evaluated the role of AR to verify that this was a receptor-mediated event. Knockdown of AR using two separate, validated siRNAs (Tsouko et al. 2014, Blessing et al. 2015, 2017) blocked the androgen-mediated expression of SLC2A12 (Fig. 2A). As further validation, co-treatment of prostate cancer cells with the antiandrogen enzalutamide suppressed androgen-mediated SLC2A12 expression (Fig. 2B and Supplementary Fig. 4). Enzalutamide’s inhibition of androgen-mediated SLC2A12 expression was overcome by saturating concentrations of the agonist, confirming AR’s role (Fig. 2B).

Androgen treatment increased SLC2A12 mRNA expression as early as 6 h after treatment (Supplementary Fig. 5), suggesting that SLC2A12 could be a direct AR target gene. Hence, we next wanted to determine whether SLC2A12 was a direct AR target. To do this, we initially co-treated prostate cancer cells with androgen in the presence or absence of cycloheximide, an inhibitor of translation (Fig. 2C). As previously described, cycloheximide treatment did not block androgen induction of FKBP5, an established direct target of AR (Frigo et al. 2009). Similar to FKBP5, cycloheximide treatment did not impair the androgen-mediated expression of SLC2A12 (Fig. 2C), suggesting that SLC2A12 was also a direct AR target. Mining of existing chromatin-immunoprecipitation sequencing (ChIP-Seq) datasets from several prostate cancer cell models indicated that AR directly bound to an intronic region of SLC2A12 in the presence of dihydrotestosterone (DHT) (Fig. 2D and Supplementary Fig. 6) (Chng et al. 2012, Tan et al. 2012). To confirm that the identified AR-binding region contained a functional androgen response element (ARE) and thus primary AR target, we cloned out the genomic region surrounding the identified binding site and tested its ability to confer androgen responsiveness to an enhancerless luciferase reporter gene. While not as robust as the well-known KLK3 ARE, the identified SLC2A12 AR-binding site indeed conferred androgen responsiveness, indicating that SLC2A12 was also a direct AR target (Fig. 2E).

**GLUT12 is required for maximal glucose uptake and cell growth in prostate cancer cells**

To investigate the functional importance of GLUT12, we depleted GLUT12 levels using siRNAs in diverse prostate cancer cell models (Fig. 3A). Interestingly, GLUT12 was still expressed in the AR-negative PC-3 cells, suggesting that in this aggressive prostate cancer model, additional, AR-independent mechanisms maintain GLUT12 expression. GLUT12 knockdown blocked androgen-mediated glucose uptake (Fig. 3B) and proliferation (Fig. 3C) in LNCaP and VCaP cells. Knockdown of GLUT12 also decreased glucose uptake (Fig. 3B) and proliferation (Fig. 3C) in PC-3 cells. These effects were not unique to LNCaP, VCaP and PC-3 cells as GLUT12 knockdown impaired the growth of additional castration-resistant prostate cancer (CRPC) cell models including C4-2 and 22Rv1 (Fig. 3C). Collectively, these results demonstrate that GLUT12 is required for maximal androgen-mediated glucose uptake and cell growth in hormone-sensitive prostate cancer cells and may also be required for basal glucose uptake and cell growth in some CRPC cell models.
Figure 2

SLC2A12 is a direct AR target. (A) LNCaP cells were transfected with siRNAs targeting a scramble sequence (siControl) or AR and then treated ± androgen (10 nM R1881) for 72 h. SLC2A12 relative mRNA expression was quantified via qRT-PCR and data were normalized to 36B4 levels. *Significant \((P<0.05)\) changes from vehicle treatment. (B) LNCaP cells were co-treated ± 10 µM enzalutamide (antiandrogen) with increasing concentrations of androgen (0, 100 pM or 10 nM R1881) for 72 h. SLC2A12 mRNA levels were quantified using qRT-PCR and normalized to 36B4 levels. *Significant \((P<0.05)\) changes from vehicle (no androgen) treatment. #Significant \((P<0.05)\) changes from vehicle (no enzalutamide) treatment. (C) LNCaP cells were treated for 16 h ± 1 µg/µL cycloheximide ± androgen (10 nM R1881). FKBP5 is a direct transcriptional target of AR (Frigo et al. 2009). CXCR4 is an indirect transcriptional target of AR (Frigo et al. 2009). Data are normalized to 36B4 levels. *Significant \((P<0.05)\) changes from double vehicle. (D) ChIP-Seq tracks of LNCaP cells treated with vehicle or DHT for 2 h. An AR binding site located within an intronic region of SLC2A12 is highlighted. Similar data for VCaP and C4-2B cells are presented in Supplementary Fig. 6. (E) Enhancer luciferase reporter constructs including a known enhancer of the KLK3 (PSA) gene (left graph; positive control) or the AR-binding region identified in (D) with intact or candidate AREs deleted (right graphs) were transfected into LNCaP cells and treated overnight with 0, 0.1 or 10 nM androgen (R1881). After treatment, cells were harvested and assayed for luciferase activity. Luciferase values were normalized to the β-galactosidase transfection control. Data are the mean relative light units (RLUs) ± s.e.m. for one representative experiment conducted in triplicate \((n=3)\). *Significant \((P<0.05)\) changes from vehicle-treated cells.
TBC1D4 is regulated by AR and CaMK2-AMPK signaling

While SLC2A12 is a direct AR target, the activity of several GLUTs can be controlled by additional regulatory mechanisms (Barron et al. 2016). Previous work from our laboratory and others demonstrated that androgens increased glucose metabolism and prostate cancer cell growth in part through a CaMK2-AMPK-mediated mechanism (Massie et al. 2011, Tennakoon et al. 2014). Accordingly, inhibition of CaMK2 using the antagonist STO-609 or depletion of AMPK using siRNAs (Supplementary Fig. 7) decreased androgen-mediated LNCaP prostate cancer cell growth (Fig. 4A and B) and basal cell growth in AR-negative PC-3 cells (Supplementary Fig. 8A and B). This occurred in LNCaP cells despite the fact that knockdown of AMPK led to an increase in AR (perhaps as a compensatory survival mechanism) (Supplementary Fig. 7), consistent with the work of others suggesting that AMPK may feedback to inhibit AR under certain contexts (Jurmeister et al. 2014, Shen et al. 2014). In contrast, STO-609 appeared to decrease basal AR levels (Fig. 4C), matching previous reports that indicated CaMK2 signaling could augment AR activity (Karacosta et al. 2012, 2016). However, this effect was lost in the presence of androgens. Of interest, AMPK has been shown to promote glucose uptake through the phosphorylation and regulation of tre-2/USP6, BUB2, cdc16 domain family member 4 (TBC1D4; also called Akt substrate of 160 kDa (AS160)), a protein that controls the trafficking of GLUT-containing vesicles (Kramer et al. 2006, Treebak et al. 2006). To determine whether this regulation also occurred in prostate cancer, we treated cells ± STO-609 ± androgen and quantified the levels of T642 phosphorylated TBC1D4 (known AMPK target site (Kramer et al. 2006, Treebak et al. 2006)). As predicted, androgens increased AMPK signaling and p-TBC1D4 levels, an effect that was inhibited by STO-609 (Fig. 4C). Surprisingly, androgens also increased the total protein levels of TBC1D4 (relative to GAPDH loading control) in a CaMK2-dependent manner (Fig. 4C), a CaMK2-mediated effect that was also observed in PC-3 cells (Supplementary Fig. 8C). This was of further importance because the expression of TBC1D4 was elevated in samples from prostate cancer patients in multiple clinical cohorts (Table 1) (Welsh et al. 2001, Luo et al. 2002, Vanaja et al. 2003, Varambally et al. 2005, Wallace et al. 2008, Taylor et al. 2010, Grasso et al. 2012). To verify the effects on TBC1D4 expression, we created stable LNCaP derivatives that could inducibly overexpress CAMKK2, and thus, increase AMPK signaling, in the presence of doxycycline (Fig. 4D, E and Supplementary Fig. 9). Here, expression of CAMKK2 was sufficient to increase AMPK activity as previously described (Friso et al. 2011). In addition, CAMKK2 expression increased the T642 phosphorylation of TBC1D4 (relative to total TBC1D4) and increased the total levels of TBC1D4 (relative to GAPDH loading control) (Fig. 4E). However, it should be noted that the CaMKK2-mediated increase in TBC1D4 levels was more moderate than that with androgen treatment (Fig. 4E), suggesting that additional AR-regulated pathways likely also govern TBC1D4 expression.

To confirm AR's role in TBC1D4 expression, we depleted AR using siRNAs to demonstrate that AR was required for both basal and androgen-mediated TBC1D4 expression (Fig. 4F). Additionally, treatment of LNCaP cells with androgens increased TBC1D4 mRNA levels (Supplementary Fig. 9). In support of these data, the AR antagonist enzalutamide decreased androgen-mediated TBC1D4 mRNA and TBC1D4 protein expression (Supplementary Fig. 10). However, overexpression of CAMKK2 alone was not sufficient to increase TBC1D4 mRNA levels (Supplementary Fig. 9), implying that CaMK2's effects on TBC1D4 levels are at the posttranscriptional level. Finally, we determined that AR gene signatures (Fig. 4G) positively correlated with TBC1D4 mRNA levels in multiple clinical cohorts (Taylor et al. 2010, Grasso et al. 2012) including TCGA (Cerami et al. 2012). Taken together, these data indicate that AR and CaMK2 signaling can increase the expression and activity of TBC1D4 through separate mechanisms.

TBC1D4 regulates glucose uptake via GLUT12 trafficking to the plasma membrane

Given TBC1D4's known role in glucose uptake, we next wanted to test its functional role in prostate cancer cells. To test this, we depleted TBC1D4 levels using siRNAs (Fig. 5A) to impair TBC1D4-mediated vesicle trafficking. Knockdown of TBC1D4 blocked androgen-mediated glucose uptake in LNCaP and VCaP cells and basal glucose uptake in PC-3 cells (Fig. 5B). The impaired glucose uptake corresponded with decreased cell growth (Fig. 5C). Interestingly, while androgens also increased TBC1D4 protein levels in non-transformed RWPE-1 cells, knockdown of TBC1D4 did not translate to significant effects on glucose uptake or cell growth (Supplementary Fig. 11), suggesting that TBC1D4-mediated glucose uptake may be more selectively used by cancer cells. We next
assessed the role of TBC1D4 in GLUT12 translocation to the plasma membrane. To do this, we depleted TBC1D4 using siRNA, treated cells ± androgen (AR+ LNCaP and VCaP cells only) and collected protein from whole cell lysates (WCL) or following isolation of the plasma membrane fraction (PM). Knockdown of TBC1D4 did not affect the total levels of GLUT12 (WCL samples), but did decrease GLUT12 in the plasma membrane fraction (PM samples) (Fig. 5D). These results indicated that TBC1D4 was required for maximal GLUT12 PM localization. Consistent with a role for CaMKK2 signaling in this process, treatment with the CaMKK2 inhibitor STO-609 blocked the expression of GLUT12 specifically at the PM (Supplementary Fig. 12). Due to concerns regarding potential off-target effects of STO-609, we decided to complement our pharmacological experiments with a molecular approach. First, we utilized our LNCaP-CAMKK2 stable cells (Fig. 4D and E) to demonstrate that, similar to androgen treatment, overexpression of CAMKK2 alone was sufficient to increase GLUT12 levels in the PM fraction (Supplementary Fig. 13). However, despite multiple attempts, we were unable to obtain a PM fraction from these cells that was completely devoid of GAPDH or CaMKK2. While previous work has indicated (1) AMPK signaling at the PM (Miyamoto et al. 2015 and Supplementary Fig. 13), and therefore, possibly CaMKK2 and (2) GAPDH localization to the plasma membrane (Sirover 2012, Seidler 2013), we could not exclude the possibility that the increased GLUT12 levels we observed in the PM fractions derived from the doxycycline-treated samples were due to incomplete fractionation. Thus, we also decided to assess GLUT12 levels following the knockdown of CAMKK2. To do this, we created stable cells that could overexpress an shRNA targeting CAMKK2, and therefore decrease AMPK signaling, in the presence of doxycycline (Fig. 5E). Initial attempts to create LNCaP-shCAMKK2 cells were unsuccessful (could not achieve significant doxycycline-mediated knockdown) and so we created stable derivatives of the CRPC 22Rv1 cells, which we have found to be more amenable to lentiviral manipulations (D Frigo, unpublished observations). Here, CaMKK2 signaling was decreased following 72h of doxycycline treatment (Fig. 5E). Correspondingly, PM GLUT12 levels were also decreased (Fig. 5F). Interestingly, in this model, silencing CaMKK2 also led to a decrease in overall GLUT12 levels, suggesting that impairing GLUT12 trafficking could destabilize the protein in some cell contexts. In support of this finding, overexpression of CAMKK2 alone increased total GLUT12 protein levels (Supplementary Fig. 13). Surprisingly, while androgens increased GLUT12 trafficking to the PM in VCaP and LNCaP-CAMKK2 cells (Fig. 5D and Supplementary Fig. 13), androgen treatment did not result in an increased level of GLUT12 at the PM in our parental LNCaP cells (Fig. 5D and Supplementary Fig. 12A). This was unexpected because androgens increased total TBC1D4 and p-TBC1D4 levels (Fig. 5D) as well as glucose uptake (Fig. 5B) at this same time point. Taken together, androgen signaling may promote GLUT12 translocation to the PM through a CaMKK2-AMPK-TBC1D4-mediated mechanism but simultaneously cause its turnover at this same location through additional mechanisms that are unknown at this time.

**Discussion**

The data presented here suggest a working model, depicted in Fig. 6, for GLUT12 regulation in prostate cancer. SLC2A12, the gene encoding GLUT12, can directly be targeted by AR. This leads to a pulse of new GLUT12 synthesis. Simultaneously, androgens can directly increase the expression of CAMKK2 (Frigo et al. 2011). CaMKK2 can then later phosphorylate and activate the master metabolic regulator AMPK (Hawley et al. 2005, Hurley et al. 2005, Woods et al. 2005). Interestingly, as shown here in PC-3 cells (Supplementary Fig. 8) and by others in DU145 cells (Fu et al. 2015), CaMKK2-AMPK signaling is maintained in AR-negative prostate cancer cells, indicating that other transcription factors beyond AR promote CaMKK2-AMPK signaling. Regardless, CaMKK2-AMPK signaling will in turn increase GLUT12 translocation to the PM by modulating the activity, and to a lesser extent the expression, of TBC1D4, a known regulator of vesicle trafficking (Kramer et al. 2006, Treebak et al. 2006). Hence, we speculate that the initial pulse of GLUT12 synthesis following androgen treatment (with max levels achieved ~24h after treatment) provides new protein that can later be modified and/or translocated as a result of CaMKK2 signaling at a later time (ex. 72h). This newly modified/translocated GLUT12 would then enable the turnover of older protein. In this regard, androgen signaling could serve to replenish the active levels of GLUT12 at the PM rather than simply increasing total GLUT12 levels throughout the cell. In addition to CaMKK2 signaling, our data (Figs 4 and 5, and Supplementary Figs 9 and 13) suggest that androgens may promote the expression of TBC1D4 through additional, CaMKK2-independent mechanisms (indicated in Fig. 6 as the expression of gene X, unknown at this time, that codes for protein X).
While the regulation of TBC1D4 by AMPK has been previously reported (Kramer et al. 2006, Treebak et al. 2006), to our knowledge, the androgen and CaMKK2-AMPK modulation of TBC1D4 expression has not been previously described. Ultimately, this combined series of transcriptional and posttranscriptional events converge to increase the levels of functional GLUT12 at the PM where it can promote glucose uptake and subsequent cell growth.

GLUT12, as its name implies, was the 12th of 14 known glucose transporters of the SLC2A family to be identified (Rogers et al. 2002). As such, there is relatively
Figure 4
TBC1D4 is regulated by AR and CaMKK2-AMPK signaling. (A) LNCaP cells were co-treated ± 30 μM STO-609 ± 10 nM R1881 (androgen) for 72 h. Relative cell numbers were then measured using a fluorescent DNA stain. *Significant (P < 0.05) changes from double vehicle-treated cells. (B) LNCaP cells were transfected with siRNAs targeting a scramble sequence (siControl) or AMPK and treated ± androgen (R1881) for 72 h. Relative cell numbers were then measured as in A. *Significant (P < 0.05) changes from vehicle treatment. †Significant (P < 0.05) changes from siControl. (C) LNCaP cells were treated for 72 h ±STO-609 ± androgen (R1881) after which whole cell lysates were analyzed via Western blot. (D) The construct used to create the LNCaP-CaMKK2 stable cell line in which CaMKK2 can be overexpressed in the presence of doxycycline. (E) LNCaP-CaMKK2 stable cells were treated for 72 h with increasing concentrations of doxycycline (Dox) or androgen (R1881). Whole-cell lysates were then analyzed via Western blot. (F) LNCaP cells were transfected with mock control or siRNAs targeting a scramble sequence (siControl) or AR (siAR) and treated ± androgen (R1881) for 72 h. Whole cell lysates were then analyzed via Western blot. (G) Expression of TBC1D4 correlated significantly with a previously described AR gene signature (Hieronymus et al. 2006) in transcriptomic profiles of prostate cancer patients from the Taylor et al. (2010) prostate cancer clinical cohort. Similar results were obtained using this AR activity signature as well as an additional AR activity signature (Nelson et al. 2002) across multiple clinical cohorts (Taylor et al. 2010, Grasso et al. 2012) including TCGA (Cerami et al. 2012).
Figure 5
TBC1D4 regulates glucose uptake by modulating GLUT12 trafficking to the plasma membrane. (A) Indicated prostate cancer cells were transfected with mock or siRNAs targeting scramble control (siControl) or TBC1D4 (siTBC1D4). LNCaP or VCaP cells were also treated for 72 h ± androgen (10 nM R1881). Whole cell lysates were extracted and analyzed by Western blot. (B) Indicated prostate cancer cells were transfected and/or treated as in A. Glucose uptake was measured using a bioanalyzer, and data were normalized to cellular DNA content that was assessed using a fluorescent DNA stain. *Significant (P < 0.05) changes from vehicle-treated (LNCaP and VCaP) or siControl (PC-3) cells. (C) Indicated prostate cancer cells were transfected and/or treated as in B. Relative cell numbers were then measured using the fluorescent DNA stain. *Significant (P < 0.05) changes from vehicle-treated (LNCaP and VCaP) or siControl (PC-3) cells. (D) Indicated prostate cancer cells were transfected and/or treated as in A. Afterward, whole cell lysates (WCLs) were extracted and analyzed by Western blot. Duplicate cells were also subjected to a plasma membrane (PM) fractionation where the PM fraction was extracted and analyzed by Western blot. ATPase serves as a positive control for the enriched PM fraction. GAPDH serves as a loading control for the WCL. (E) Top, diagram of the construct used to create the 22Rv1-shCAMKK2 stable cell line in which shRNA targeting CAMKK2 can be overexpressed in the presence of doxycycline. Bottom, Western blot validation of the 22Rv1-shCAMKK2 stable cell line. 22Rv1-shCAMKK2 stable cells were treated for 72 h ± 800 ng/mL doxycycline (Dox). Whole cell lysates were then analyzed via Western blot. (F) 22Rv1-shCAMKK2 cells were treated the same as in E and then subjected to fractionation and Western blot analysis as described in D.
little known about its regulation and function compared to many other GLUTs such as GLUT1 and GLUT4. Interestingly, GLUT12 was first discovered in MCF-7 breast cancer cells (Rogers et al. 2002). Since then, it has been found to be expressed in a number of malignancies including rhabdomyosarcomas, oligodendrogliomas, oligoastrocytomas, astrocytomas as well as breast, lung, colorectal and prostate cancers (Rogers et al. 2002, 2003, Chandler et al. 2003, Stockhammer et al. 2008, Pujol-Gimenez et al. 2015). Hence, a potential oncogenic role for this transporter has been emerging. In agreement with the mRNA expression data listed in Table 1, immunohistochemical staining for GLUT12 revealed the transporter’s protein expression in malignant but not benign prostatic hyperplasia tissue (Chandler et al. 2003). In estrogen receptor-positive (ER+) MCF-7 breast cancer cells, 24-h treatment with the steroid hormone estrogen was shown to increase GLUT12 protein but not mRNA levels, suggesting estrogen increased GLUT12 levels by promoting translation or protein stability (Macheda et al. 2005). They also demonstrated that GLUT12 protein could not be detected in ER-negative breast cancer cells, further supporting a model of ER regulation. The authors additionally stated (but did not show the data) that 24-h DHT treatment increased GLUT12 protein levels in MCF-7 cells, which express AR (Macheda et al. 2005). It was not reported whether DHT increased mRNA expression. Like ER signaling in breast cancer cells, our data demonstrate that AR signaling increased GLUT12 expression in prostate cancer (Figs 1, 2 and Supplementary Figs 1, 4, 5 and 6). However, our findings indicate that, unlike ER signaling in breast cancer, SLC2A12 is a direct AR target in prostate cancer cells (Fig. 2 and Supplementary Figs 5 and 6). Further, AR-negative PC-3 prostate cancer cells express GLUT12, indicating that AR is not the only transcription factor that regulates SLC2A12 (Figs 3 and 5 and Supplementary Figs 2 and 12).

GLUT transporters are grouped into three classes based on their sequence homology and structure (Joost & Thorens 2001, Barron et al. 2016). GLUT12 belongs to class III while many of the originally identified GLUTs including the insulin-regulated GLUT4 belong to class I. Despite belonging to two different classes, GLUT12 was originally identified by homology to GLUT4 (Rogers et al. 2002). Like GLUT4, evidence suggests that GLUT12’s subcellular location is also regulated by signals such as insulin (Rogers et al. 2002, Stuart et al. 2009). In this regard, it is of note that GLUT12 possesses the motifs found in GLUT4 that facilitate this transporter’s subcellular trafficking (Rogers et al. 2002). To that end,
it has been speculated that GLUT4 and GLUT12 might even be colocalized to the same vesicles under baseline conditions. Our data here suggest that like GLUT4 (Sano et al. 2003, Miinea et al. 2005), GLUT12 translocation is also regulated by TBC1D4 (Fig. 5).

Under physiological conditions in muscle or fat, TBC1D4 can be phosphorylated by Akt or AMPK (Miinea et al. 2005, Sakamoto & Holman 2008, Stuart et al. 2009, Cartee 2015). This phosphorylation inhibits its Rab GTPase-activating activity and promotes GLUT4 and, as our data suggest here, GLUT12 translocation. Consistent with this, inhibition of CaMKK2-AMPK kinase signaling by STO-609 or shRNA-mediated silencing of CaMKK2 decreased GLUT12 PM levels (Fig. 5 and Supplementary Fig. 12) and glucose uptake (Mianne et al. 2011). The existence of this regulatory mechanism for GLUT12 is consistent with the presence of the conserved motif, described earlier, in both transporters that are known to be responsible for insulin-mediated GLUT4 translocation. As this phosphorylation is an inhibitory event, it is not unexpected that knockdown of TBC1D4 or its paralog TBC1D1 have been described before to increase cell surface GLUT4 levels and glucose uptake in muscle and fat cells (Eguez et al. 2005, Sano et al. 2007, Chavez et al. 2008).

Paradoxically, silencing of TBC1D4 in prostate cancer cells decreased the plasma membrane-localized levels of GLUT12 while having no effect on total GLUT12 levels (Fig. 5D). These findings suggest that TBC1D4 might be required to establish basal GLUT12 vesicle trafficking that can then be further regulated in response to cues such as CaMKK2 signaling. Future experiments will be critical to determine whether this regulation is specific to GLUT12 and/or prostate cancer.

At this time, we cannot exclude the possibility that androgens or CaMKK2-AMPK signaling could promote glucose uptake through additional, GLUT12-independent mechanisms. As described earlier, TBC1D4 was first demonstrated to regulate the translocation of GLUT4 (Kramer et al. 2006, Treebak et al. 2006), a transporter that is also expressed (albeit lower) in the prostate (Fig. 1) (Gonzalez-Menendez et al. 2014). In addition, AMPK has been reported to increase GLUT1 levels through a variety of mechanisms (Barnes et al. 2002, Wu et al. 2013). Further, AMPK can directly phosphorylate two of the four isoenzymes of 6-phosphofructo-2-kinase/fructose-2, 6-bisphosphate 2-phosphatase (PFKFB), an enzyme that represents the rate-limiting step of glycolysis (Marsin et al. 2000, Almeida et al. 2004, Domenech et al. 2015). These phosphorylation events on PFKFB2 and PFKFB3 increase their kinase activity and, therefore, promote forward flux through glycolysis. Because glucose uptake through the facilitative transporters is regulated by the concentration gradient, increased cellular catabolism of glucose would stimulate the uptake of more sugar by other GLUTs that might also function independently of TBC1D4. Likewise, androgens and CaMKK2-AMPK signaling have also been demonstrated to increase glucose metabolism through the tricarboxylic acid (TCA) cycle, the hexosamine biosynthetic pathway and the pentose phosphate pathway (Massie et al. 2011, Itkonen et al. 2013, Tennakoon et al. 2014, Tsouko et al. 2014).

Collectively, these effects suggest that GLUT12 represents only one of potentially several mechanisms downstream of androgen and CaMKK2 signaling that could increase glucose uptake. Given prostate cancer’s heterogeneous makeup (Cancer Genome Atlas Research 2015), it is likely that some subtypes may favor GLUT12- and/or TBC1D4-independent glucose uptake.

This study examined the regulation and role of the transporter GLUT12 in glucose uptake. It still remains to be determined how glucose is then used by the cancer cells. As described earlier, androgen and CaMKK2 signaling have previously been demonstrated to promote glucose metabolism through the hexosamine biosynthetic pathway, the pentose phosphate pathway, glycolysis and the TCA cycle in prostate cancer (Massie et al. 2011, Itkonen et al. 2013, Tennakoon et al. 2014, Tsouko et al. 2014). In addition, androgens are known to stimulate the formation of intracellular fat depots, thought to promote growth, in prostate cancer cells (Swinnen et al. 2006, Shi et al. 2013). These fat reservoirs are likely created in part through the breakdown of sugars and shuttling of their carbons into de novo lipogenesis. Future studies using isotopic tracing techniques will undoubtedly help determine the exact contribution of each one of these pathways to the growing tumor.

### Supplementary data
This is linked to the online version of the paper at [https://doi.org/10.1530/ERC-17-0051](https://doi.org/10.1530/ERC-17-0051).

### Declaration of interest
The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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