

The role of microRNA deregulation in the pathogenesis of adrenocortical carcinoma

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Abstract

Adrenocortical carcinoma (ACC) is an aggressive tumor showing frequent metastatic spread and poor survival. Although recent genome-wide studies of ACC have contributed to our understanding of the disease, major challenges remain for both diagnostic and prognostic assessments. The aim of this study was to identify specific microRNAs (miRNAs) associated with malignancy and survival of ACC patients. miRNA expression profiles were determined in a series of ACC, adenoma, and normal cortices using microarray. A subset of miRNAs showed distinct expression patterns in the ACC compared with adrenal cortices and adenomas. Among others, *miR-483-3p*, *miR-483-5p*, *miR-210*, and *miR-21* were found overexpressed, while *miR-195*, *miR-497*, and *miR-1974* were underexpressed in ACC. Inhibition of *miR-483-3p* or *miR-483-5p* and overexpression of *miR-195* or *miR-497* reduced cell proliferation in human NCI-H295R ACC cells. In addition, downregulation of *miR-483-3p*, but not *miR-483-5p*, and increased expression of *miR-195* or *miR-497* led to significant induction of cell death. Protein expression of p53 upregulated modulator of apoptosis (PUMA), a potential target of *miR-483-3p*, was significantly decreased in ACC, and inversely correlated with *miR-483-3p* expression. In addition, high expression of *miR-503*, *miR-1202*, and *miR-1275* were found significantly associated with shorter overall survival among patients with ACC (*P* values: 0.006, 0.005, and 0.042 respectively). In summary, we identified additional miRNAs associated with ACC, elucidated the functional role of four miRNAs in the pathogenesis of ACC cells, demonstrated the potential involvement of the pro-apoptotic factor PUMA (a *miR-483-3p* target) in adrenocortical tumors, and found novel miRNAs associated with survival in ACC.

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Introduction

Patients with tumors of the adrenal cortex may present an incidentally detected or hormonally overproducing adrenocortical adenoma, or uncommonly an adrenocortical carcinoma (ACC). ACC is an aggressive tumor associated with poor prognosis in adults (Bertherat *et al.* 2006). Differential diagnosis between ACC and the more common adenoma is difficult, particularly for the tumors in the marginal category, i.e. Weiss score 2 or 3 vs 4. Moreover, prognostic tools for early recognition of

metastatic ACC disease and adverse outcome are presently lacking in routine diagnostics. Profiling of mRNA and microRNA (miRNA) expressions in human cancer have revealed important alterations that do not only have possible diagnostic and prognostic value but also point toward molecular alterations of mechanistic importance for tumor etiology and progression.

In adrenocortical tumors, mRNA expression profilings by several independent groups have revealed significant and reproducible alterations for the

discrimination of adenoma and ACC and for the determination of ACC prognosis (Giordano *et al.* 2003, de Fraipont *et al.* 2005, Velazquez-Fernandez *et al.* 2005, Giordano *et al.* 2009, Laurell *et al.* 2009, de Reynies *et al.* 2009, Soon *et al.* 2009a). One of the most striking finding was the significant overexpression of insulin-like growth factor 2 (*IGF2*) in ACC, which was observed in >80% of cases compared with adrenal cortices and hormonally active/inactive adenomas (Giordano *et al.* 2009, Laurell *et al.* 2009, de Reynies *et al.* 2009). In addition, the expression levels of other genes such as *DLG1* and *PINK1* have been associated with the clinical outcome of ACC (de Reynies *et al.* 2009).

miRNAs are ~22 nucleotide long single-stranded non-coding RNAs generated by the RNase-III enzyme Dicer from endogenous hairpin-shaped transcripts (Iorio & Croce 2009). miRNAs have diverse roles in many biological processes and have also been shown important for tumor development, by acting as oncogenic or tumor suppressive species. Besides, accumulating evidence also supports a role of miRNAs as diagnostic and prognostic biomarkers of human cancers (Iorio & Croce 2009, Ferracin *et al.* 2010). Profiling of adrenocortical tumors have identified several deregulated miRNAs in benign adrenocortical diseases (Iliopoulos *et al.* 2009, Bimpaki *et al.* 2010), as well as ACC (Soon *et al.* 2009b, Tombol *et al.* 2009, Doghman *et al.* 2010, Patterson *et al.* 2011). Several deregulated miRNAs have been reported in ACC, including overexpression of *miR-483-3p* and *miR-483-5p*, which are transcribed from an intronic sequence of the *IGF2* gene (Soon *et al.* 2009b, Doghman *et al.* 2010). *miR-483-3p* has recently been shown to target the pro-apoptotic gene BCL-2-binding component 3/p53 upregulated modulator of apoptosis (*BBC3/PUMA*) in cancers of the colon, liver, and breast (Veronese *et al.* 2010), suggesting its anti-apoptotic role in a variety of tumor types.

In this study, we characterized miRNA expression patterns of ACC compared with adenomas and normal adrenal cortices, and related the findings to outcome at follow-up. The functional consequences of *miR-483-3p*, *miR-483-5p*, *miR-195*, and *miR-497* deregulations were studied *in vitro* concerning cell proliferation and apoptosis. We further explored the involvement of PUMA, a potential target of *miR-483-3p*, in clinical samples. Our findings may contribute to further understanding of ACC development, and suggest a role for selected miRNAs as diagnostic and prognostic biomarkers in ACC.

Materials and methods

Tumors and normal tissues

A total of 68 snap-frozen primary sporadic adrenocortical tumors collected at the Karolinska University Hospital were included in this study. In addition, histopathologically verified normal adrenal cortical samples were obtained from ten patients undergoing nephrectomy for other reasons and used as non-neoplastic reference tissues. All samples were obtained with informed consent and the study of the tissue materials was approved by the Local Ethics Committee. Tumors were diagnosed following the WHO classification (DeLellis *et al.* 2004) as adrenocortical adenoma (Ad 1–43) or ACC (Ca 1–25). Clinical details have been partly published for subsets of the cases in previous studies (Laurell *et al.* 2009). All 25 carcinoma cases were followed-up until May 2011 or until their death, and the follow-up data is detailed together with clinical and histopathological information in [Supplementary Table 1](#), see section on [supplementary data](#) given at the end of this article. Thirteen of the 43 adenoma cases were cortisol-producing tumors (Cushing) from one male and 12 female patients with a mean age of diagnosis at 56 years (range 27–81) and a mean tumor size of 3.8 cm (range 1.5–6.5). Sixteen adenomas were aldosteronomas (five male and 11 female patients) with a mean age of 46 years (range 16–79) and a mean tumor size of 2.0 cm (range 0.9–4.7). The last 14 adenomas were classified as incidentalomas (six male and eight female patients) with a mean age of 59 years (range 42–65) and a mean size of 3.9 cm (range 2.5–5.3). The median follow-up time for adenomas was 39 months (range 12–258). At the end of the follow-up, no adenoma patient had disease progression or metastasis.

Cell line

The ACC cell line NCI-H295R was purchased from the American Type Culture Collection (ATCC# CRL-2128; LGC Standards, Middlesex, UK). Cells were maintained in DMEM:F12 medium containing 2.5% of NuSerum (cat. no. 355500; BD Biosciences, Bedford, MA, USA), 1% penicillin/streptomycin and 1% insulin-transferin-sodium selenite (ITS + 1) liquid media supplement (cat. no. I2521; Sigma-Aldrich Logistik GmbH) at 37 °C and 5% CO₂. Authentication of the cell line was evaluated and verified by Bio-Synthesis, Inc. (Lewisville, TX, USA) employing genotyping of 15 short tandem repeat (STR) loci and the amelogenin gene (*AMEL*), and comparison with genotype information at the ATCC ([Supplementary Table 2](#), see section on [supplementary data](#) given at the end of this article).

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RNA extraction

Total RNA isolation was performed by mirVana miRNA Isolation Kit (Applied Biosystems/Ambion, Austin, TX, USA). Measurement of RNA concentrations was done using a NanoDrop ND-1000 spectrophotometer (NanoDrop Technologies, Wilmington, DE, USA).

Microarray-based miRNA profiling

Global miRNA expression profiling of adrenocortical tumors and normal cortices was performed by the Human Agilent's miRNA microarray system (Agilent, Santa Clara, CA, USA) with probes matching 903 human miRNAs (miRBase release 14). Array hybridizations and data analyses were performed essentially as described previously (Caramuta *et al.* 2010). In brief, samples of 120 ng total RNA were labeled with Cyanine 3-pCp, hybridized onto arrays for 18–20 h at 55 °C, followed by washing. Slides were scanned in an Agilent microarray scanner G2565BA and images were processed with Feature Extraction Software v10.7.3.1 (Agilent). Cluster 3.0 Software (<http://bonsai.hgc.jp/~mdehoon/software/cluster/>) was used for normalization and median centering (de Hoon *et al.* 2004). Normalized miRNAs with <50% missing values were included in subsequent analyses for hierarchical clustering and significance analysis of microarrays (SAM). The data from microarray analysis have been deposited at NCBI Gene Expression Omnibus (GEO accession number, GSE22816).

qRT-PCR analysis of individual miRNAs

Selected mature miRNAs were quantified using commercially available TaqMan qRT-PCR assays (Applied Biosystems) and a 7900HT Real-Time PCR System (Applied Biosystems). cDNA was synthesized from 25 ng total RNA using TaqMan mRNA RT Kit (Applied Biosystems) and used for quantification of *miR-483-3p* (ID 002339), *miR-483-5p* (ID 002338), *miR-497* (ID 001043), *miR-195* (ID 000494), *miR-1974* (ID 121209_mat), *miR-210* (ID 000512), *miR-21* (ID 000397), *miR-503* (ID 001048), *miR-1202* (ID 002858), *miR-1275* (ID 002840), *miR-638* (ID 001582), *miR-1915* (ID 121111_mat), and *miR-572* (ID 001614) with normalization against *RNU6B* (ID 001093). All reactions were performed in triplicate, and relative expression levels were determined with the $\Delta\Delta C_T$ method and reported as $2^{-\Delta C_T}$.

miRNA inhibition and overexpression in NCI-H295R cells

NCI-H295R ACC cells were transfected using Nucleofector Technology (Amaxa Biosystems, Gaithersburg, MD, USA). In brief, 3×10^6 cells were resuspended in 100 μ l Nucleofector solution R, and mixed with 100 pmol of miRNA inhibitors (anti-miR-483-3p or anti-miR-483-5p; Applied Biosystems/Ambion) or miRNA precursors (pre-miR-195 or pre-miR-497; Applied Biosystems/Ambion). Anti-miR-negative control #1 or pre-miR-negative control #1 (Applied Biosystems/Ambion) containing a non-targeting sequence molecule was used as a negative control. Cells were then electroporated using the program T-20, allowed to recover in DMEM:F12 media for 15 min at 37 °C and seeded in T-25 flasks with 5 ml DMEM:F12 media. Culture media was replaced after 24 h and cells were cultured for additional 72 h. All transfection experiments were repeated at least three times and used for proliferation and apoptosis assays. Transfection efficiency was determined by measuring the endogenous *miR-483-3p*, *miR-483-5p*, *miR-195*, or *miR-497* expression levels by qRT-PCR (Supplementary Figure 1, see section on supplementary data given at the end of this article). To demonstrate that the transfected miRNAs achieved physiologically relevant expression levels, we compared the expression fold change differences between the experimental cell culture systems (anti-miRNA/pre-miR versus control) and the clinical materials (carcinomas versus normal cortices) for *miR-483-3p*, *miR-483-5p*, *miR-195*, and *miR-497*. In the transfected cells expressing specific miRNA assayed, we observed a reduction of ~12-fold for *miR-483-3p* and approximately ninefold for *miR-483-5p* while we noticed ~15- and 30-fold increase for *miR-195* and *miR-497*, respectively, compared with the negative control. The effect of transfections had a similar magnitude of miRNA expression levels as for the comparison between carcinomas and normal adrenal cortices. In carcinomas, we observed an average increased expression of 166-fold (range 0.1–1330) and 73-fold (range 0.1–400) for *miR-483-3p* and *miR-483-5p*, respectively; but a reduced expression of sixfold (range 0.6–26) and 18-fold (range 0.3–170) for *miR-195* and *miR-497* respectively.

Cell proliferation WST-1 colorimetric assay

WST-1 (4-(3-(4-iodophenyl)-2-(4-nitrophenyl)-2H-5-tetrazolio)-1,3-benzene disulfonate; cat. no. 11644807001; Roche Applied Science) colorimetric assay was carried out to determine the effects of

miR-483-3p and *miR-483-5p* inhibition or *miR-195* and *miR-497* overexpression on cell proliferation. The assay was performed by 96-well plates, with seeding of ~20 000 cells in each well. After 72 h of transfection, 20 µl of the WST-1 solution were added to the culture medium and incubated for 2.5 h at 37 °C. Absorbance was subsequently determined using a microplate ELISA reader (VERSAmax; Molecular Devices, Sunnyvale, CA, USA) and analyzed with SoftMax Pro 5 Software (Molecular Devices) applying the wavelengths 450 nm for measurements and 650 nm as reference. All experiments were conducted in eight wells for each condition, and replicated at least three times. Cell proliferation was calculated by comparing the absorbance values of the samples after background subtraction. The fraction of surviving cells was calculated by defining the anti-miR- (or pre-miR)-negative control #1-treated cells as one.

Apoptosis caspase-3 colorimetric assay

The caspase-3 colorimetric assay (cat. no. L00289; Genscript, Piscataway, NJ, USA) was used to evaluate the effects of *miR-483-3p* and *miR-483-5p* inhibition or *miR-195* and *miR-497* overexpression on apoptosis; 3×10^6 cells were transfected and seeded in T-25 flasks. After 72 h of transfection, proteins were isolated and quantified by Bio-Rad Protein Assay (Bio-Rad). Fifty microliters of lysate containing 50 µg protein was mixed with 50 µl of 2× reaction buffer and 5 µl of caspase-3 substrate and incubated for 4 h at 37 °C. Absorbance was subsequently determined using a microplate ELISA reader (VERSAmax; Molecular Devices) and analyzed with SoftMax Pro 5 Software (Molecular Devices) applying the wavelengths 405 nm for measurement. Apoptosis was calculated by comparing the absorbance values of the anti-miR/pre-miR treated cells with the respective negative control-treated cells. All the experiments were replicated three times.

Western blot analysis

Whole cell lysates were prepared from normal and tumor tissue samples by homogenization in NP-40 lysis buffer (cat. no. FNN0021; Invitrogen), with addition of protease inhibitor (complete protease inhibitor cocktail; Roche Diagnostics Corporation) and 1 mM of phenylmethanesulfonyl fluoride (Sigma–Aldrich). After quantification by Bio-Rad Protein Assay (Bio-Rad), 60 µg of lysate was separated in Novex 10% Tricine gels (Invitrogen) and transferred to nitrocellulose membranes (LC2001; Invitrogen). Filters were blocked with 5% non-fat milk diluted in TBS/0.05% Tween 20, and incubated with rabbit polyclonal anti-PUMA

antibody (#4976, Cell Signaling Technology, Danvers, MA, USA) at 1:1000 dilution, followed by an anti-rabbit IgG–HRP (1:3000; #170-6515; Bio-Rad Laboratories) used as secondary antibody. Detection was carried out with Novex ECL HRP chemiluminescent substrate reagent (#WP20005; Invitrogen). Novex Sharp Pre-stained protein standards (#57318; Invitrogen) and MagicMark XP (#LC5602; Invitrogen) markers were used to determine relative molecular weights. Protein levels were quantified on X-ray films from immunoblots using ImageJ Software (<http://rsb.info.nih.gov/ij/>). Subsequent incubation of the filters with an anti-GAPDH antibody (sc-47724; Santa Cruz Biotechnology, Inc, Santa Cruz, CA, USA) diluted at 1:5000 was performed for normalization purposes.

Statistical analysis

All analyses were performed by Statistica 8.0 (StatSoft, Inc., Tulsa, OK, USA) or MS Office Excel 2007, unless otherwise specified. Unpaired Student's *t*-test was conducted to compare miRNA expressions in different groups, while paired Student's *t*-test was performed to analyze transfection experiments. Correlations between *miR-195* and *miR-497*, and *miR-483-3p* and PUMA expression levels were assessed by Pearson's correlation analyses and *P* values were estimated by permuting the samples. SAM survival analysis was used to identify miRNAs associated with survival. *P* values were obtained for the Cox score statistics using the χ^2 -distribution. For survival analysis, all patients who died of unknown causes or causes unrelated to ACC (i.e. Ca 3, Ca 7, Ca 12, and Ca 14) during follow-up were considered as 'censored'. Selected miRNAs potentially associated with survival were analyzed by TaqMan qRT-PCR and carcinoma patients were classified into different groups with high or low expression of each miRNA according to median level. The inter-relationship of miRNAs with survival was studied using Kaplan–Meier plots, and the curves for each group were compared by log-rank test. All the analyses were two-tailed and *P* values <0.05 were considered significant.

Results

Deregulated miRNAs in ACC

In this study, we characterized miRNA expression patterns in a cohort of 48 adrenocortical tumors (26 adenomas and 22 carcinomas) and four normal adrenal cortices using a microarray-based approach. Unsupervised clustering analysis classified the samples in different subgroups based on the similarity of their miRNA expression profiles. In Fig. 1, clustering based

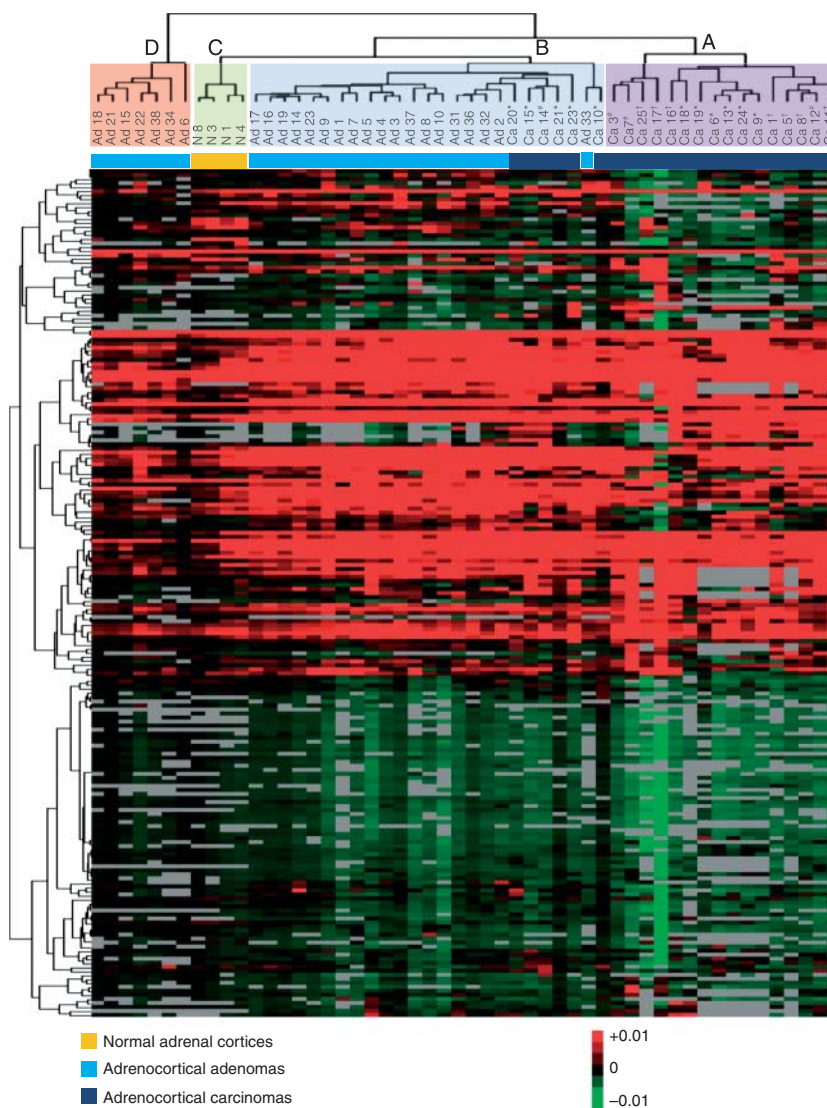


Figure 1 Unsupervised clustering analysis of microarray-based miRNA expression data for four normal adrenal cortices (N), 26 adenomas (Ad), and 22 carcinomas (Ca). Samples were clustered using the Spearman rank correlation and complete linkage. The four main clusters obtained (A–D) are indicated above the heatmap. Red and green colors indicate relatively high and low expression respectively. Missing values are indicated in gray. *Alive; †dead of disease; #dead of unrelated or unknown causes.

on 213 miRNAs revealed four main clusters: cluster A consisted of the majority of carcinoma samples (16 of the 22 ACC; 72%) and cluster B comprised 19 adenomas and the remaining six carcinomas. Five of the six ACC patients in cluster B are still alive at the end of follow-up and one died of unrelated causes with ACC after 65 months. All four normal adrenal cortices were grouped in cluster C, and the remaining seven adenomas were included in cluster D.

We also performed SAM analysis to identify the most significant deregulated miRNAs that could distinguish carcinomas from adenomas and adrenal cortices. The analysis identified 72 differentially

expressed miRNAs with a false discovery rate (FDR) of 0% (Supplementary Table 3, see section on supplementary data given at the end of this article). Several of these miRNAs were also found to have significantly differential expression in the comparisons between carcinomas and adenomas or normal adrenal cortices (Supplementary Tables 4 and 5, see section on supplementary data given at the end of this article). To verify the significance of the results obtained by microarray analysis, we evaluated the expression levels of seven miRNAs (*miR-483-3p*, *miR-483-5p*, *miR-210*, *miR-21*, *miR-1974*, *miR-195*, and *miR-497*) by qRT-PCR in a series of 68 adrenocortical tumors

(43 adenomas and 25 carcinomas) and ten normal adrenal cortices (Fig. 2). These miRNAs were selected among those with the highest score in the SAM analysis, or because of their involvement in adrenal cortical tumors or other tumor types. In concordance with the microarray results, the qRT-PCR analyses revealed significantly higher expression of *miR-483-3p*, *miR-483-5p*, *miR-210*, and *miR-21*, as well as lower expression of *miR-1974*, *miR-195*, and *miR-497* in carcinomas compared with adrenal cortices or adenomas (Fig. 2 and Supplementary Figure 2, see section on supplementary data given at the end of this article). Interestingly, two of these miRNAs, *miR-483-3p* and *miR-483-5p*, were also significantly differential expressed between the carcinomas in cluster B and cluster A observed in unsupervised clustering (Supplementary Table 6, see section on supplementary data given at the end of this article). The precursor of *miR-497* is located at a distance of ~200 bases upstream of *miR-195* at chromosomal region 17p13.1, suggesting that *miR-497* and *miR-195* belong to the same miRNA cluster and are likely to be co-expressed. We found that the expression of *miR-195* was strongly correlated with the *miR-497* expression (corr=0.9, $P < 0.00001$; Supplementary Figure 3, see section on supplementary data given at the end of this article).

Effect of altered *miR-483*, *miR-195*, and *miR-497* expression on cell proliferation and apoptosis in NCI-H295R cells

We further evaluated the functional consequences from dysregulation of four miRNAs in human NCI-H295R ACC cells. These miRNAs were chosen because of their associations with malignancy in adrenal cortical tumors. In Fig. 3A, inhibition of *miR-483-3p* or *miR-483-5p* expression resulted in significant reduction (20 and 30% respectively), of cell proliferation compared with negative control cells (treated with anti-miR-negative control #1). In addition, cells transfected with anti-miR-483-3p, but not with anti-miR-483-5p, showed a significant increase (approximately twofold) in apoptosis compared with cells transfected with negative control (Fig. 3A).

We also explored the functional consequences of *miR-497* and *miR-195* overexpression, which were found significantly downregulated in ACC compared with adenomas. In line with the effects observed for *miR-483-3p*, overexpression of *miR-195* or *miR-497* led to a significant decrease in cell growth (30 and 40% respectively), and a concomitant induction of cell death (40 and 30% respectively), compared with negative control (Fig. 3B).

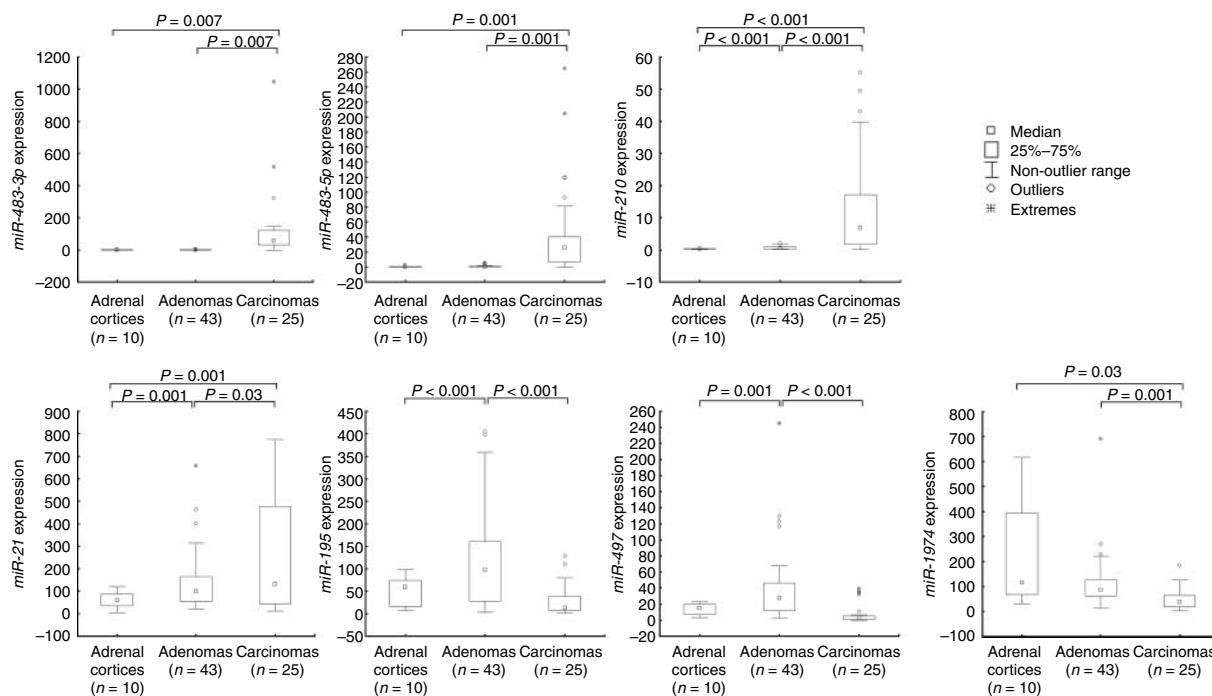


Figure 2 Relative expression levels of *miR-483-3p*, *miR-483-5p*, *miR-210*, *miR-21*, *miR-195*, *miR-497*, and *miR-1974* in the different sample groups. Box plots show miRNA expression levels determined by qRT-PCR in adrenocortical carcinomas, adenomas, and adrenal cortices. Statistical significances between the groups were determined with two-tailed unpaired *t*-test and $P < 0.05$ were considered significant.

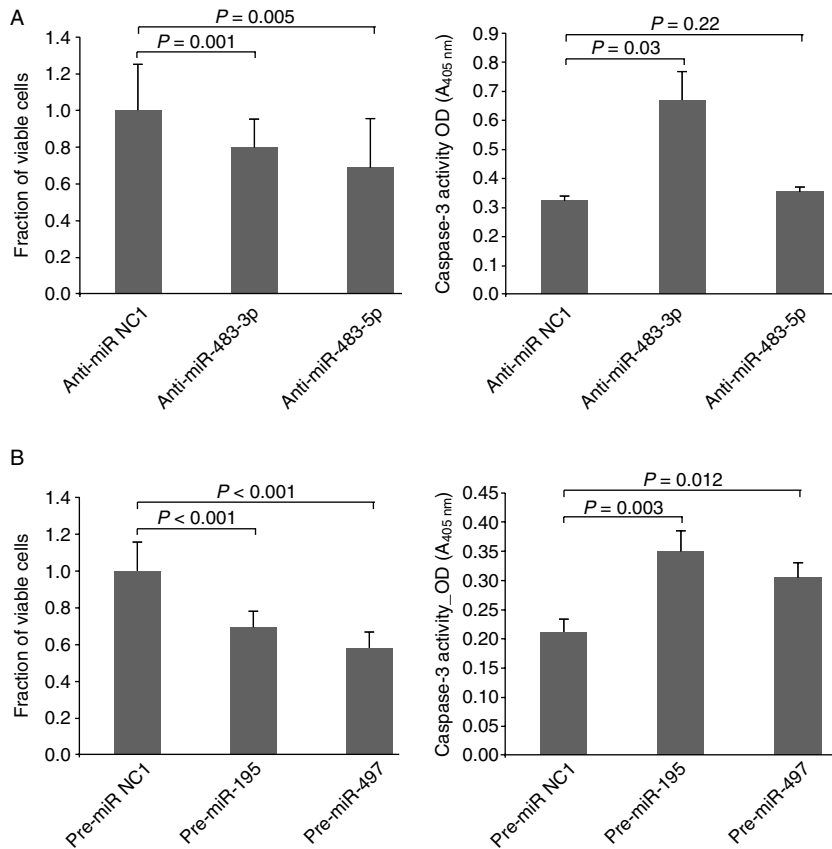


Figure 3 Effects of *miR-483* inhibition and *miR-497* or *miR-195* overexpression on cell proliferation and apoptosis in NCI-H295R ACC cells. Cell viability was evaluated by WST-1 colorimetric assay and cell apoptosis by caspase-3 colorimetric assay upon (A) inhibition of *miR-483-3p* and *miR-483-5p* or (B) overexpression of *miR-195* and *miR-497*. Error bars represent s.d. of the mean from three independent experiments. Statistical significance between the two treatment conditions was evaluated by Student's paired *t*-test and $P < 0.05$ was considered significant. NC1, negative control #1.

Relationship between *miR-483-3p* and PUMA expression in adrenocortical tumors

Recent findings of the pro-apoptotic gene *BBC3/PUMA* as a direct target of *miR-483-3p* (Veronese *et al.* 2010), in combination with our observations of the pro-apoptotic and anti-proliferative effects of *miR-483-3p* silencing, prompted us to investigate a possible connection between PUMA and *miR-483-3p* expression in adrenocortical tumors. We measured the PUMA protein expression levels by western blot analysis in three normal adrenal cortical samples, 26 adenomas, and 22 carcinomas, and compared with the expression of *miR-483-3p*. In Fig. 4, high levels of PUMA expression were detected in almost all adenomas and adrenal cortices, but only in a small proportion of carcinomas. The PUMA protein expression was inversely correlated with the *miR-483-3p* expression (corr = -0.31 , $P = 0.025$). Notably, those carcinomas expressing low levels of

miR-483-3p had a higher PUMA protein expression (e.g. Ca 20), and *vice versa* (e.g. Ca 6; Fig. 4).

miRNAs associated with survival of ACC patients

In an attempt to identify specific miRNAs associated with survival, we performed SAM survival analysis on the carcinoma cases. The analysis identified 11 miRNAs that could predict the disease outcome with a SAM survival score > 2.6 (FDR: 0%; Fig. 5A). Specifically high expression of *miR-638*, *miR-1246*, *miR-1915*, *miR-1275*, *miR-503*, *miR-671-5p*, *miR-1268*, *miR-762*, *miR-331-3p*, *miR-1202*, and *miR-572* were found associated with short survival (Fig. 5A). Although two of the carcinoma cases (Ca 23 and Ca 24) had a shorter follow-up (25 and 20 months respectively), exclusion of these cases from the survival analyses did not affect the significance of our results. Using the miRNA set selected by SAM survival analysis, we performed a hierarchical

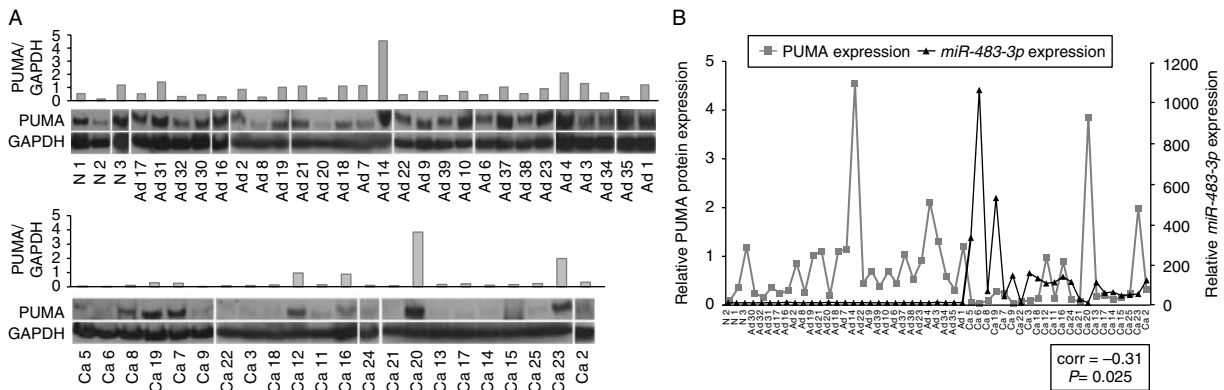


Figure 4 Correlation between expression of *miR-483-3p* and its putative target PUMA. (A) Western blot analyses of PUMA expression and GAPDH used as reference in three adrenal cortical samples (N), 26 adenomas (Ad), and 22 carcinomas (Ca). The expression levels of PUMA shown in the diagram above were calculated by normalization against GAPDH protein levels. (B) Comparison between *miR-483-3p* expression and PUMA protein levels in individual samples. Correlation was assessed by Pearson correlation analysis.

clustering analysis and observed that the samples were classified into three clusters. As shown in Fig. 5A, the survival analysis revealed a significant difference between cluster 3 and the other two clusters ($P=0.03$, log-rank test). Furthermore, using Kaplan–Meier survival and log-rank analyses, we evaluated the association with survival for each individual miRNA detected by SAM. The analysis revealed six of these 11 miRNAs as significantly associated with survival (Supplementary Figure 4, see section on supplementary data given at the end of this article). High expression of *miR-503* ($P<0.001$; log-rank test), *miR-1202* ($P=0.009$), *miR-1915* ($P=0.013$), *miR-638* ($P=0.018$), *miR-572* ($P=0.024$), and *miR-1275* ($P=0.035$) were significantly associated with shorter overall survival (Supplementary Figure 4).

To validate the significance of these prognostic miRNAs, we evaluated the expression of six miRNAs in the cohort of 25 ACC by qRT-PCR. Using Kaplan–Meier survival curves and log-rank analysis, we confirmed the significant association of *miR-503* ($P=0.006$), *miR-1202* ($P=0.005$), and *miR-1275* ($P=0.042$) with overall survival (Fig. 5B), which is in concordance with the array-based results. However, *miR-572*, *miR-1915*, and *miR-638* were not found statistically significant (data not shown).

Discussion

In this study, we identified a set of miRNAs that is differentially expressed between malignant and non-malignant samples, and a small number of miRNAs associated with survival in ACC patients.

miRNA deregulation in ACC

Using a microarray approach, we identified distinct miRNA expression patterns between carcinoma and non-carcinoma samples. The consistency of the findings was supported by the validation of seven of the most significant differentially expressed miRNAs in an extended series of adrenocortical tumors by TaqMan qRT-PCR methodology. Taken together, the observations indicate the significant importance of these miRNAs in the pathogenesis of ACC.

We showed that *miR-497* expression is significantly reduced in ACC compared with adenomas. In concordance with our results, *miR-497* was reported underexpressed in childhood adrenocortical tumors (Doghman et al. 2010). Decreased expression of *miR-497* has also been observed in peritoneal carcinoma (Flavin et al. 2009) and male breast cancer (Lehmann et al. 2010). Downregulation of *miR-195* was recently reported in adult ACC (Soon et al. 2009b), which is consistent with our results. The precursors of *miR-497* and *miR-195* are likely generated from the same miRNA cluster because of their close proximity in genomic location. A strong positive correlation between *miR-497* and *miR-195* expressions also indicates that these two miRNAs were co-expressed. The co-expression of the *miR-497–195* cluster suggests that these miRNAs could be co-regulated by common transcriptional factor(s) and they may have some common functions. Although the factor(s) regulating the transcription of this miRNA cluster are yet unknown, a common deletion within this chromosomal region (17p13) has been reported in ACC (Kjellman et al. 1996, Soon et al. 2008), but not in benign adrenocortical conditions (Almeida et al. 2011). This may, at least in

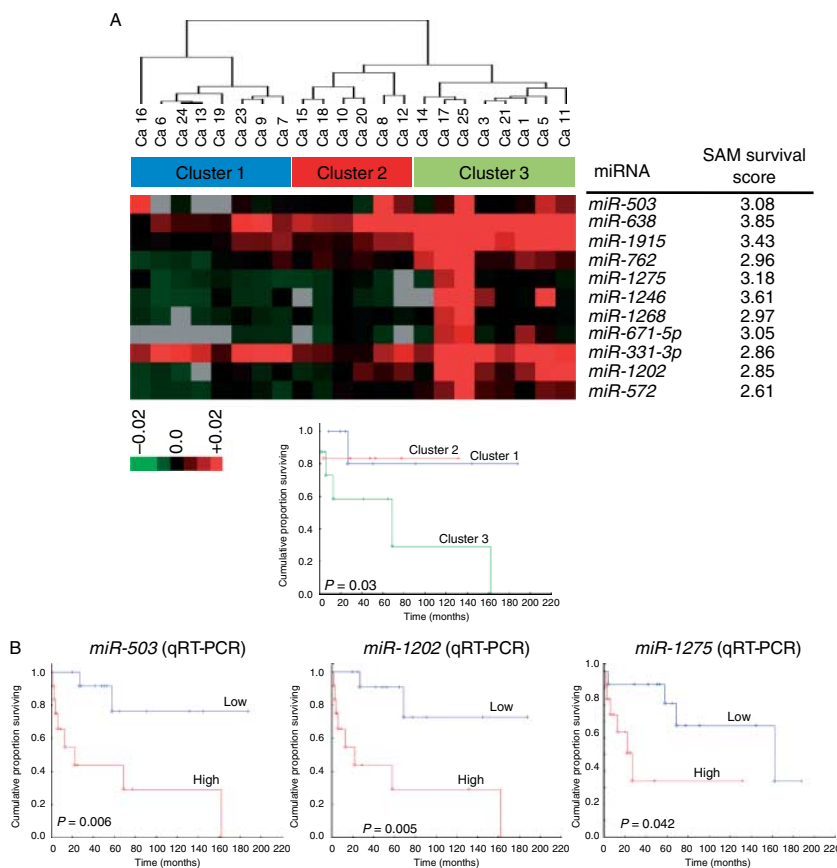


Figure 5 miRNAs associated with survival of carcinoma patients. (A) Clustering analysis of 11 miRNAs identified by SAM survival analysis of microarray data, and subsequent Kaplan–Meier analysis for cases in cluster 1–3. (B) Kaplan–Meier curves for *miR-503*, *miR-1202*, and *miR-1275* based on qRT-PCR results. Differences in survival were calculated using log-rank test.

part, explain the reduced expression of the *miR-497–195* cluster in ACC.

Despite the expression association, functional roles of miRNAs in ACC cells remain uncharacterized. In this study, we demonstrate the functional role of deregulated miRNAs in ACC cells using experimental cell culture systems. Overexpression of *miR-195* or *miR-497* in the NCI-H295R human ACC cell line could significantly reduce cell proliferation in combination with a pro-apoptotic effect. Consistent with our findings, *miR-497* was previously shown to promote neural death by negatively regulating expression of anti-apoptotic proteins, bcl-2 and bcl-w (Yin *et al.* 2010). Moreover, overexpression of *miR-195* could induce cell cycle arrest in hepatocellular carcinoma cells (Xu *et al.* 2009) and promote cell apoptosis in human colorectal cancer cells (Liu *et al.* 2010). Taken together, these findings revealed the potential tumor suppressive role and pro-apoptotic activity of this miRNA cluster in ACC and other cell types. Further investigation is certainly warranted to

identify their potential targets and elucidate their mechanisms of action in ACC development.

We also demonstrated that *miR-483-3p* and *miR-483-5p* expression levels were significantly higher in carcinomas compared with the benign or normal counterparts. Concordantly, overexpression of *miR-483-3p* and *miR-483-5p* were recently reported in childhood adrenocortical tumors (Doghman *et al.* 2010) and adult ACC (Patterson *et al.* 2011) respectively. However, Soon *et al.* (2009b) observed a non-significant trend for *miR-483* overexpression in adult ACC versus adenomas, where the lack of statistical significance could result from the relatively limited sample size. *miR-483-3p* and *miR-483-5p* are derived from different arms (3' and 5' arms respectively), of the same miRNA precursor. Although both *miR-483-3p* and *miR-483-5p* were significantly upregulated in carcinomas, we noted that the 3'-strand was much more abundant than the 5'-strand.

Overexpression of *miR-483-3p* may be a common alteration of tumor cells in several different tumor

types. In addition to ACC, overexpression of *miR-483-3p* has also been demonstrated in Wilms' tumors, and in colon, breast, and liver cancers (Veronese et al. 2010). Importantly, Veronese et al. (2010) recently demonstrated that silencing of *miR-483-3p* could suppress cell proliferation and induce apoptosis in the hepatocarcinoma cell line HepG2, as well as inhibit tumorigenicity *in vivo*. In line with their findings, we also observed a significant decrease in cell proliferation and increase in apoptosis by suppressing *miR-483-3p* expression in the human ACC cell line NCI-H295R. PUMA was identified as a target of *miR-483-3p* (Veronese et al. 2010). We also observed a significant inverse correlation between PUMA protein expression and *miR-483-3p* expression levels in ACC. The data support the expected inverse relationship between miRNA and target gene expressions.

miR-483-5p overexpression has been reported in tumors of the adrenal gland, such as ACC (Patterson et al. 2011) and pheochromocytoma (Meyer-Rochow et al. 2010). Although Ma et al. (2011) recently demonstrated that *miR-483-5p* directly targets *Socs3* in the mouse hepatoma Hepa 1–6 cells, its function remains uncharacterized. In this study, we show that *miR-483-5p* also promotes cell proliferation but has no effect on apoptosis in NCI-H295R ACC cells. The differential effect on apoptosis by *miR-483-3p* and *miR-483-5p* may be related to the pro-apoptotic PUMA. While PUMA is a target of *miR-483-3p*, computational prediction and expression correlation between *miR-483-5p* and PUMA expression (data not shown) suggest that PUMA is not a target of *miR-483-5p*. Taken together, *miR-483* is involved in various cancer types and *miR-483-3p* plays an important role in anti-apoptotic protection.

In addition, we also observed a significantly increased expression of *miR-21* and *miR-210* in ACC compared with adenomas and adrenal cortices. Overexpression of *miR-21* has been reported in several tumor types, such as glioblastoma (Chan et al. 2005), breast cancer (Iorio et al. 2005, Yan et al. 2008), chronic lymphocytic leukemia (Fulci et al. 2007), and cervical cancer (Lui et al. 2007). Several studies also showed that *miR-21* knockdown could impair cell growth, induce apoptosis and interfere with cell migration and invasion of cancer cells (Chan et al. 2005, Si et al. 2007, Asangani et al. 2008, Zhu et al. 2008). Although overexpression of *miR-21* has not been reported in previous miRNA profiling studies of ACC (Soon et al. 2009b, Tombol et al. 2009, Doghman et al. 2010, Patterson et al. 2011), its upregulation has been shown to promote cell proliferation in human NCI-H295R ACC cells (Romero et al. 2008). These

lines of evidence suggest that *miR-21* plays a common oncogenic role in several tumor types, including ACC.

Overexpression of *miR-210* appears to be a common feature in many tumor types (Camps et al. 2008, Huang et al. 2009, Zhang et al. 2009, Greither et al. 2010) and several reports have shown that its expression is regulated by hypoxia-inducible factor 1 α (HIF1 α). Hypoxia is frequently found in tumors and is associated with radiation-resistance and chemotherapy-resistance, increased metastatic potential and poor outcome (Pouyssegur et al. 2006, Lee et al. 2007). However, virtually no studies have so far been reported that evaluate the role of hypoxia in the pathogenesis of ACC. Further investigations are warranted to determine the involvement of hypoxia and the role of *miR-210* in ACC development or progression.

miRNAs associated with survival in ACC

Recently, Soon et al. (2009b) identified two miRNAs (i.e. *miR-483-5p* and *miR-195*) associated with survival in ACC using Exiqon miRCURY LNA microarray, however these miRNAs were not significantly associated with survival in our cohort using both Agilent microarray and qRT-PCR methods (data not shown). The discrepancy could be due to differences in sample characteristics, platforms, and data analysis methods.

In this study, we demonstrate that ACC can be divided into different subgroups with different clinical outcomes based on their miRNA expression profiles. Cluster 3 is associated with poorer outcome, in which most patients were died of the disease. The overall survival is apparently better in the other two clusters in which most patients are still alive. However, no distinct histologic features were found between the two subgroups (data not shown). Survival analysis revealed that high expression of *miR-503*, *miR-1202*, and *miR-1275* are significantly associated with poor survival of ACC patients. Notably, increased expression of *miR-503* has been observed in various human tumors, including parathyroid carcinoma (Corbetta et al. 2010), retinoblastoma (Zhao et al. 2009), and ACC (Soon et al. 2009b, Tombol et al. 2009). However, to our knowledge, the association between *miR-503* expression and survival has not been reported in any cancer type. Functionally, *miR-503* has been shown to directly target cell cycle regulators, which leads to induction of G1 cell cycle arrest in various cancer cell lines and promotion of cell differentiation in monocytes and myoblasts (Forrest et al. 2010, Sarkar et al. 2010). Although *miR-503* is important for promoting cell cycle arrest and differentiation, its role in tumor progression has yet to be

elucidated. The other two miRNAs demonstrating significant association with survival, *miR-1202* and *miR-1275*, have not been described in any cancer types. Nothing is known about their expression levels and function.

In conclusion, our findings show deregulation of a subset of miRNAs in ACC together with a potential role of *miR-483*, *miR-195*, and *miR-497* in the pathogenesis of this neoplasm. Our study also reveals that high expression of *miR-503*, *miR-1202*, and *miR-1275* are associated with poor survival of ACC patients, suggesting their potential prognostic value in ACC.

Supplementary data

This is linked to the online version of the paper at <http://dx.doi.org/10.1530/ERC-11-0082>.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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Author contribution statement

D M Ö, S C, C L, and W-O L conceived and designed the experiments; D M Ö, S C, D V-F, P A, and H X performed the experiments; D M Ö, S C, D V-F, P A, H X, and W-O L analyzed the data; M B, C L, A H, and J Z contributed to clinical and histopathological information of the cases; D M Ö, S C, C L, and W-O L wrote the manuscript; All authors have read and approved the final version of the manuscript.

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References

Almeida MQ, Harran M, Bimpaki EI, Hsiao HP, Horvath A, Cheadle C, Watkins T, Nesterova M & Stratakis CA 2011

- Integrated genomic analysis of nodular tissue in macronodular adrenocortical hyperplasia: progression of tumorigenesis in a disorder associated with multiple benign lesions. *Journal of Clinical Endocrinology and Metabolism* **96** E728–E738. (doi:10.1210/jc.2010-2420)
- Asangani IA, Rasheed SA, Nikolova DA, Leupold JH, Colburn NH, Post S & Allgayer H 2008 MicroRNA-21 (miR-21) post-transcriptionally downregulates tumor suppressor Pcd4 and stimulates invasion, intravasation and metastasis in colorectal cancer. *Oncogene* **27** 2128–2136. (doi:10.1038/sj.onc.1210856)
- Bertherat J, Groussin L & Bertagna X 2006 Mechanisms of disease: adrenocortical tumors – molecular advances and clinical perspectives. *Nature Clinical Practice. Endocrinology & Metabolism* **2** 632–641. (doi:10.1038/ncpendmet0321)
- Bimpaki EI, Iliopoulos D, Moraitis A & Stratakis CA 2010 MicroRNA signature in massive macronodular adrenocortical disease and implications for adrenocortical tumorigenesis. *Clinical Endocrinology* **72** 744–751. (doi:10.1111/j.1365-2265.2009.03725.x)
- Camps C, Buffa FM, Colella S, Moore J, Sotiriou C, Sheldon H, Harris AL, Gleadle JM & Ragoussis J 2008 hsa-miR-210 Is induced by hypoxia and is an independent prognostic factor in breast cancer. *Clinical Cancer Research* **14** 1340–1348. (doi:10.1158/1078-0432.CCR-07-1755)
- Caramuta S, Egyhazi S, Rodolfo M, Witten D, Hansson J, Larsson C & Lui WO 2010 MicroRNA expression profiles associated with mutational status and survival in malignant melanoma. *Journal of Investigative Dermatology* **130** 2062–2070. (doi:10.1038/jid.2010.63)
- Chan JA, Krichevsky AM & Kosik KS 2005 MicroRNA-21 is an antiapoptotic factor in human glioblastoma cells. *Cancer Research* **65** 6029–6033. (doi:10.1158/0008-5472.CAN-05-0137)
- Corbetta S, Vaira V, Guarnieri V, Scillitani A, Eller-Vainicher C, Ferrero S, Vicentini L, Chiodini I, Bisceglia M, Beck-Peccoz P *et al.* 2010 Differential expression of microRNAs in human parathyroid carcinomas compared with normal parathyroid tissue. *Endocrine-Related Cancer* **17** 135–146. (doi:10.1677/ERC-09-0134)
- DeLellis RA, Lloyd RV, Heitz PU & Eng C 2004 Pathology and genetics of tumours of endocrine organs. In *World Health Organization Classification of Tumours*. Lyon, France: IARC Press.
- Doghman M, El Wakil A, Cardinaud B, Thomas E, Wang J, Zhao W, Peralta-Del Valle MH, Figueiredo BC, Zambetti GP & Lalli E 2010 Regulation of insulin-like growth factor-mammalian target of rapamycin signaling by microRNA in childhood adrenocortical tumors. *Cancer Research* **70** 4666–4675. (doi:10.1158/0008-5472.CAN-09-3970)
- Ferracin M, Veronese A & Negrini M 2010 Micromarkers: miRNAs in cancer diagnosis and prognosis. *Expert Review of Molecular Diagnostics* **10** 297–308. (doi:10.1586/erm.10.11)

- Flavin RJ, Smyth PC, Laios A, O'Toole SA, Barrett C, Finn SP, Russell S, Ring M, Denning KM, Li J et al. 2009 Potentially important microRNA cluster on chromosome 17p13.1 in primary peritoneal carcinoma. *Modern Pathology* **22** 197–205. (doi:10.1038/modpathol.2008.135)
- Forrest AR, Kanamori-Katayama M, Tomaru Y, Lassmann T, Ninomiya N, Takahashi Y, de Hoon MJ, Kubosaki A, Kaiho A, Suzuki M et al. 2010 Induction of microRNAs, mir-155, mir-222, mir-424 and mir-503, promotes monocytic differentiation through combinatorial regulation. *Leukemia* **24** 460–466. (doi:10.1038/leu.2009.246)
- de Fraipont F, El Atifi M, Cherradi N, Le Moigne G, Defaye G, Houlgatte R, Bertherat J, Bertagna X, Plouin PF, Baudin E et al. 2005 Gene expression profiling of human adrenocortical tumors using complementary deoxyribonucleic acid microarrays identifies several candidate genes as markers of malignancy. *Journal of Clinical Endocrinology and Metabolism* **90** 1819–1829. (doi:10.1210/jc.2004-1075)
- Fulci V, Chiaretti S, Goldoni M, Azzalin G, Carucci N, Tavolaro S, Castellano L, Magrelli A, Citarella F, Messina M et al. 2007 Quantitative technologies establish a novel microRNA profile of chronic lymphocytic leukemia. *Blood* **109** 4944–4951. (doi:10.1182/blood-2006-12-062398)
- Giordano TJ, Thomas DG, Kuick R, Lizyness M, Misek DE, Smith AL, Sanders D, Aljundi RT, Gauger PG, Thompson NW et al. 2003 Distinct transcriptional profiles of adrenocortical tumors uncovered by DNA microarray analysis. *American Journal of Pathology* **162** 521–531. (doi:10.1016/S0002-9440(10)63846-1)
- Giordano TJ, Kuick R, Else T, Gauger PG, Vinco M, Bauersfeld J, Sanders D, Thomas DG, Doherty G & Hammer G 2009 Molecular classification and prognostication of adrenocortical tumors by transcriptome profiling. *Clinical Cancer Research* **15** 668–676. (doi:10.1158/1078-0432.CCR-08-1067)
- Greither T, Grochola LF, Udelnow A, Lautenschlager C, Wurl P & Taubert H 2010 Elevated expression of microRNAs 155, 203, 210 and 222 in pancreatic tumors is associated with poorer survival. *International Journal of Cancer* **126** 73–80. (doi:10.1002/ijc.24687)
- de Hoon MJ, Imoto S, Nolan J & Miyano S 2004 Open source clustering software. *Bioinformatics* **20** 1453–1454. (doi:10.1093/bioinformatics/bth078)
- Huang X, Ding L, Bennewith KL, Tong RT, Welford SM, Ang KK, Story M, Le QT & Giaccia AJ 2009 Hypoxia-inducible mir-210 regulates normoxic gene expression involved in tumor initiation. *Molecular Cell* **35** 856–867. (doi:10.1016/j.molcel.2009.09.006)
- Iliopoulos D, Bimpaki EI, Nesterova M & Stratakis CA 2009 MicroRNA signature of primary pigmented nodular adrenocortical disease: clinical correlations and regulation of Wnt signaling. *Cancer Research* **69** 3278–3282. (doi:10.1158/0008-5472.CAN-09-0155)
- Iorio MV & Croce CM 2009 MicroRNAs in cancer: small molecules with a huge impact. *Journal of Clinical Oncology* **27** 5848–5856. (doi:10.1200/JCO.2009.24.0317)
- Iorio MV, Ferracin M, Liu CG, Veronese A, Spizzo R, Sabbioni S, Magri E, Pedriali M, Fabbri M, Campiglio M et al. 2005 MicroRNA gene expression deregulation in human breast cancer. *Cancer Research* **65** 7065–7070. (doi:10.1158/0008-5472.CAN-05-1783)
- Kjellman M, Kallioniemi OP, Karhu R, Hoog A, Farnebo LO, Auer G, Larsson C & Backdahl M 1996 Genetic aberrations in adrenocortical tumors detected using comparative genomic hybridization correlate with tumor size and malignancy. *Cancer Research* **56** 4219–4223.
- Laurell C, Velazquez-Fernandez D, Lindsten K, Juhlin C, Enberg U, Geli J, Hoog A, Kjellman M, Lundeberg J, Hamberger B et al. 2009 Transcriptional profiling enables molecular classification of adrenocortical tumours. *European Journal of Endocrinology* **161** 141–152. (doi:10.1530/EJE-09-0068)
- Lee KA, Roth RA & LaPres JJ 2007 Hypoxia, drug therapy and toxicity. *Pharmacology & Therapeutics* **113** 229–246. (doi:10.1016/j.pharmthera.2006.08.001)
- Lehmann U, Streichert T, Otto B, Albat C, Hasemeier B, Christgen H, Schipper E, Hille U, Kreipe HH & Langer F 2010 Identification of differentially expressed microRNAs in human male breast cancer. *BMC Cancer* **10** 109. (doi:10.1186/1471-2407-10-109)
- Liu L, Chen L, Xu Y, Li R & Du X 2010 microRNA-195 promotes apoptosis and suppresses tumorigenicity of human colorectal cancer cells. *Biochemical and Biophysical Research Communications* **400** 236–240. (doi:10.1016/j.bbrc.2010.08.046)
- Lui WO, Pourmand N, Patterson BK & Fire A 2007 Patterns of known and novel small RNAs in human cervical cancer. *Cancer Research* **67** 6031–6043. (doi:10.1158/0008-5472.CAN-06-0561)
- Ma N, Wang X, Qiao Y, Li F, Hui Y, Zou C, Jin J, Lv G, Peng Y, Wang L et al. 2011 Coexpression of an intronic microRNA and its host gene reveals a potential role for miR-483-5p as an IGF2 partner. *Molecular and Cellular Endocrinology* **333** 96–101. (doi:10.1016/j.mce.2010.11.027)
- Meyer-Rochow GY, Jackson NE, Conaglen JV, Whittle DE, Kunnimalaiyaan M, Chen H, Westin G, Sandgren J, Stalberg P, Khanafshar E et al. 2010 MicroRNA profiling of benign and malignant pheochromocytomas identifies novel diagnostic and therapeutic targets. *Endocrine-Related Cancer* **17** 835–846. (doi:10.1677/ERC-10-0142)
- Patterson EE, Holloway AK, Weng J, Fojo T & Kebebew E 2011 MicroRNA profiling of adrenocortical tumors reveals miR-483 as a marker of malignancy. *Cancer* **117** 1630–1639. (doi:10.1002/cncr.25724)
- Pouyssegur J, Dayan F & Mazure NM 2006 Hypoxia signalling in cancer and approaches to enforce tumour regression. *Nature* **441** 437–443. (doi:10.1038/nature04871)

- de Reynies A, Assie G, Rickman DS, Tissier F, Groussin L, Rene-Corail F, Dousset B, Bertagna X, Clauser E & Bertherat J 2009 Gene expression profiling reveals a new classification of adrenocortical tumors and identifies molecular predictors of malignancy and survival. *Journal of Clinical Oncology* **27** 1108–1115. (doi:10.1200/JCO.2008.18.5678)
- Romero DG, Plonczynski MW, Carvajal CA, Gomez-Sanchez EP & Gomez-Sanchez CE 2008 Microribonucleic acid-21 increases aldosterone secretion and proliferation in H295R human adrenocortical cells. *Endocrinology* **149** 2477–2483. (doi:10.1210/en.2007-1686)
- Sarkar S, Dey BK & Dutta A 2010 MiR-322/424 and -503 are induced during muscle differentiation and promote cell cycle quiescence and differentiation by down-regulation of Cdc25A. *Molecular Biology of the Cell* **21** 2138–2149. (doi:10.1091/mbc.E10-01-0062)
- Si ML, Zhu S, Wu H, Lu Z, Wu F & Mo YY 2007 miR-21-mediated tumor growth. *Oncogene* **26** 2799–2803. (doi:10.1038/sj.onc.1210083)
- Soon PS, Libe R, Benn DE, Gill A, Shaw J, Sywak MS, Groussin L, Bertagna X, Gicquel C, Bertherat J *et al.* 2008 Loss of heterozygosity of 17p13, with possible involvement of ACADVL and ALOX15B, in the pathogenesis of adrenocortical tumors. *Annals of Surgery* **247** 157–164. (doi:10.1097/SLA.0b013e318153ff55)
- Soon PS, Gill AJ, Benn DE, Clarkson A, Robinson BG, McDonald KL & Sidhu SB 2009a Microarray gene expression and immunohistochemistry analyses of adrenocortical tumors identify IGF2 and Ki-67 as useful in differentiating carcinomas from adenomas. *Endocrine-Related Cancer* **16** 573–583. (doi:10.1677/ERC-08-0237)
- Soon PS, Tacon LJ, Gill AJ, Bambach CP, Sywak MS, Campbell PR, Yeh MW, Wong SG, Clifton-Bligh RJ, Robinson BG *et al.* 2009b miR-195 and miR-483-5p identified as predictors of poor prognosis in adrenocortical cancer. *Clinical Cancer Research* **15** 7684–7692. (doi:10.1158/1078-0432.CCR-09-1587)
- Tombol Z, Szabo PM, Molnar V, Wiener Z, Tolgyesi G, Horanyi J, Riesz P, Reismann P, Patocs A, Liko I *et al.* 2009 Integrative molecular bioinformatics study of human adrenocortical tumors: microRNA, tissue-specific target prediction, and pathway analysis. *Endocrine-Related Cancer* **16** 895–906. (doi:10.1677/ERC-09-0096)
- Velazquez-Fernandez D, Laurell C, Geli J, Hoog A, Odeberg J, Kjellman M, Lundeberg J, Hamberger B, Nilsson P & Backdahl M 2005 Expression profiling of adrenocortical neoplasms suggests a molecular signature of malignancy. *Surgery* **138** 1087–1094. (doi:10.1016/j.surg.2005.09.031)
- Veronese A, Lupini L, Consiglio J, Visone R, Ferracin M, Fornari F, Zanasi N, Alder H, D'Elia G, Gramantieri L *et al.* 2010 Oncogenic role of miR-483-3p at the IGF2/483 locus. *Cancer Research* **70** 3140–3149. (doi:10.1158/0008-5472.CAN-09-4456)
- Xu T, Zhu Y, Xiong Y, Ge YY, Yun JP & Zhuang SM 2009 MicroRNA-195 suppresses tumorigenicity and regulates G1/S transition of human hepatocellular carcinoma cells. *Hepatology* **50** 113–121. (doi:10.1002/hep.22919)
- Yan LX, Huang XF, Shao Q, Huang MY, Deng L, Wu QL, Zeng YX & Shao JY 2008 MicroRNA miR-21 over-expression in human breast cancer is associated with advanced clinical stage, lymph node metastasis and patient poor prognosis. *RNA* **14** 2348–2360. (doi:10.1261/rna.1034808)
- Yin KJ, Deng Z, Huang H, Hamblin M, Xie C, Zhang J & Chen YE 2010 miR-497 regulates neuronal death in mouse brain after transient focal cerebral ischemia. *Neurobiological Disorders* **38** 17–26. (doi:10.1016/j.nbd.2009.12.021)
- Zhang Z, Sun H, Dai H, Walsh RM, Imakura M, Schelter J, Burchard J, Dai X, Chang AN, Diaz RL *et al.* 2009 MicroRNA miR-210 modulates cellular response to hypoxia through the MYC antagonist MNT. *Cell Cycle* **8** 2756–2768. (doi:10.4161/cc.8.17.9387)
- Zhao JJ, Yang J, Lin J, Yao N, Zhu Y, Zheng J, Xu J, Cheng JQ, Lin JY & Ma X 2009 Identification of miRNAs associated with tumorigenesis of retinoblastoma by miRNA microarray analysis. *Child's Nervous System* **25** 13–20. (doi:10.1007/s00381-008-0701-x)
- Zhu S, Wu H, Wu F, Nie D, Sheng S & Mo YY 2008 MicroRNA-21 targets tumor suppressor genes in invasion and metastasis. *Cell Research* **18** 350–359. (doi:10.1038/cr.2008.24)

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